

Screening methods in alveolar echinococcosis: a follow-up study comparing Emc- and Emf-ELISA with Em2plus-ELISA and ultrasonography

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SUMMARY

The purpose of the present study was to identify *Echinococcus multilocularis* infection in follow-up of 95 subjects initially seropositive by Emc-ELISA or Emf-ELISA antibody assays and to compare the utility of these assays with specific Em2plus-ELISA and ultrasound screening for *E. multilocularis* infection. At follow-up seven subjects were seropositive with both methods, while three were seropositive only with Emc-ELISA and 11 only with Emf-ELISA. All subjects were seronegative with Em2plus-ELISA. There were no manifestations of *E. multilocularis* infestation by ultrasonographic screening. Seropositivity on Emc-ELISA and Emf-ELISA screening tests does not appear to correlate with manifest alveolar echinococcosis identified by ultrasound. A recommendation for further follow-up of subjects found to be seropositive with Emc-ELISA and Emf-ELISA but with no sonographic evidence of disease is not justified at this time.

INTRODUCTION

Alveolar echinococcosis (AE) is a zoonosis caused by the larval stage of the fox tapeworm, *Echinococcus multilocularis*. Humans serve as an intermediate host for the cestode and actually are an inappropriate or 'dead-end' host [1, 2]. Almost without exception, the primary site of disease manifestation in humans is the liver [1, 3]. More than 10 years may pass between first infection and clinical manifestation [1]. Untreated, this disease remains fatal [4, 5]. The incidence of the

disease varies in the endemic regions of Central Europe between 0·02 and 1·4/100 000 persons [6, 7]. At present, only three studies have assessed prevalence using both serology and ultrasound; the remaining prevalence data are based solely on serological studies [8–10].

Earlier prevalence studies have used Em2-, EgHF-, EmP-, Emc- and Emf-ELISAs for screening larger populations [9–13]. Currently, reports in the literature describe a total of 14 different enzyme-linked immunosorbent assays (ELISAs) (Emc-, Emf-, Em2-, Em2plus-, EgHF-, EmP-, Em10-, Em13-, Em16-, Em18-, Em70-, Em90-, EmAP- and EmII/3-10-ELISA) that have been used to diagnose AE [9–19].

Previous data on the change in *E. multilocularis* antibody concentrations in AE over time are available

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only from a rural population in southern Germany [10, 20]. In this population, follow-up revealed no increase in disease progression or significant change in serological antibody concentrations. Data for an urban population have not yet been published. In November, 2002, a cross-sectional survey drawn from an urban population of Leutkirch, a town in south-western Germany, using crude Emc- and Emf-ELISAs demonstrated that 95 individuals, or 3.9% of those tested, were seropositive to at least one of the two crude antigen ELISAs used [21]. No clinical or ultrasonographic signs of AE were found in any of these subjects.

The objective of the present study was to re-examine these seropositive subjects and to assess the reliability of Emc- and Emf-ELISAs in predicting AE compared with the specific Em2plus-ELISA and diagnostic ultrasound screening.

METHODS

Follow-up study population

At the time of the initial survey in 2002, a total of 95 study participants tested positive with at least one of two raw antigen ELISAs, with 16 probands positive for Emc and 79 for Emf. Emc antigen was prepared from *E. multilocularis* metacystode tissue grown intraperitoneally in a laboratory strain of common voles as described previously by Gottstein *et al.* [22]. Emf antigen was vesicular fluid of *E. multilocularis* metacystodes grown *in vitro* as previously described by Hemphill & Gottstein [23]. All seropositive subjects identified during the initial EMIL study received a written invitation 10 months after completion of the screening to attend a follow-up appointment to include blood sampling and ultrasound examination. The follow-up examinations were conducted between November 2003 and February 2004 at the University Hospital Ulm Medical Centre.

Subjects who had not responded to the written invitation within 2 months were contacted by phone and again invited to a follow-up appointment. The most frequent reason stated for declining the follow-up appointment was the long distance to Ulm. Logistical and financial resources did not allow re-examination of the study participants at the original study site. Despite written invitation and telephone reminders, only 44 of the initial 95 seropositive subjects (46.3%) agreed to participate in the follow-up study appointment. Of these 44 subjects, 25 were

female (average age 40.6 years, range 13–65 years) and 19 were male (average age 37.3 years, range 13–65 years). Only 31 subjects underwent both blood sampling and ultrasound examination. Twelve subjects underwent only ultrasound examination, which, in four cases, was performed at the University Hospital Ulm, and in the remaining eight cases at the practices of their respective primary-care physicians.

The study was conducted in accordance with the principles of the Helsinki Declaration and the GCP (Good Clinical Practice) recommendations. It was approved by the ethics commission of the Landesärztekammer Baden-Württemberg [21].

Serological testing

Initial investigations in 2002

In the initial survey, 10 ml of whole blood was obtained from each subject for serological testing (Emc- and Emf-ELISA). After sedimentation at 7000–8000 g for 10 min, samples were stored at -70°C until final processing. Samples were tested for antibodies to *E. multilocularis* using an Emf-ELISA and Emc-ELISA (metacystode production *in vivo* in *Meriones unguiculatus* or metacystode *in vitro* culture according to Hemphill & Gottstein [23]). Antigens were coated onto microtitre plates (Nunc MaxiSorp, Nunc GmbH & Co., Wiesbaden, Germany) with a protein concentration of 3 mg/ml of Emf and Emc antigen. The participants' sera were investigated by ELISA at a dilution of 1:200. Anti-human IgG horseradish peroxidase was applied as conjugate at a standard dilution of 1:10 000 as indicated by the manufacturer (Dako, Deutschland GmbH, Hamburg, Germany). Tetramethylbenzidine dihydrochloride (TMB) served as substrate, and absorbance values were read at 450 nm.

Serological findings were considered as positive if the index reached a level of mean optical density plus 3 standard deviations, corresponding to >72 for Emc-ELISA and >110.3 for Emf-ELISA. All other findings were considered negative. All samples were tested in the laboratory of the State Health Department, Stuttgart, Germany.

Follow-up in 2004

As part of the follow-up examination conducted in 2003–2004, 10 ml of whole blood were obtained from 31 subjects for serological testing (Emc- and Emf-ELISA). Blood samples were processed and stored as described above.

Repeat serological testing in 2006

In 2006, in order to exclude methodologically based changes in concentrations and to enhance comparability of test results, we re-tested all sera obtained at the initial survey in 2002 and at the follow-up examinations in 2003–2004. For Emc- and Emf-ELISA, the methods used were identical to those used in 2002. All blood samples were tested in the laboratory of the State Health Department, Stuttgart, Germany.

In addition, all samples from the initial survey in 2002 and from the follow-up examinations in 2003–2004 were tested using Em2plus-ELISA (DPC Biermann GmbH, Bad Nauheim, Germany). The sensitivity of this commercially available test is reported as 97% [14]. The *E. multilocularis* Em2plus-ELISA is a solid-phase ELISA using microtitre plates for detecting serum IgG antibodies to *E. multilocularis*. The microtitre strips included with the test kit are coated with *E. multilocularis* Em2plus antigen. This is a mixed antigen comprised of an affinity-purified Em2 antigen and a recombinant II/3–10 antigen from *E. multilocularis*. The absorbance values obtained by the Em2plus-ELISA were measured at 405 nm. The Em2plus-ELISA testing was conducted in the Department of Medical Microbiology and Hygiene of the University Hospital Ulm.

Ultrasound examination

Ultrasound follow-up examinations were performed at the University Hospital Ulm using a HDI 5000 ultrasound scanner (ATL Ultrasound, Philips Medical Systems, Bothell, WA, USA) of the same type as used at the initial survey in Leutkirch in 2002 [21]. Ultrasound follow-up examinations outside the University Hospital were done by primary-care physicians. All of the physicians had a licence to perform ultrasound examinations from the Medical Council. On examination, the liver was assessed for evidence of infection with *E. multilocularis*; the examination also included assessment of the kidneys, gallbladder, biliary tract and spleen.

RESULTS

In the initial survey of the EMIL study in 2002, 95/2445 subjects from an urban population in southwest Germany were found to be positive on at least one of two raw antigen ELISAs (Emc-ELISA, $n=16$; Emf-ELISA, $n=79$) for evidence of infection with *E. multilocularis*. The following data are results of

the re-testing in 2006, conducted under identical laboratory conditions, of cryopreserved sera of 31 subjects who participated in the initial (2002) and follow-up examinations (2003–2004).

Initial sera (2002)

Of the 31 sera found to be positive (Emc and Emf) at the initial survey in 2002, only 26 tested positive at re-testing conducted in 2006. Of these, eight tested positive with Emc-ELISA and 13 with Emf-ELISA, while three remained positive with both tests. Five initially positive sera tested negative at re-testing despite application of identical testing procedures. All sera initially positive in 2002 tested negative by Em2plus-ELISA (see Table 1).

Follow-up sera (2004)

Of the 31 sera obtained at the follow-up examinations in 2004, only 21 were positive when tested again in 2006. Seven of the 21 sera were positive by both Emc- and Emf-ELISA, while three sera were positive only by Emc-ELISA and 11 only by Emf-ELISA. All 21 sera testing positive in the follow-up examinations in 2004 tested negative with the Em2plus-ELISA (see Table 1).

Comparison of the 2002 and 2004 sera

A comparison of Emc-ELISA concentration results obtained in 2002 and 2004 showed that the number of subjects seropositive by this assay increased from eight to ten. Among the total of 14 subjects positive by Emc-ELISA in 2002 or 2004, follow-up testing in 2004 revealed agreement with the 2002 results in only four cases. A comparison of Emf-ELISA concentration results obtained in 2002 and 2004 showed that the number of subjects seropositive by this assay increased from 13 to 18. Among the total of 22 subjects positive by Emf-ELISA in 2002 or 2004, follow-up testing in 2004 revealed agreement with the 2002 results in nine cases (Table 1).

Ultrasound

Study subjects undergoing follow-up ultrasound examinations at the University Hospital Ulm or by their respective primary-care physician did not identify any findings specific for *E. multilocularis* disease. In 30 subjects, the findings of the diagnostic ultrasound were within normal limits and appropriate for their

Table 1. Results of Emc- and Emf-ELISA in 2002 and at follow-up in 2004

| Subject no. | Emc-ELISA | | Emf-ELISA | | Em2plus | |
|-------------|-----------|-------|-----------|-------|---------|------|
| | 2002 | 2004 | 2002 | 2004 | 2002 | 2004 |
| 1 | + | - | + | - | - | - |
| 2 | + | - | + | - | - | - |
| 3 | + | - | + | - | - | - |
| 4 | + | - | - | - | - | - |
| 5 | + | + | - | + | - | - |
| 6 | + | + | - | - | - | - |
| 7 | + | + | - | - | - | - |
| 8 | + | + | - | - | - | - |
| 9 | - | + | + | + | - | - |
| 10 | - | + | + | + | - | - |
| 11 | - | + | + | + | - | - |
| 12 | - | + | - | + | - | - |
| 13 | - | + | - | + | - | - |
| 14 | - | + | - | + | - | - |
| 15 | - | - | + | + | - | - |
| 16 | - | - | + | + | - | - |
| 17 | - | - | + | + | - | - |
| 18 | - | - | + | + | - | - |
| 19 | - | - | + | + | - | - |
| 20 | - | - | + | + | - | - |
| 21 | - | - | + | - | - | - |
| 22 | - | - | - | + | - | - |
| 23 | - | - | - | + | - | - |
| 24 | - | - | - | + | - | - |
| 25 | - | - | - | + | - | - |
| 29 | - | - | - | + | - | - |
| 27 | - | - | - | - | - | - |
| 28 | - | - | - | - | - | - |
| 26 | - | - | - | - | - | - |
| 30 | - | - | - | - | - | - |
| 31 | - | - | - | - | - | - |
| Sum | 8/23 | 10/21 | 13/18 | 18/13 | 0/31 | 0/31 |
| | pos./neg. | | | | | |

Emc-ELISA, Antigen obtained from metacystodes; Emf-ELISA, antigen obtained from vesicular tissue; +, positive; -, negative.

Grey shading indicates sera which have been positive for Emc-ELISA and Emf-ELISA in 2002 and 2004.

respective ages. Coincidental findings included hepatic steatosis in five cases, liver cysts in two cases, a haemangioma in one case, gallbladder stones in two cases, and gallbladder polyps in four cases. None of the findings at follow-up ultrasound had immediate diagnostic or therapeutic relevance.

DISCUSSION

Few studies have used serology and diagnostic ultrasound to assess the prevalence or seroprevalence of

E. multilocularis in follow-up [10, 20]. It is still unclear to what extent seropositive individuals may be at risk of subsequently developing manifest AE. Because seroprevalence rates up to 20% can be found in highly endemic areas [9, 10], understanding the relationship between seropositivity without detectable lesions on ultrasound examination and future disease is of great importance in developing follow-up strategies to reduce morbidity and mortality associated with AE.

The findings of the present study show significant changes in Emc- and Emf-ELISA antibody concentrations over time using these non-specific assays. The results from antibody concentration determinations performed in 2006 did not reproduce the initial concentrations from 2002 despite using identical test procedures. Agreement between the two test methods was observed in only three subjects in 2002 and in only seven subjects in 2004. For Emc-ELISA, comparison of 2002 and 2004 results revealed agreement in only four subjects, while, for Emf-ELISA, there was agreement in only nine cases. The inconsistency between 2002 and 2004 measurement results must, therefore, be seen as a basic method-related problem with this screening test and underscores the poor reliability of these methods. The highly specific Em2plus ELISA was negative for all sera from both the 2002 and 2004 series, consistent with both the results of the ultrasound examination and the reported high specificity of this test [14].

A study conducted in 1996 in the southwestern German town of Römerstein [10] used the same serological tests as were used in our study. Three years after completion of the initial Römerstein study, 36/47 (76.6%) seropositive patients participated in a follow-up study [20]. At follow-up, ultrasound examination did not demonstrate findings suggestive of *E. multilocularis* lesions in any study subject.

The studies described above of patients with persistently elevated antibody concentrations to *E. multilocularis* antigens without evidence of typical liver lesions on ultrasound may represent latent AE, which, with its long incubation period of 10–15 years [1] has not yet become manifest. An alternate explanation for persistently positive serologies in individuals without evidence of active disease may be immunity among persons residing in areas highly endemic for *E. multilocularis* [24]. According to Gottstein and colleagues [7, 25], only about 10–30% of persons undergoing seroconversion following

exposure to *E. multilocularis* subsequently develop AE [25]. In addition, there may be individuals who, despite a high risk of exposure to *E. multilocularis*, fail to mount an immunological response to parasitic antigens. Finally, *E. multilocularis* seropositivity in clinically healthy subjects might be due to cross-reactivity of the raw antigen ELISAs, as has been described previously [14].

Vuitton describes four possible situations in which an individual may develop antibodies to *E. multilocularis*. First, a person may already exhibit clinically manifest AE; second, the person may suffer from latent disease that has not yet become clinically apparent; third, there may be extinct and calcified lesions in the liver; fourth, there may be no detectable lesions whatsoever [26]. A study conducted in Alaska reported cases of AE in which the parasites have spontaneously died out: in these cases, Em2-ELISA remained persistently positive [27].

There are also reports in the literature of protective factors against infection with *E. multilocularis*. The HLA allele HLA-DRB1*11 has been associated with protection against development of AE [28]. In the same study, other HLA alleles such as HLA-DQB1*02 appeared to be associated with a progressive disease course. In another study [29], it was shown that patients with the HLA DR3 + DQ2 + haplotype secrete higher levels of TH2 cytokine, which promotes disease progression.

The seropositive subjects in the present study using unspecific ELISAs such as Emf and Emc seem not to suffer from the early stages of AE. Seropositivity to these antigens may simply be a cross-reaction [30]. The results of our study do not allow us to postulate subjects' possible contact with *E. multilocularis* antibody or to assume a certain degree of immunity to the parasite, especially since the more specific Em2plus-ELISA and ultrasound remained negative at both the initial and follow-up surveys.

A limitation of the study might be the long period of sample storage. To our knowledge there exists no study, which systematically investigated the stability of antibody concentrations in blood sera over years. However the experience of the Baden-Württemberg State Health Office suggests a high quality of samples stored at -70°C over many years.

Follow-up of subjects seropositive by Emc- or Emf-ELISA screening surveys and without sonographic evidence of disease does not appear to be justified at this time.

APPENDIX. Members of the EMIL Study Group

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DECLARATION OF INTEREST

None.

REFERENCES

1. **Ammann RW, Eckert J.** Cestodes. *Echinococcus*. *Gastroenterology Clinics of North America* 1996; **25**: 655–689.
2. **Gottstein B.** Molecular and immunological diagnosis of echinococcosis. *Clinical Microbiology Reviews* 1992; **5**: 248–261.
3. **Reuter S, et al.** Extrahepatic alveolar echinococcosis without liver involvement: a rare manifestation. *Infection* 2000; **28**: 187–192.
4. **Kern P, Kratzer W, Reuter S.** Alveolar echinococcosis: diagnosis. *Deutsche Medizinische Wochenschrift* 2000; **125**: 59–62.
5. **Kern P, Kratzer W, Reuter S.** Alveolar echinococcosis: therapy. *Deutsche Medizinische Wochenschrift* 2000; **125**: 87–89.
6. **Eckert J, Deplazes P.** Alveolar echinococcosis in humans: the current situation in central Europe and

- the need of countermeasures. *Parasitology Today* 1999; **15**: 315–319.
7. **Gottstein B.** Epidemiology and systematics of cystic and alveolar hydatid disease. *Chirurg* 2000; **71**: 1–8.
 8. **Bartholomot B, et al.** Combined ultrasound and serological screening for hepatic alveolar echinococcosis in central China. *American Journal of Tropical Medicine and Hygiene* 2002; **66**: 23–29.
 9. **Craig PS, et al.** A large focus of alveolar echinococcosis in central China. *Lancet* 1992; **340**: 826–831.
 10. **Romig T, et al.** An epidemiological survey of alveolar echinococcosis in southwestern Germany. *American Journal of Tropical Medicine and Hygiene* 1999; **61**: 566–573.
 11. **Bresson-Hadni S, et al.** Seroepidemiological screening of *Echinococcus multilocularis* infection in a european area endemic for alveolar echinococcosis. *American Journal of Tropical Medicine and Hygiene* 1994; **51**: 837–846.
 12. **Gottstein B, et al.** Seroepidemiological survey for alveolar echinococcosis (by Em2-ELISA) of blood donors in an endemic area of Switzerland. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 1987; **81**: 960–964.
 13. **Gottstein B, et al.** Is high prevalence of *Echinococcus multilocularis* in wild and domestic animals associated with disease incidence in humans? *Emerging Infectious Diseases* 2001; **7**: 408–412.
 14. **Gottstein B, et al.** Primary immundiagnosis of alveolar echinococcosis in humans by an enzyme-linked immunosorbent assay using the Em2plus-antigen. *Journal of Clinical Microbiology* 1993; **31**: 373–376.
 15. **Gottstein B, et al.** *Echinococcus granulosus* antigens: immun-electrophoretic and western blot analysis of hydatid cyst fluids. *Parasitology Research* 1987; **73**: 186–189.
 16. **Korkmaz M, et al.** Use of two sensitive and specific immunoblot markers, Em70 and Em90, for diagnosis of alveolar echinococcosis. *Journal of Clinical Microbiology* 2004; **43**: 3350–3352.
 17. **Siles-Lucas MM, Gottstein BB.** Molecular tools for the diagnosis of cystic and alveolar echinococcosis. *Tropical Medicine & International Health* 2001 **6**: 463–475.
 18. **Sarciron S, et al.** Antibodies against *Echinococcus multilocularis* alkaline phosphatase as markers for the specific diagnosis and the serological monitoring of alveolar echinococcosis. *Parasite Immunology* 1997; **19**: 61–68.
 19. **Zhang W, Li J, McLanus DP.** Concepts in immunology and diagnosis of hydatid disease. *Clinical Microbiology Reviews* 2003; **16**: 18–36.
 20. **Jensen B, et al.** Long-term seropositivity against *Echinococcus multilocularis* in an epidemiological follow-up study in southwestern Germany (Römerstein). *Infection* 2001; **29**: 310–314.
 21. **Haenle MM, et al.** Overweight, physical activity, tobacco and alcohol consumption in a cross-sectional random sample of German adults. *BMC Public Health* 2006; **6**: 233.
 22. **Gottstein B, et al.** Serological differentiation between *Echinococcus granulosus* and *E. multilocularis* in man. *Parasitology Research* 1983; **69**: 347–356.
 23. **Hemphill A, Gottstein B.** Immunology and morphology studies on the proliferation of in vitro cultivated *Echinococcus multilocularis* metacestodes. *Parasitology Research* 1995; **81**: 605–614.
 24. **Lanier AP, et al.** Comparison of serological tests for the diagnosis and follow-up of alveolar hydatid disease. *American Journal of Tropical Medicine and Hygiene* 1987; **37**: 609–615.
 25. **Gottstein B, Felleisen R.** Protective immune mechanisms against the metacestode of *Echinococcus multilocularis*. *Parasitology Today* 1995; **11**: 320–326.
 26. **Vuitton DA.** Echinococcosis and allergy. *Clinical Reviews in Allergy and Immunology* 2004; **26**: 93–104.
 27. **Stehr-Green JK, et al.** Risk factors for infection with *Echinococcus multilocularis* in Alaska. *American Journal of Tropical Medicine and Hygiene* 1988; **38/2**: 380–385.
 28. **Eiermann TH, et al.** HLA and alveolar echinococcosis. *Tissue Antigens* 1998; **52**: 124–129.
 29. **Godot V, et al.** Resistance/susceptibility to *Echinococcus multilocularis* infection and cytokine profile in humans. II. Influence of the HLA B8, DR3, DQ2 haplotype. *Clinical & Experimental Immunology* 2000; **121**: 491–498.
 30. **Muller N, et al.** Improved serodiagnosis of alveolar echinococcosis of humans using an in vitro-produced *Echinococcus multilocularis* antigen. *Parasitology* 2007; **134**: 879–888.