

Impact of the use of an alcohol-based hand sanitizer in the home on reduction in probability of infection by respiratory and enteric viruses

A. H. TAMIMI¹, S. MAXWELL¹, S. L. EDMONDS^{2*} AND C. P. GERBA¹

¹ *Department of Soil, Water and Environmental Science, University of Arizona, Tucson, AZ, USA*

² *Research and Development, GOJO Industries, Akron, OH, USA*

*Received 21 April 2014; Final revision 8 December 2014; Accepted 7 January 2015;
first published online 31 March 2015*

SUMMARY

The goal of this study was to determine the reduction in risk of infection by viruses with the use of an alcohol-based hand sanitizer, used in addition to routine hand washing, in family members in households. A quantitative microbial risk model was used to determine the probability of infection from the concentration of virus on the hands. The model incorporated variation in hand size, frequency of touching orifices (nose, mouth, eyes), and percent transfer to the site of infection, as well as, dose-response for each virus. Data on the occurrence of virus on household members' hands from an intervention study using MS-2 coliphage was used to determine the reduction of viruses on the hands pre- and post-intervention. It was found that the risk of rhinovirus, rotavirus or norovirus infection after the intervention was reduced by 47–98% depending upon the initial concentration of virus on the hands.

Key words: Hand hygiene, risk assessment, virus infection.

INTRODUCTION

Every year more than 800 million cases of respiratory and gastrointestinal infections occur in the United States [1]. Viruses are a major cause of these illnesses. Rhinovirus is the leading cause of the common cold. Rotavirus is a leading cause of gastroenteritis in children, while noroviruses are usually associated with gastroenteritis in adults. These viruses are known to be readily transmitted by contact with contaminated surfaces and self-inoculation by hand contact with the nose, mouth or eyes [2, 3]. Hand hygiene involving hand washing or the use of hand sanitizers is believed to play a significant role in reducing infection risks of respiratory and enteric infections [4]. We recently conducted a study on the impact of the spread of viruses

from a contaminated hand to fomites and to other family members' hands in the household [5]. In that study it was found that use of a hand sanitizer in addition to routine hand washing resulted in the reduction of a tracer virus by 97–99·9% on fomites and the hands of household members when the sanitizer was used three times over an 8-h period. The results on the occurrence of virus on family members' hands from that study were used in a quantitative microbial risk assessment model to determine the possible reduction in probability of infection from respiratory and enteric viruses with the use of the hand sanitizer.

METHODS

The objective of this study was to determine the risk of infection from viruses using an alcohol-based hand sanitizer (Purell Advanced Hand Sanitizer, GOJO Industries, USA) three times over an 8-h period. An

* Author for correspondence: Ms. S. L. Edmonds, GOJO Industries Inc., PO Box 991, Akron, OH 44309, USA.
(Email: edmondss@gojo.com)

alcohol gel hand sanitizer known to be effective against rhinoviruses, rotavirus, norovirus and coliphage MS-2 was used in this study [6–8]. The viruses selected for modelling were rhinoviruses, rotavirus and norovirus. Because there is uncertainty in using the available dose-response data for norovirus the dose-response for rotavirus developed in adults was used for modelling both viruses. The dose-response data available for norovirus was developed using a non-infectivity assay for the virus and there were questions by the authors about the presence of aggregates in the preparation given to the human volunteers [8].

Pre-intervention consisted of the family members' routine hand washing with soap and water. Post-intervention involved both routine hand washing with soap and water, and the use of a hand sanitizer at least three times during 8 h. Total concentration of MS-2 coliphage inoculated to the hands of the subjects was 1×10^8 plaque-forming units (p.f.u.) to ensure detection during laboratory analysis.

Dose-response modelling

For rhinovirus and rotavirus dose-response models, datasets from human exposure experiments conducted by Hendley *et al.* and Ward *et al.* were used [3, 9]. For each of the viruses, different statistical distribution models were assessed to determine best fit using maximum likelihood estimation techniques. It was determined that the beta-Poisson model best fits the dose-response data-sets for both rhinovirus and rotavirus. The general form of the beta-Poisson distribution is given by equation (1), in which $P(\text{response})$ is the probability of response or infection, a and b are the optimization coefficients that follow a beta distribution, and D is the concentration or dose of the pathogen under investigation [10].

$$P(\text{response}) = 1 - \left(1 + \frac{D}{\beta}\right)^{-\alpha} \quad (1)$$

The Solver routine in Microsoft Excel program was used in this study to optimize the beta-Poisson model by varying a and b coefficients in equation (1) with the objective being maximization of the χ^2 goodness of fit between the observed and the expected probability of risk of infection.

Exposure model

By knowing the count of viruses on the hand post- and pre-intervention, the amount of virus ingested or

coming into contact with the eye or nose can be calculated from equation (2).

$$D = \frac{C_{\text{hand}}}{A_{\text{hand}}} \times \sum_{i=1}^m (f_{2,i} \times A_i \times N_i \times T_i), \quad (2)$$

where D is the dose or total count of viable (infectious) virus that is ingested or exposed to the mouth, eye or nose by a person in time T ; C_{hand} is the concentration of viable (infectious) virus measured on hand per cm^2 ; A_{hand} is the area of two hands (cm^2); $f_{2,i}$ is hand-to-orifice ' i ' transfer efficiency of virus (fraction; dimensionless); i is orifice i through which viruses can be ingested such as mouth, nose or eye (1, 2, 3, 4, ..., m); m is the total number of orifices; A_i is the surface area of the hand that touches orifice ' i ' (cm^2); N_i is the number of times a person touches his/her orifice ' i ' (per minute); T_i is the time duration of exposure (minutes).

Parameters for exposure model

To determine hand-to-nose, hand-to-eye and hand-to-mouth contacts per minute, data from Nicas & Best were utilized to generate the distributions shown in Figure 1 (*a-c*, respectively) [11]. Hand-to-nose contact distribution has an average of 0.0848/min [95% confidence interval (CI) 0.0546–0.1148]; hand-to-eye contact distribution has an average of 0.0411/min (95% CI 0.0247–0.0586); and hand-to-mouth contact distribution has an average of 0.1332/min (95% CI 0.0694–0.2283).

To determine the area of hand, nose, eye and mouth, data from Snyder *et al.* were utilized to generate the distributions shown in Figure 2 [10]. Data obtained from U.S. EPA and from Snyder *et al.* were used to determine the distribution of area of hands (A_{hand}) [10, 12, 13]. Bootstrapping was used to determine the distribution of areas of hands shown in Figure 2*a*. The value t^* represents hand area (cm^2) and the y-axis represent the probability density function of the distribution [13–17]. The area of hand distribution is not a normal distribution with an average area of 658 cm^2 (95% CI 533.1–881.6). The area of nose has an average of 5.12 cm^2 (95% CI 4.608–5.654); area of eye has an average of 1.56 cm^2 (95% CI 1.523–1.602); and area of mouth has an average of 6.89 cm^2 (95% CI 6.63–7.13).

A point estimate of 0.339 for transfer efficiency of virus from hand to nose, hand to eye and hand to mouth was used in the model based on Rusin *et al.* since they used MS-2 to develop the data [18]. This is the only data available in the literature. Parameters derived from the bootstrapping are shown in Table 1.

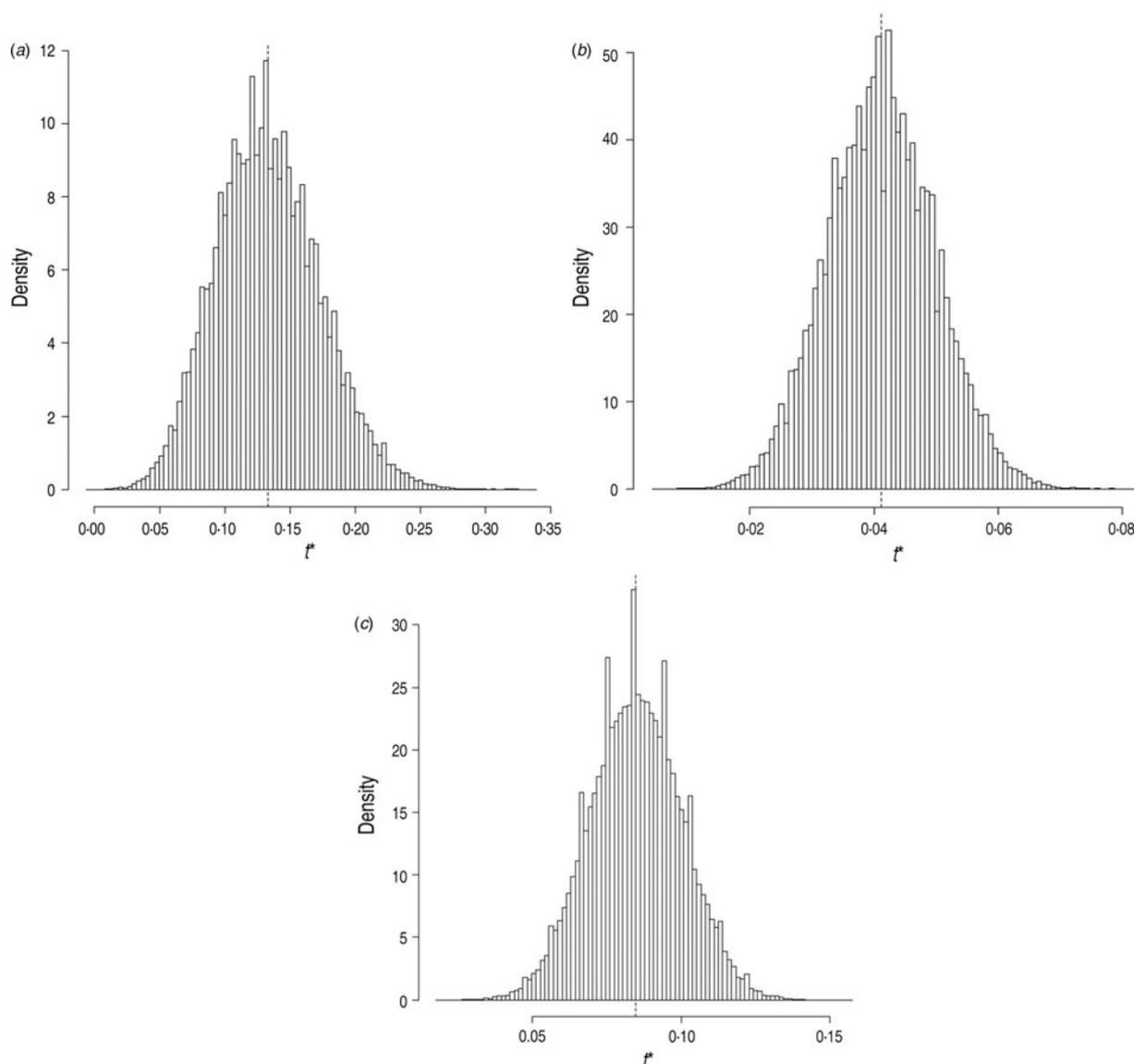


Fig. 1. Generated distributions for face contact frequency using bootstrapping. (a) Hand-to-mouth contacts/min; (b) hand-to-eye contacts/min; (c) hand-to-nose contacts/min.

RESULTS

Rhinovirus probability of infection

Rhinovirus is transmitted by contact with the nose and the eyes; hence plugging the parameters presented in Table 1 into equation (2) yields equation (3) shown below, in which C_{hand} is the concentration of rhinovirus found on the subjects’ hands.

$$D = \frac{C_{hand} \text{ (number of rhinoviruses)}}{658 \cdot 94 \text{ cm}^2} \times \left(\begin{matrix} 0 \cdot 39 \times 5 \cdot 12 \text{ cm}^2 \times \frac{0 \cdot 0848}{\text{minute}} \times 480 \text{ min} + \\ 0 \cdot 39 \times 1 \cdot 56 \text{ cm}^2 \times \frac{0 \cdot 0411}{\text{minute}} \times 480 \text{ min} \end{matrix} \right). \quad (3)$$

Table 2 shows different scenarios of concentrations of rhinovirus on the hands and the corresponding probability of infection before and after the use of the hand sanitizer three times during an 8-h period. Table 2 also gives the calculated percent risk reduction of infection due to the use of a hand sanitizer. Because of the nature of the dose-response curve for rhinovirus, greater reduction in risk of infection is seen with a lower initial concentration of virus on the hands.

Figure 3 shows rhinovirus risk of infection pre- and post-intervention through the use of a hand sanitizer three times during 8 h assuming an initial concentration of 1000 infectious rhinoviruses on the hands (from Table 2) for each individual in the studied households.

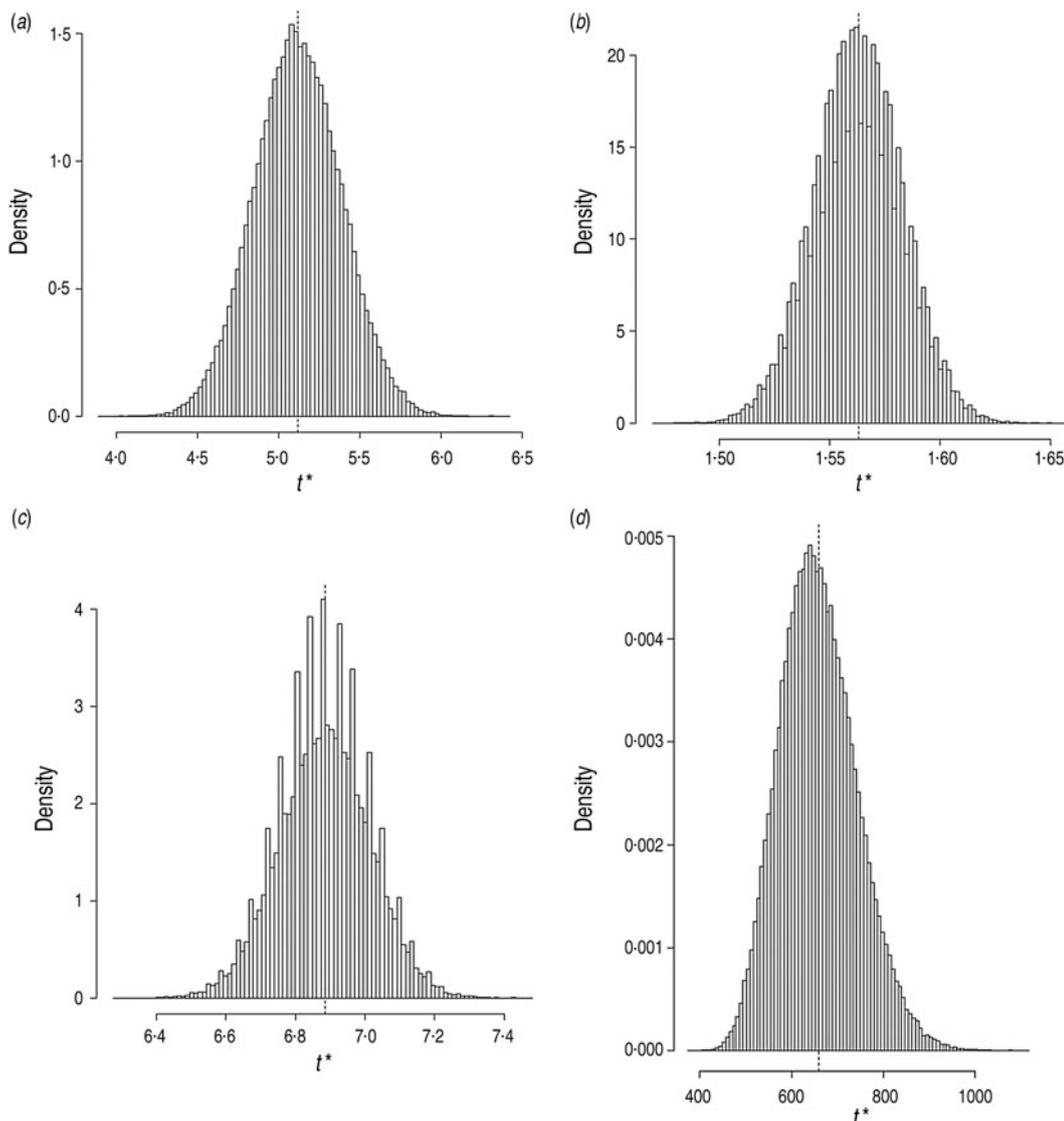


Fig. 2. Generated distributions for the area (cm²) of the (a) nose; (b) eye; (c) mouth; (d) hand, using bootstrapping techniques.

The risk reduction of infection varies with each individual depending upon the initial average reduction of the surrogate on the hands of each individual.

Rotavirus risk of infection

Rotavirus is transmitted by contact with the mouth; hence plugging the parameters presented in Table 1 into equation (2) yields equation (4), in which C_{hand} is the concentration of rotavirus found on the subjects’ hands.

$$D = \frac{C_{hand} \text{ (number of viruses)}}{658 \cdot 94 \text{ cm}^2} \times \left(39 \times 6 \cdot 89 \text{ cm}^2 \times \frac{0 \cdot 13}{\text{minute}} \times 480 \text{ min} \right). \quad (4)$$

Table 3 shows the reduction in percent of infection for rotavirus with different initial concentrations of rotavirus on the hands. Again, because of the nature of the dose-response curve for rotavirus, greater reduction in risk of infection is seen with a lower initial concentration of virus on the hands.

DISCUSSION

MS-2 coliphage was used as a surrogate for rhinovirus, rotavirus and norovirus to quantify the exposure to virus both with and without use of a hand sanitizer three times a day. MS-2 virus is similar in shape and size to these viruses and has been used extensively as a surrogate to study the behaviour

Table 1. Parameters for use with equation (3)

Parameter	Parameter unit	Distribution	95% CI	Mean value used in the model	Source
A_{hand}	cm ²	Bootstrapping	533.1–881.6	658.94	[12, 13]
$f_{2,\text{eye}}$	fraction	Point estimate	–	0.339	[18]
$f_{2,\text{nose}}$	fraction	Point estimate	–	0.339	[18]
$f_{2,\text{mouth}}$	fraction	Point estimate	–	0.339	[18]
A_{eye}	cm ²	Bootstrapping	1.523–1.602	1.56	[12]
A_{nose}	cm ²	Bootstrapping	4.608–5.654	5.12	[12]
A_{mouth}	cm ²	Bootstrapping	6.63–7.13	6.89	[12]
N_{eye}	per minute	Bootstrapping	0.0247–0.0586	0.0411	[11]
N_{nose}	per minute	Bootstrapping	0.0546–0.1148	0.0848	[11]
N_{mouth}	per minute	Bootstrapping	0.0694–0.2283	0.1332	[11]

CI, Confidence interval.

Table 2. Probability of infection as a function of initial rhinovirus concentration on the hand

Concentration of rhinovirus on hands (TCID ₅₀)	Probability of infection		
	Before	After	% Reduction
3000	0.7484	0.3980	46.8
1500	0.7126	0.3361	52.8
1000	0.6894	0.2987	56.7
500	0.6456	0.2350	63.6
250	0.5961	0.1753	70.6
75	0.4967	0.0925	81.4
30	0.4113	0.0518	87.4
15	0.3439	0.0323	90.6
3	0.1929	0.0098	94.9
1	0.1378	0.0055	96.0

TCID, Tissue culture infective dose.

of human pathogenic viruses in the environment and assessment of disinfectants [19]. Its distribution on fomites in the household after addition to the hands [20] was very similar to that observed by Winther *et al.* who contaminated households with rhinovirus [21].

Total concentration of MS-2 coliphage inoculated on the hands of the subjects (one member only in each family) was 1×10^8 p.f.u. to ensure detection during laboratory analysis so that reductions of the virus on the hands could be quantified after the intervention. Rotavirus concentrations of 10^{12} rotavirus particles can be found in the stool of infected persons [22]. Data on the occurrence of rotavirus on the hands of infected persons is not available, although it is readily detected on individuals' hands in households in developing countries [22]. Norovirus has been detected

on the hands of infected individuals ranging from $10^{3.3}$ to $10^{4.45}$ genome copies by qPCR in hand rinse samples [3]. The concentration of rhinovirus in nasal mucus has been found to vary with the stage of infection and ranges from 10 to 1 000 000 tissue culture infectious viruses/ml [23].

An exposure model was developed based on the work of Nicas & Best to estimate the amount of virus that would reach the nose, eyes and mouth during the 8-h study period [11]. Using dose-response models for rhinovirus, rotavirus and norovirus, the risk of infection could then be quantified based on the amount of virus on the hands. Because data is not available on the actual concentration of virus that might be on the hands of infected and exposed individuals a range of values of risk of infection were determined for each of the viruses (Tables 2 and 3). Using the data on the reduction in concentrations of MS-2 on the hands post-intervention, the reduction in risk of infection can be determined based on different concentrations of the actual viral pathogens on the hands. It was found that because of the nature of the dose-response models, the less the amount of virus on the hands the greater the reduction in probability of infection. Thus, the risk of infection was reduced by 96% with one virus on the hands *vs.* 46.8% when 3000 viruses were present on the hands for rhinovirus. The same was true for rotavirus and norovirus. Risk reductions are only estimated for individuals who do not have protective immunity.

The potential for reduction in risk varied both between households and by household members (Fig. 3) based on the amount of MS-2 on their hands pre- and post-intervention. Variation in risk pre-intervention may vary depending on numerous factors including how well and how often family

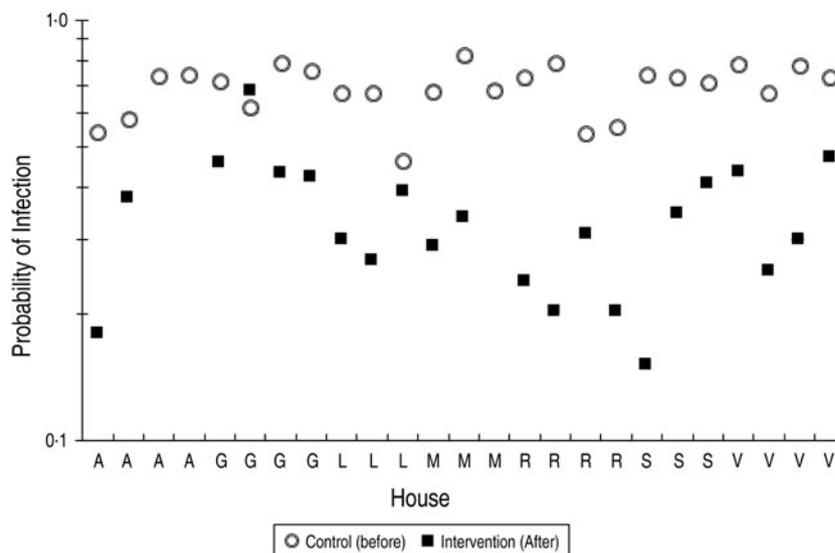


Fig. 3. Rhinovirus risk of infection before and after intervention assuming a concentration of 1000 infectious rhinoviruses on the hands. Each letter represents an individual in the household. Same letter represents a different individual in the same household.

Table 3. Reduction in probability of infection as a function of initial rotavirus concentration on the hand

Concentration of rotavirus on hands (TCID ₅₀)	Probability of infection		
	Before	After	% Reduction
3000	0.8079	0.4099	49.3
1500	0.7703	0.3350	56.5
1000	0.7451	0.2907	61.0
500	0.6960	0.2182	68.7
250	0.6388	0.1546	75.8
75	0.5211	0.0752	85.6
30	0.4199	0.0406	90.3
15	0.3410	0.0248	92.7
3	0.1726	0.0070	96.0
1	0.0897	0.0026	97.1

TCID, Tissue culture infective dose.

members practice routine hand washing and their activities within the home (i.e. how often they used the toilet, food preparation, playing, cleaning, etc.). Reduction in MS-2 on the hands of individuals post-intervention may be indicative of when they use the hand sanitizer during the day (that was left to their option).

Uncertainty arises from varying concentrations of respiratory and enteric viruses on the hands of infected individuals. Moreover, dose-response data was developed in normal healthy adult individuals

and may not be reflective of children, the immunocompromised and the elderly. Differences may also exist between the surrogate and the human pathogenic viruses in their survival on the hands/fomites and transfer efficiency to and from fomites, which could affect the degree of exposure [19, 21, 24].

Several epidemiological studies have shown the potential for reduction in transmission of respiratory and enteric infections with the use of hand sanitizers [25–27]. In a review of 16 studies on the impact of reduction of respiratory infections from the use of hand sanitizers Warren-Gash *et al.* concluded that success was dependent on setting, context and compliance [28]. The present study also provides evidence for the potential of alcohol-based hand sanitizers to reduce the risk of illness from respiratory and enteric infections in household settings.

The results of the present study suggest that adding the use of an alcohol-based hand sanitizer in the home can result in increased reduction in risk of infection by respiratory and enteric viruses, such as rotavirus, rhinovirus and norovirus, even when routine hand washing is already practised. Future studies should evaluate whether other hand hygiene interventions, such as use of hand sanitizing wipes, would have a similar effect.

DECLARATION OF INTEREST

None.

REFERENCES

1. CDC. Centers for Disease Control (<http://www.cdc.gov/>). Accessed 1 April 2014.
2. Gwaltney JM, Hendley JO. Transmission of experimental rhinovirus infection by contaminated surfaces. *American Journal of Epidemiology* 1981; **116**: 828–833.
3. Ward RL, et al. Prevention of surface-to-human transmission of rotaviruses by treatment with disinfectant spray. *Journal of Clinical Microbiology* 1991; **29**: 1991–1996.
4. Liu P, et al. Laboratory evidence of Norwalk virus contamination on the hands of infected individuals. *Applied and Environmental Microbiology* 2013; **79**: 7875–7881.
5. Hubner N, et al. Effectiveness of alcohol-based hand disinfectants in a public administration: impact on health and work performance related to acute respiratory symptoms and diarrhea. *BMC Infectious Diseases* 2010; **10**: 250.
6. Sattar SA, et al. Activity of an alcohol-based hand gel against human adeno-, rhino, and rotaviruses using the fingerpad method. *Infection Control and Hospital Epidemiology* 2000; **21**: 516–519.
7. Macinga DR, et al. Improved inactivation of non-enveloped enteric viruses and their surrogates by a novel alcohol-based hand sanitizer. *Applied and Environmental Microbiology* 2008; **74**: 5047–5052.
8. Teunis PF, et al. Norwalk virus: how infectious is it? *Journal of Medical Virology* 2008; **80**: 1486–1476.
9. Hendley JO, Edmondson Jr. WP, Gwaltney Jr. JM. Relation between naturally acquired immunity and infection of two rhinoviruses in volunteers. *Journal of Infectious Diseases* 1972; **125**: 243–248.
10. Haas CN, Rose JB, Gerba CP. *Quantitative Microbial Risk Assessment*, 2nd edn. New York: John Wiley, 2014.
11. Nicas M, Best D. A study quantifying the hand-to-face contact rate and its potential application to predicting respiratory tract infection. *Journal of Occupational and Environmental Hygiene* 2008; **5**: 347–352.
12. U.S. EPA. *Exposure Factors Handbook*. Washington, DC, 2011.
13. Snyder RG, et al. Anthropometry of infants, children, and youths to age 18 for product safety design. Final Report. Washington, DC, USA: University of Michigan Highway Safety Research Institute Consumer Product Safety Commission, 1977.
14. Mooney C, Duval R. *Bootstrapping: A Nonparametric Approach to Statistical Inference*, 1st edn. California: SAGE Publications Inc., 1993.
15. Ott RL, Longnecker M. *An Introduction to Statistical Methods and Data Analysis*, 5th edn. California: Duxbury Press, 2001.
16. Kabacoff RI. *R in Action: Data Analysis and Graphics with R*. New York: Manning Publications Co., 2011.
17. Zume N, Mount J. *Practical Data Science with R*, 1st edn. New York: Manning Publications, 2014.
18. Rusin P, Maxwell S, Gerba C. Comparative surface to hand and fingertip to mouth transfer efficiency of gram positive bacteria, gram negative bacteria, and phage. *Journal of Applied Microbiology* 2002; **93**: 585–592.
19. Ward RL, et al. Human rotavirus studies in volunteers: determination of infectious dose and serological response to infection. *Journal of Infectious Disease* 1986; **154**: 871–880.
20. Tamimi AH, et al. Impact of a hand sanitizer intervention on the spread of viruses in homes. *Food and Environmental Virology* 2014; **6**: 140–144.
21. Winther B, et al. Environmental contamination with rhinovirus and transfer to fingers of healthy individuals by daily life activity. *Journal of Medical Virology* 2007; **79**: 1606–1610.
22. Flewett TH. Clinical features of rotavirus infections. In: Tyrrell DAJ, Kapikan AZ, eds. *Virus Infections of the Gastrointestinal Tract*. New York: Marcel Dekker, 1982, pp. 125–146.
23. Sinclair JR, et al. Selection of microbial surrogates for studying the fate and control of pathogens in the environment. *Applied and Environmental Microbiology* 2012; **78**: 1969–1977.
24. Ansari AS, et al. Rotavirus survival on human hands and transfer of infectious virus to animate and nonporous inanimate surfaces. *Journal of Clinical Microbiology* 1988; **26**: 1513–1518.
25. Lopez GU, et al. Transfer efficiency of bacteria and viruses from porous and nonporous fomites to fingers under different relative humidity conditions. *Applied and Environmental Microbiology* 2013; **79**: 5728–5734.
26. Fendler EJ, et al. The impact of alcohol hand sanitizer use on infection rates in an extended care facility. *American Journal of Infection Control* 2002; **30**: 226–233.
27. Prazuck T, et al. Reducing gastroenteritis occurrences and their consequences in elementary schools with alcohol-based hand sanitizers. *Pediatric Infectious Disease Journal* 2010; **29**: 994–998.
28. Warren-Gash C, Fragaszy E, Hayward AC. Hand hygiene community transmission of influenza and acute respiratory tract infection: a systematic review. *Influenza and Other Respiratory Viruses* 2012; **7**: 738–449.