

Biodegradable and biobased mulch residues had limited impacts on soil properties but reduced yield of the following crop in a low fertility soil

Research Paper

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Abstract

Biodegradable and biobased mulch films and fabrics (BDMs) are potentially sustainable alternatives to polyethylene plastic mulch film (PE) in vegetable production because BDMs can be incorporated into the soil by tillage at the end of the growing season for decomposition. However, grower adoption has been limited in part by concerns about slow degradation rates and possible adverse effects on soil health and productivity. The objective of this study was to measure the effects of soil incorporated residues from two BDMs and compost on soil chemical and physical properties and vegetable crop yield across two diverse locations [Lincoln (LNK) and Scottsbluff (SBF), NE, USA] during a 2-yr study. The BDMs, including a polylactic acid biofabric with embedded wood particles (PLA; 1.14 mm thick and 298 g m⁻²), and a starch-polyester bioplastic mulch film (STP; 0.015 mm thick and 20 g m⁻²), were applied in May 2017 for vegetable production. Mulches were incorporated in soil by tillage in September 2017 in half of the experimental plots and removed in the other half as a control. Compost was applied in fall 2017 and 2018 at rates between 42 and 60 Mg ha⁻¹ to establish high and low (no compost) fertility soil environments within each location. Sweet corn (*Zea mays*) was grown in 2018 and cabbage (*Brassica oleraceae*) in 2019, and yield data were collected. The soil was sampled at ~6 month intervals for two years. The BDM residues had little effect on soil pH, organic matter or physical properties, but the incorporation of PLA in the soil at SBF reduced soil nitrate 6 months after the incorporation of residues. Nitrogen immobilization likely contributed to the 16% ± 5% reduction in sweet corn yield observed at SBF in plots without compost where BDM residues were incorporated compared to removed. No additional yield differences were detected in sweet corn (2018) or cabbage (2019) across locations or treatments, which suggests that BDM residues are less likely to immobilize nitrogen and reduce yield in high fertility soil environments. Given the potential environmental benefits of BDMs as an alternative to PE, future research should seek to mitigate the negative effects of BDM residues on crop yield, particularly in lower fertility soils.

Introduction

Small fruit and vegetable growers use polyethylene plastic mulch films (PE) for weed management, increased soil temperature, water retention and increased crop yields (Tofaneli and Wortman, 2020). While valuable, PE must be removed from the field after use and most is landfilled or burned (Kasirajan and Ngouajio, 2012). Even after removal, up to 11% (or 8.4 kg ha⁻¹) of PE is left behind in the field (He *et al.*, 2018). The PE fragments left in the soil can persist for decades (Chamas *et al.*, 2020) releasing toxins, adsorbing pollutants, and contaminating water or other organisms (Feuilloley *et al.*, 2005; Ashton *et al.*, 2010; Wang *et al.*, 2013; Briassoulis *et al.*, 2015).

Biodegradable and biobased mulch films and fabrics (BDMs) are potentially sustainable alternatives to PE because they can provide similar agronomic benefits and be left in the soil to decompose after their useful life (Tofaneli and Wortman, 2020). Commercially available BDMs include bioplastic films manufactured from starch and polyester polymer blends and paper mulch membranes. Many other experimental BDM products are in development including composite biofabrics manufactured from polylactic acid and agricultural residues (Samuelson *et al.*, 2022). Despite their promise, on-farm adoption of BDMs in lieu of PE has been limited. In a survey of Tennessee, Texas and Washington growers, premature deterioration of BDMs during the growing season and slow degradation after soil incorporation were identified as barriers to adoption (Goldberger *et al.*, 2015). Consistent with these grower concerns, recent studies have documented inconsistent durability of bioplastic and paper BDMs

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and the persistence of biofabric BDM residues in soil (Wortman *et al.*, 2015; Dharmalingam *et al.*, 2016; Thompson *et al.*, 2019).

As BDMs decompose or persist in soil, they have the potential to influence soil chemical and physical properties. Any changes to soil properties that negatively affect soil fertility and productivity could further contribute to BDM hesitancy among growers. The C/N ratio of BDMs is greater than 300 and there may be potential for soil N immobilization during BDM biodegradation (Thompson *et al.*, 2019). Nitrogen immobilization occurs in high C/N soil environments where microbial communities scavenge and store nitrogen within their cells, which renders it temporarily unavailable for crop uptake (Mooshammer *et al.*, 2014). This effect is commonly observed following soil incorporation of straw residues and cover crops, but less is known about how the decomposition of C-rich BDMs will influence soil nitrate availability (Reichel *et al.*, 2018).

Soil additions of C-rich amendments including wood chips and straw can increase soil organic matter (SOM), but the effects of soil-incorporated BDMs are unknown (Powlson *et al.*, 2008; Li *et al.*, 2018). Depending on BDM type, row spacing, and deterioration during the growing season, the mass of BDMs added to soil likely ranges from 50 (e.g., bioplastic films) to 2000 kg ha⁻¹ (e.g., biofabric and paper membranes). For comparison, manure and compost amendments typically range from 1000 to 50,000 kg ha⁻¹ (Wortman *et al.*, 2017) and cover crop biomass incorporated in soil often ranges from 2000 to 14,000 kg ha⁻¹ (Osipitan *et al.*, 2019). While BDM mass contributions are considerably lower than other amendments, potential effects on SOM are difficult to predict given the higher C/N ratio of BDMs (Thompson *et al.*, 2019).

Changes to soil microbial communities after incorporation of BDMs could indirectly contribute to changes in SOM and physical properties. After soil incorporation, some BDMs, including those containing polylactic acid polymers, can increase the abundance of soil fungi (Bandopadhyay *et al.*, 2018; Janczak *et al.*, 2018). One study found that the recovered mass of a polylactic acid BDM increased 160% over the course of 20 months due in part to the accumulation of mycelia (Ghimire *et al.*, 2017). Increases in soil fungi, often associated with the addition of organic amendments, have been shown to improve soil aggregate stability (Bossuyt *et al.*, 2001). Management practices that promote aggregate stability are particularly relevant in vegetable systems because intensive tillage that is typical for seedbed preparation and weed management reduces the abundance of C-rich macroaggregates (Six *et al.*, 2000). If BDM incorporation in soil influences SOM and aggregate stability, it is also likely to influence related physical properties including soil sorptivity (initial water infiltration), aggregate tensile strength, and others (Murphy and Murphy, 2015).

While polylactic acid BDMs may contribute to improved soil structure, there is some evidence that bioplastic BDMs could behave like PE films in soil and degrade soil structure until fully decomposed. Repeated incorporation of BDMs in the soil can increase the abundance of microplastics (Yu *et al.*, 2021), but it is not clear if degrading BDM fragments and microplastics affect soil physical properties in the same way that PE microplastics do (de Souza Machado *et al.*, 2018). Long-term accumulation of PE in soil has been shown to reduce soil organic C (Liu *et al.*, 2021), and Jiang *et al.* (2017) reported decreased soil water content, bulk density, and saturated hydraulic conductivity, and increased porosity when PE residues were present in the soil. However, Qi *et al.* (2020a) found that residues of PE and

bioplastic BDM had no effect on aggregate stability. Gao *et al.* (2021) found that BDM residues [from a polylactic acid/polybutylene-adipate-co-terephthalate film] increased SOM in the first year after soil incorporation, but had no effect on aggregate stability, bulk density, porosity, EC or pH; however, PE residues increased bulk density and reduced porosity.

Changes in soil properties reported by Gao *et al.* (2021) influenced subsequent crop yield. The BDM residues in soil from a previous potato (*Solanum tuberosum*) crop (51.6 kg ha⁻¹ at time of planting) did not affect subsequent rice (*Oryza sativa*) crop yield, but 30.0 kg ha⁻¹ PE residue reduced rice yield compared to the no residue control. Similarly, Hu *et al.* (2020) found that PE residues greater than 300 kg ha⁻¹ negatively impacted maize (*Zea mays*) root and shoot growth under field conditions, partly due to reduced water use efficiency. However, a pot study conducted by Qi *et al.* (2018) demonstrated that residues of a starch-based BDM reduced wheat (*Triticum aestivum*) growth, particularly in the first two months of growth, and the negative effects of PE were less pronounced compared to BDM. Variable effects of mulch residues on soil properties and crop productivity suggest additional research is needed to better understand the importance of mulch composition and mass, local conditions, management and subsequent crops.

The objective of this study was to evaluate the effects of two different types of BDM residue and compost on soil chemical and physical properties and vegetable crop yield across two diverse environments. Generally, we hypothesized that differences in BDM composition and residue management (incorporation in soil compared to removal) would drive differences in soil nitrogen dynamics, soil physical properties, and crop yields in the two years following the initial use of the mulch.

Materials and methods

Experimental locations and design

We conducted experiments at two climatically distinct locations in Nebraska between 2017 and 2019. Locations included the University of Nebraska-Lincoln East Campus Research Farm in Lincoln, NE (40.84 N, 96.65 W elevation = 351 m) and the Panhandle Research and Extension Center in Scottsbluff, NE (41.89 N, 103.68 W). The soil at Lincoln (LNK) is Zook silty clay loam, which is a fine, smectitic, mesic Cumulic Vertic Endoaquolls, and the climate is humid continental with mean annual precipitation of 735 mm and a mean daily temperature 10.8°C. The soil at Scottsbluff (SBF) is a Tripp very fine sandy loam, which is a coarse-silty, mixed, superactive, mesic Aridic Haplustoll, and the climate is semi-arid with mean annual precipitation of 399 mm and a mean daily temperature of 9.3°C. Relative precipitation and temperature differences observed between locations during the experiment were similar to historical trends (Fig. 1).

The experiments were designed using a randomized complete block, split-split plot design with three replicate blocks at each location. The main plot was mulch type applied in 2017 for a sweet pepper (*Capsicum annuum*) crop. Mulches included a black starch-polyester bioplastic BDM (Mater-Bi®, Novamont S.P.A.; Shelton, CT, USA; hereafter abbreviated as STP) or a novel polylactic acid-based biofabric BDM (3M Company, St. Paul, MN, USA; hereafter abbreviated as PLA). The STP mulch was 0.015 mm thick and had an area density of 20 g m⁻². The PLA mulch was comprised of two layers of spunbond

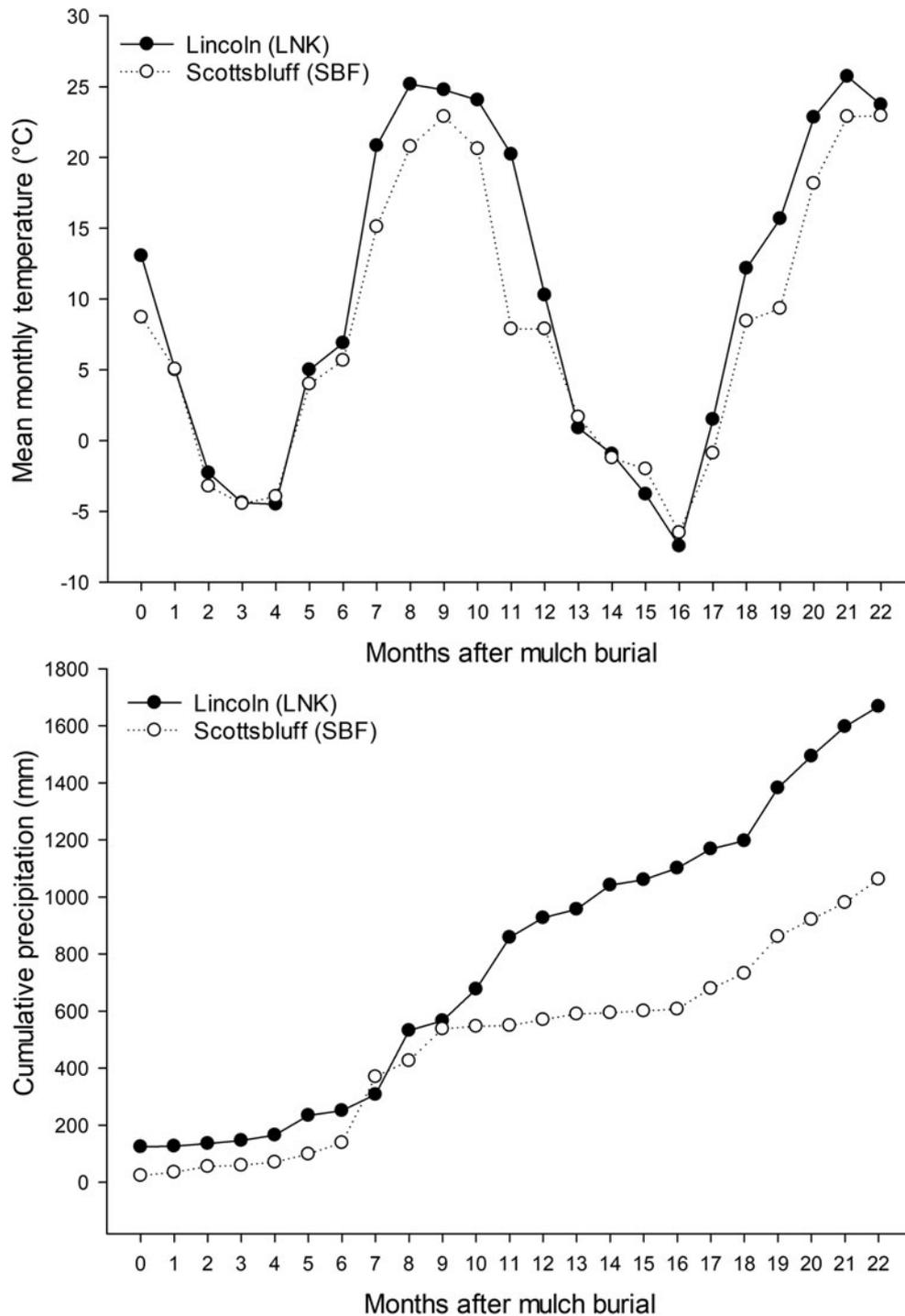


Fig. 1. Mean monthly air temperature (top) and cumulative precipitation (bottom) for Lincoln (LNK) and Scottsbluff (SBF) locations from Oct. 2017 through Aug. 2019 (Samuelson *et al.*, 2022).

polylactic acid, with a meltblown interior layer that included fine-grade wood particles (62% of total mulch mass) in the fiber matrix. The PLA mulch was 1.14 mm thick with an area density of 298 g m⁻². The STP mulch is certified biodegradable in soil but is not 100% biobased and therefore cannot be incorporated in soil on certified organic farms in the U.S. (Miles *et al.*, 2017). In contrast, the PLA mulch is 100% biobased and certified compostable, and was included to explore its potential as an allowable input and replacement for PE on certified organic farms (though it

was later discovered that biodegradation rates in soil were unlikely to meet the required standards; Thompson *et al.*, 2019).

The subplot treatment in the experiment was the incorporation status of mulch, where the mulch was either incorporated in soil via a reciprocating spading machine (Celli Y70, Celli SpA; Forlì, Italy) after the initial 2017 growing season or removed from the field as a control. The experiment also included sub-subplot treatments to assess the effects of organic soil amendments (including compost, compost extracts, cover crops, and a combination of all

amendments) on the degradation rate of mulch (Samuelson *et al.*, 2022), but data analyzed for this paper were limited to a comparison of compost-amended plots and control plots without compost to establish an extreme gradient of soil fertility within and between locations.

Mulch type main plots were 65.8 m long and 1.83 m wide and randomized within replicate blocks; incorporation status subplots were 32.9 m long and 1.83 wide and randomized within main plots; and compost and control sub-subplot experimental units measured 5.5 m long and 1.83 m wide and were randomized within subplots. The 1.83 m plot width included a 0.76 m raised bed top with a single crop row and half of the between row area on each side of the bed top (1.07 m). There was also a 0.3 m buffer between rows of plots which resulted in a total distance of 2.13 m between rows of crops in different sub-subplots.

Cropping systems management

In spring 2017, prior to crop planting, the STP and PLA mulches were applied with a raised bed mulch layer (RB448; Nolt's Produce Supplies, Leoloa, PA, USA) with dripline applied under the mulch for irrigation. Plots were irrigated regularly to maintain soil matric potential above 25 kPa at SBF and 75 kPa at LNK (Irmak *et al.*, 2016). Sweet peppers (cv. 'Carmen') were started from seed in the greenhouse 8 weeks before transplanting into the field on 16 May 2017 at LNK and on 6 June 2017 at SBF. Peppers (and planting holes in the mulch) were spaced 46 cm between plants in a single row and 12 plants per sub-subplot. Fish emulsion fertilizer (2.9-3.5-0.3 NPK; OrganicGem, Advanced Marine Technologies, New Bedford, MA, USA) was applied at a rate of 0.3 g N, 0.4 g P and 0.03 g K per planting hole at planting and twice more throughout the season at the same rate. A cumulative total of 84 kg ha⁻¹ N blood meal fertilizer (13-1-0 NPK, Earthworks Health, LLC, Norfolk, NE, USA) was added during the season. After the 2017 pepper harvest, crop residue was shredded and mulch was incorporated via tillage (incorporation plots) or removed by hand (control plots). Mulch was incorporated or removed on 29 September 2017 at LNK and on 5 October 2017 at SBF. Both mulch types were largely intact (free of major rips or tears) at the time of soil incorporation.

Compost was applied to the appropriate sub-subplots and incorporated in soil at the same time as mulch residues in fall 2017. Compost was applied a second time approximately 12 months later after crop harvest but was not incorporated in the soil. The compost used at LNK was a municipal yard waste compost and the compost used at SBF was a composted beef feedlot manure. To account for the different feedstocks, compost rate was standardized across locations by total organic nitrogen with a target application of 504 kg ha⁻¹ N. This resulted in application rates of 57 Mg ha⁻¹ in 2017 and 60 Mg ha⁻¹ in 2018 (dry weight) at LNK; and 42 Mg ha⁻¹ in 2017 and 51 Mg ha⁻¹ in 2018 at SBF.

In preparation for planting sweet corn (*Zea mays*) in 2018, organic soybean meal fertilizer (7-1-2 NPK; Phyta-Grow, California Organic Fertilizers, Inc., Hanford, CA, USA) was applied at a rate of 67 kg ha⁻¹ N to all plots. Sweet corn (cv. 'Xtra-Tender 2171') was sown on 24 May 2018 at LNK and on 31 May 2018 at SBF. Two rows spaced 60 cm apart were planted on each bed top the length of the experiment. Weeds were managed throughout the season with hand hoes to avoid the disturbance of buried mulches. Sweet corn was harvested on 23 July 2018 at LNK and on 30 July 2018 at SBF. Total and marketable yield were measured by harvesting whole ears from all

plants in a plot. After harvest, crop and weed residues were shredded on 12 September 2018 at LNK and on 21 September 2018 at SBF.

Cabbage (*Brassica oleraceae* var. *sabauda* cv. 'Melissa') followed sweet corn in the crop rotation. Bloodmeal fertilizer (13-1-0 NPK, Earthworks Health, LLC, Norfolk, NE, USA) was applied to all plots prior to planting at a rate of 34 kg ha⁻¹ N and raked into the surface soil before application of a white on black PE mulch for weed management. As in sweet corn, the experimental areas were not mechanically tilled and beds were not created so as to avoid disturbing buried mulch residues. Cabbage seeds were started in the greenhouse approximately eight weeks prior to transplanting on 16 May 2019 at LNK and on 5 June 2019 at SBF. Plants were spaced 46 cm apart within rows and 12 plugs were planted in each sub-subplot. Fish emulsion fertilizer (2.9-3.5-0.3 NPK; OrganicGem, Advanced Marine Technologies, New Bedford, MA, USA) was applied after transplanting at a rate of 0.5 g N, 0.7 g P and 0.05 g K per planting hole. A *B.t.* insecticide (GardenSafe Spectrum Brands, Middleton, WI, USA) was sprayed weekly at a rate of 3.9 mL L⁻¹ from 24 June through harvest at LNK and from 30 July through harvest at SBF. Cabbage was harvested on 5 August 2019 at LNK and on 19 August 2019 at SBF. Cabbage was separated as marketable (dense heads with no insect damage) or cull and then weighed.

Soil sampling and analysis

For the determination of nitrate and SOM concentrations, the soil was sampled in the spring and fall of 2018 and 2019 at approximately 6, 12, 18 and 22 months after mulch was incorporated or removed in fall 2017. Eight soil cores (1.9 cm diameter by 20 cm depth) were collected from each sub-subplot. These samples were analyzed for nitrate (KCl extraction), pH (1:1 soil:water dilution) and SOM (loss of weight on ignition) (Ward, 2021).

Soil sorptivity was measured in all sub-subplots at both locations at 6 months after mulch incorporation and in control sub-subplots at 18 months after mulch incorporation in soil using methods detailed by Smith (1999). Metal rings (10.5 cm height by 9.8 cm diameter) were carefully pressed into undisturbed soil and 75 mL of water was poured into the ring along the edge to minimize soil disturbance. Four readings were taken from each sub-subplot at each time. The time required for infiltration of the 75 mL of water was recorded, and four readings were collected in each plot. From these readings, soil sorptivity was calculated as:

$$\text{Soil sorptivity} = \frac{H}{\sqrt{t}}$$

whereby H is the head of water in cm and t is the time to infiltration in seconds.

Soil penetration resistance was measured in all sub-subplots at both locations 6 and 18 months after mulch incorporation in soil using the force meter with an 8 mm diameter cone head. Three measurements per sub-subplot were taken to a depth of 5 cm with a speed of approximately 1 cm s⁻¹, and readings were converted to MPa based on the basal cone area and averaged (Rakkar *et al.*, 2017).

Wet aggregate stability was determined by the wet sieving method to determine the proportion of macroaggregates (>0.25 mm diameter) and microaggregates (<0.25 mm diameter);

(Nimmo and Perkins, 2002; Rakkar *et al.*, 2017). Two bulk soil samples for wet aggregate stability were collected to a depth of 5 cm from all sub-subplots at both locations at 6 months after mulch incorporation and from control sub-subplots at 18 months after mulch incorporation in soil. Air-dried soil samples were dry sieved through an 8 mm sieve and 50 ± 1 g were placed on top of a stack of sieves. The three sieves had openings of 2, 1 and 0.25 mm. Soil samples were saturated by capillary action for 10 min and then oscillated up and down 3 cm for 10 min in a water tank at 30 strokes per minute. Aggregates remaining on each sieve were collected into pre-weighed beakers, which were then oven-dried at 105°C and weighed again. For samples collected at SBF, the amount of soil aggregates was corrected for sand content (because of the sandy loam soil texture at SBF, compared to silty clay loam at LNK) (Nimmo and Perkins, 2002). Sand correction accounts for loose sand accumulated from sieving, which can cause inaccuracies in the ratio of total stable aggregates and make it difficult to distinguish differences in macroaggregates (Nimmo and Perkins, 2002). Sand and aggregates were collected from each sieve, dried and weighed, then treated with 30 mL of 0.5% (NaPO_3)₆ to break up aggregates. Samples were then washed in a 53- μm sieve. The remaining sand was dried at 105°C, weighed, and used to correct the original aggregate mass collected on each sieve.

Soil tensile strength was measured at 6 and 18 months after mulch incorporation in the soil in all plots with a force meter equipped with a rounded, flat head (FDX Force Ten, Wagner Instruments, Greenwich, CT, USA) following the methods of Öztaş *et al.* (1999) and Dexter and Kroesbergen (1985). To minimize variation, 20 air-dried aggregates (collected from top 5 cm of surface) between 4.75 and 8 mm were crushed individually between a flat metal disk sitting on a metal plate and the force meter head. The maximum force required to crush each aggregate was measured and tensile strength was calculated as:

$$\text{Tensile strength (kPa)} = 0.576 \left(\frac{F}{D^2} \right)$$

whereby F is the force required to break the aggregate (N), D is the diameter of the aggregate (m), and 0.576 is the value of the coefficient of proportionality between the applied compressive load and the inner tensile strength of the aggregate (Dexter and Kroesbergen, 1985).

Statistical analysis

Analysis of variance was conducted with the *glimmix* procedure in SAS 9.4 (SAS Institute, Cary, NC, USA) to test for fixed effects of mulch type, incorporation status, compost, and all possible two- and three-way interactions on soil properties and crop yield. Replicate block and the interaction of block and mulch type were included as random effects. Locations and sample dates were analyzed individually due to confounding and interacting effects (e.g., different sample dates across locations), and because many of the soil properties measured are strongly influenced by temporal dynamics beyond the scope of our objectives (e.g., soil nitrate abundance fluctuates depending on crop growth stage). Instead, analysis of mulch type, incorporation status, and compost at individual locations and sample times allows for a relative comparison of soil property changes independent of the expected spatial and temporal dynamics. Means separation was determined for

significant effects and interactions using the Tukey's HSD test with a significance threshold of $P < 0.05$.

Results and discussion

Mulch residue in soil

Mulch was applied in spring 2017 at both locations at a rate of 1744 kg ha⁻¹ for PLA and 133 kg ha⁻¹ for STP (per ha estimate assumes 1.83 m spacing between crop rows). Based on mulch degradation estimates from Samuelson *et al.* (2022), the mass of PLA at LNK ranged from approximately 1302 kg ha⁻¹ at 6 months after incorporation in soil to 580 kg ha⁻¹ at 22 months; and mass of STP at LNK ranged from 127 kg ha⁻¹ at 6 months and was fully degraded by 22 months. At SBF, the mass of PLA in soil ranged from 827 kg ha⁻¹ at 6 months to 512 kg ha⁻¹ by 22 months; and STP ranged from 132 kg ha⁻¹ at 6 months to 64 kg ha⁻¹ at 22 months. The mass of mulch residue added to soil was greater than Hu *et al.* (2020) where 51.6 kg ha⁻¹ was added before rice production; however, residue levels added – particularly for STP – were consistent with Ghimire *et al.* (2020) and followed similar trends in mulch degradation between 6 and 22 months. Effects of initial mulch residue levels in soil greater than 1000 kg ha⁻¹, as in the case of PLA, have not been previously studied.

Soil chemical properties

Soil pH at LNK was affected by the interacting effects of mulch incorporation and compost at 6 months whereby the combination of compost and BDM residues incorporated in soil lowered soil pH (6.7 ± 0.2) compared to compost without BDM residues in soil (7.2 ± 0.2). However, there were no differences in soil pH between 12 and 22 months after mulch incorporation (Table 1). At SBF, soil pH was influenced by the interaction of mulch by incorporation at 12 months because incorporation of STP residues increased soil pH (8.44 ± 0.04) compared to removal (8.32 ± 0.04) (Table 2); no additional changes in pH were observed at SBF. Qi *et al.* (2020b) reported a short-term increase in pH (6.72 to 6.90) during a two-month soil incubation of micro-fragments of a starch-polyester BDM; however, after four months, there were no differences in pH. In a similar field study, Li *et al.* (2014a) did not observe any effect of BDM residues (including starch-polyester bioplastics, a polylactic acid biofabric, and a paper mulch) on soil pH between 0 and 18 months after mulch incorporation across three diverse locations throughout the U.S.

Biodegradation and hydrolysis of polylactic acid releases lactic acid and should acidify soil, but this hydrolysis is unlikely to occur under typical field soil conditions because the glass-transition temperature of polylactic acid is 49.2°C (Karamanlioglu and Robson, 2013; Hayes *et al.*, 2017). The PLA residues in soil, excluding degraded wood fibers, were relatively stable throughout the 22 months of this experiment (Samuelson *et al.*, 2022); thus, we would not expect that polylactic acid in the PLA mulch would have driven changes in soil pH. One difference in the polylactic acid-based mulch studied here, compared to other biofabrics (e.g., Li *et al.*, 2014a), is that PLA contained 62% wood fibers by weight. However, wood chip mulch and soil amendments do not typically alter soil pH, even as the rate of amendment or thickness of mulch increases (Greenly and Rakow, 1995; Tahboub *et al.*, 2008). Given the minor and inconsistent changes in soil pH observed here and the unexplained connections to

Table 1. Soil pH, organic matter (SOM) and nitrate sampled approximately 6, 12, 18 and 22 months after mulch residues (a starch-polyester bioplastic [STP] or a polylactic acid biofabric with embedded wood particles [PLA]) were removed or incorporated in soil in Lincoln, NE

| | Lincoln | | | | | | | | | | | |
|---------------------------------|----------|---------|--------------------------------|-----------|---------|--------------------------------|-----------|---------|--------------------------------|-----------|---------|--------------------------------|
| | 6 months | | | 12 months | | | 18 months | | | 22 months | | |
| | pH | SOM (%) | Nitrate (mg kg ⁻¹) | pH | SOM (%) | Nitrate (mg kg ⁻¹) | pH | SOM (%) | Nitrate (mg kg ⁻¹) | pH | SOM (%) | Nitrate (mg kg ⁻¹) |
| <i>STP</i> | | | | | | | | | | | | |
| Removed | | | | | | | | | | | | |
| Compost | 7.0 | 3.5 | 15.4 | 7.3 | 3.7 | 7.6 | 7.4 | 4.1 | 6.4 | 7.4 | 4.3 | 7.4 |
| None | 6.7 | 3.6 | 14.3 | 6.9 | 3.8 | 6.4 | 6.9 | 4.2 | 4.3 | 7.0 | 4.3 | 5.2 |
| Incorporated | | | | | | | | | | | | |
| Compost | 6.6 | 4.2 | 14.4 | 7.1 | 4.1 | 6.4 | 7.1 | 5.0 | 6.3 | 7.3 | 5.1 | 5.9 |
| None | 7.0 | 3.7 | 14.2 | 7.1 | 3.7 | 5.2 | 7.1 | 4.2 | 4.4 | 7.2 | 4.3 | 5.1 |
| <i>PLA</i> | | | | | | | | | | | | |
| Removed | | | | | | | | | | | | |
| Compost | 7.3 | 3.6 | 16.0 | 7.3 | 3.7 | 6.0 | 7.4 | 4.5 | 9.7 | 7.4 | 4.5 | 6.9 |
| None | 7.0 | 3.5 | 14.5 | 7.2 | 3.5 | 4.4 | 7.2 | 3.8 | 4.0 | 7.4 | 4.0 | 4.9 |
| Incorporated | | | | | | | | | | | | |
| Compost | 6.8 | 4.3 | 23.0 | 7.0 | 4.3 | 7.8 | 7.2 | 5.2 | 10.4 | 7.3 | 5.2 | 9.6 |
| None | 7.1 | 3.7 | 15.9 | 7.3 | 3.7 | 5.5 | 7.3 | 4.3 | 4.7 | 7.4 | 4.4 | 4.9 |
| <i>Standard error</i> | 0.2 | 0.4 | 2.0 | 0.2 | 0.4 | 0.8 | 0.2 | 0.4 | 1.2 | 0.2 | 0.5 | 1.1 |
| <i>Significance</i> | | | | | | | | | | | | |
| Mulch | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns |
| Incorporation | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns |
| Mulch × incorporation | ns | ns | ns | ns | ns | * | ns | ns | ns | ns | ns | ns |
| Compost | ns | ns | ns | ns | ns | ** | ns | ns | *** | ns | ns | ** |
| Mulch × compost | ns | ns | ns | ns | ns | ns | ns | ns | ** | ns | ns | ns |
| Incorporation × compost | * | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns |
| Mulch × incorporation × compost | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns |

Results of analysis of variance for effects of mulch (STP or PLA), incorporation (removed or incorporated), and compost for each soil property are included at the bottom of the table and significance of each factor is indicated as: 'ns' = not significant; * = $P < 0.05$; ** = $P < 0.01$, *** = $P < 0.001$.

mulch type or incorporation, it is unlikely that biodegradation of STP or PLA residues alter soil pH enough to impact productivity in the short term.

Compost generally increased soil nitrate throughout the experiment as expected, but mulch type and incorporation also influenced soil nitrogen dynamics across both locations (Tables 1 and 2). The most notable changes in soil nitrate were observed at 6 months at SBF and at 18 to 22 months across both locations. Soil nitrate at 6 months at SBF was reduced following incorporation of PLA residues ($3.4 \pm 0.3 \text{ mg kg}^{-1}$) compared to removal ($4.7 \pm 0.3 \text{ mg kg}^{-1}$), but there was no difference between removal and incorporation of STP residues (e.g., no N immobilization). At 18 months, soil nitrate was greatest in the PLA × compost plots at LNK and SBF. At LNK, soil nitrate in PLA × compost plots was $10.1 \pm 1.0 \text{ mg kg}^{-1}$ compared to $6.3 \pm 1.0 \text{ mg kg}^{-1}$ in STP × compost plots and $4.3 \pm 0.9 \text{ mg kg}^{-1}$ in plots without compost. Similarly, soil nitrate in PLA × compost plots was $3.1 \pm 0.2 \text{ mg kg}^{-1}$ compared to $2.0 \pm 0.2 \text{ mg kg}^{-1}$ in STP × compost plots

and less in plots without compost. And the trend continued at SBF at 22 months where soil nitrate was greatest in plots where PLA residues were incorporated with compost ($10.2 \pm 1.6 \text{ mg kg}^{-1}$ compared to $\leq 7.1 \text{ mg kg}^{-1}$ in all other treatments) (Tables 1 and 2).

Given the mass of PLA added and its high C/N, we expected short-term N immobilization following incorporation in soil. Immobilization was observed at SBF but not at LNK, and only 6 months after incorporation. This nitrogen immobilization, though modest and site-specific, contrasts with Li *et al.* (2014a) where no BDM residues influenced soil nitrate between 6 and 18 months after mulch incorporation. Instead, Li *et al.* (2014a) found that differences in soil nitrate were driven by location, time and production system. Nitrogen immobilization following soil amendment with C-rich organic materials (e.g., wheat straw or wood chips) is most often observed when applied at rates (2500 to 5000 kg ha⁻¹; Döring *et al.*, 2005) or depths (>15 cm; Hoagland *et al.*, 2008) greater than what was added here (effective

Table 2. Soil pH, organic matter (SOM) and nitrate sampled approximately 6, 12, 18 and 22 months after mulch residues (a starch-polyester bioplastic [STP] or a polylactic acid biofabric with embedded wood particles [PLA]) were removed or incorporated in soil in Scottsbluff, NE

| | Scottsbluff | | | | | | | | | | | |
|---------------------------------|-------------|---------|--------------------------------|-----------|---------|--------------------------------|-----------|---------|--------------------------------|-----------|---------|--------------------------------|
| | 6 months | | | 12 months | | | 18 months | | | 22 months | | |
| | pH | SOM (%) | Nitrate (mg kg ⁻¹) | pH | SOM (%) | Nitrate (mg kg ⁻¹) | pH | SOM (%) | Nitrate (mg kg ⁻¹) | pH | SOM (%) | Nitrate (mg kg ⁻¹) |
| <i>STP</i> | | | | | | | | | | | | |
| Removed | | | | | | | | | | | | |
| Compost | 8.3 | 1.7 | 4.3 | 8.3 | 1.7 | 5.5 | 8.3 | 1.7 | 1.9 | 8.3 | 1.9 | 6.5 |
| None | 8.3 | 1.5 | 3.2 | 8.3 | 1.6 | 3.8 | 8.3 | 1.5 | 1.2 | 8.3 | 1.6 | 3.1 |
| Incorporated | | | | | | | | | | | | |
| Compost | 8.4 | 1.6 | 4.5 | 8.5 | 1.6 | 2.5 | 8.4 | 1.7 | 2.2 | 8.3 | 1.9 | 6.9 |
| None | 8.2 | 1.5 | 3.5 | 8.4 | 1.5 | 4.7 | 8.4 | 1.4 | 1.2 | 8.3 | 1.6 | 7.0 |
| <i>PLA</i> | | | | | | | | | | | | |
| Removed | | | | | | | | | | | | |
| Compost | 8.4 | 1.5 | 5.9 | 8.4 | 1.6 | 3.1 | 8.4 | 1.8 | 2.6 | 8.3 | 2.0 | 6.4 |
| None | 8.2 | 1.5 | 3.5 | 8.4 | 1.6 | 4.7 | 8.3 | 1.5 | 1.3 | 8.3 | 1.6 | 4.6 |
| Incorporated | | | | | | | | | | | | |
| Compost | 8.3 | 1.6 | 3.6 | 8.4 | 1.7 | 5.4 | 8.5 | 1.9 | 3.6 | 8.3 | 2.0 | 10.2 |
| None | 8.3 | 1.5 | 3.2 | 8.3 | 1.5 | 5.2 | 8.4 | 1.4 | 1.5 | 8.3 | 1.6 | 3.9 |
| <i>Standard error</i> | 0.07 | 0.05 | 0.4 | 0.05 | 0.06 | 1.2 | 0.05 | 0.07 | 0.2 | 0.08 | 0.06 | 1.5 |
| <i>Significance</i> | | | | | | | | | | | | |
| Mulch | ns | ns | ns | ns | ns | ns | ns | ns | * | ns | ns | ns |
| Incorporation | ns | ns | ns | ns | ns | ns | ns | ns | * | ns | ns | * |
| Mulch × incorporation | ns | ns | * | * | ns | ns | ns | ns | ns | ns | ns | ns |
| Compost | ns | ** | * | ns | ** | ns | ns | *** | *** | ns | *** | ** |
| Mulch × compost | ns | ns | ns | ns | ns | ns | ns | ns | ** | ns | ns | ns |
| Incorporation × compost | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns |
| Mulch × incorporation × compost | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | * |

Results of analysis of variance for effects of mulch (STP or PLA), incorporation (removed or incorporated), and compost for each soil property are included at the bottom of the table and significance of each factor is indicated as: 'ns' = not significant; * = $P < 0.05$; ** = $P < 0.01$, *** = $P < 0.001$.

depth of 1.14 mm and mass of approximately 1744 kg ha⁻¹ before incorporation in soil). Much of the C in polylactic acid is not bio-available in soil due to, as previously exposed, the high glass-transition temperature (49.2°C; Hayes *et al.*, 2017), which suggests that 38% of the 1744 kg ha⁻¹ in the PLA mulch (approximately 663 kg ha⁻¹; the proportion of PLA mass as polylactic acid polymer) would not contribute to N immobilization in the short term. However, the remaining addition of approximately 1081 kg ha⁻¹ wood particles from PLA residues was apparently enough to immobilize soil nitrate in this study at SBF (lower soil fertility), but not at LNK (higher soil fertility) (Tables 1 and 2). However, repeated applications or increased wood particle loading rates in the mulch could increase the potential for immobilization in high fertility soils.

Increased soil nitrate associated with PLA and compost 18 or more months after mulch use was surprising given the carbon-rich nature of the polylactic acid and embedded wood particles. Saprophytic fungi abundance was greater on PLA mulch residues and adjacent soil (data not shown; Samuelson, 2019), and perhaps the fungi contributed to accelerated decomposition and eventual mineralization of nitrogen from more recalcitrant organic materials in the compost (Masunga *et al.*, 2016). Moreover, saprophytic fungi have been shown to contribute to significant nitrogen immobilization in soil (Beare *et al.*, 1992); thus, it is possible that the nitrate initially immobilized in fungal biomass after use of PLA mulch was mineralized as the PLA, wood fiber carbon source and microbial biomass declined over time.

Compost amendment increased SOM at SBF, but SOM was not affected by mulch type or incorporation at either location or any of the sampling events between 6 and 22 months after mulch incorporation or removal ($P > 0.05$; Tables 1 and 2). Gao *et al.* (2021) reported a small increase in SOM after incorporation of BDM in soil (1.47 mg kg⁻¹ compared to 1.33 mg kg⁻¹ in the no residue control), but only measured to a depth of 5 cm. SOM in this study was measured to a depth of 20 cm and given the relatively small one-time addition of organic matter, we did not expect to observe measurable changes in SOM among treatments. Li *et al.* (2014b) reported modest variation in total organic C after BDM residue incorporation in soil at one of three locations in a high tunnel (e.g., 1.27 mg kg⁻¹ in the control compared to 1.64 mg kg⁻¹ 18 months after the incorporation of a paper mulch in soil), but data were not analyzed for differences among mulch types over time. Though small and potentially undetectable, any short-term changes to SOM following the incorporation of BDM residues are likely to be positive.

Soil physical properties

Soil sorptivity did not differ among mulch types or incorporation treatments across years or locations ($P > 0.05$; Tables 3 and 4). Jiang *et al.* (2017) observed increased infiltration rates due to PE film residues in soil because of paths of preferential flow and macropores created by residue fragments in the soil matrix. Similarly, Sintim *et al.* (2021) found that PE and BDM residues increased infiltration rates by 10–12% compared to a no-mulch control at one of the two locations tested. Soil sorptivity, as a measure of initial water infiltration, influences irrigation wetting patterns, and reduced sorptivity could limit the effectiveness of drip irrigation in vegetable production (Minasny and Cook, 2011). However, residues of STP and PLA did not negatively impact soil sorptivity in this study.

Compost reduced soil aggregate tensile strength at 18 months after soil incorporation at LNK, but soil aggregate tensile strength was not influenced by mulch or incorporation at either location (Tables 3 and 4). Organic soil amendments, including compost and biochar, can reduce aggregate tensile strength. Increasing application rates of corn cob biochar led to proportionate reductions in tensile strength of soil aggregates between 4 and 16 mm in diameter sampled approximately six months after application (Amoakwah *et al.*, 2017). The reduced aggregate tensile strength can improve soil tilth and workability, which are important soil physical properties in vegetable production for promoting the seedling establishment and root growth. However, longer-term changes in aggregate tensile strength are less common. After four decades of straw incorporation at rates between 0 and 12 Mg ha⁻¹, Qi *et al.* (2022) found no difference in aggregate tensile strength among treatments.

Soil penetration resistance was generally not different among mulch types or incorporation treatments across years or locations. The one exception was a mulch by incorporation interaction at SBF at 6 months, where compost had different effects on soil penetration resistance depending on mulch type (Tables 3 and 4). Mulch residues are less dense than most soil minerals and prior to decomposition can reduce bulk density and increase aeration, which may facilitate root penetration (Zhou *et al.*, 2021). Sintim *et al.* (2021) found that penetration resistance near the soil surface (0 to 5 cm) decreased after four consecutive years of BDM use and residue incorporation in soil compared to the no-mulch control. However, BDM soils were not different from PE plots where the mulch was removed annually, which suggests that BDM use – not residues in soil – reduced near-surface compaction. Mulch was only used in two of three years of this study (BDM in 2017 pepper and PE in 2019 cabbage), compared to four consecutive years in Sintim *et al.* (2021). Our results and those from Sintim *et al.* (2021) suggest that BDM residue from a single season is unlikely to alter soil penetration resistance in a predictable, consistent way.

Macroaggregate stability was not affected by mulch type, incorporation or compost at either location (Tables 3 and 4). We had expected that greater fungal abundance and glomalin on and around PLA residues would contribute to improved soil aggregation, but that was not observed (Carrizo *et al.*, 2018; Samuelson, 2019). Sintim *et al.* (2021) reported more consistently positive effects of BDMs on aggregate stability as annual use and incorporation of BDM residues increased aggregate stability by up to 16% compared to the no mulch control. However, as was the case with penetration resistance, aggregate stability was not different between BDM treatments and PE film (which was removed from the field at the end of each season). This suggests that improvements in aggregate stability observed by Sintim *et al.* (2021) were related to the use of the BDM or PE during the growing season, not the result of BDM residue biodegradation in soil. The lack of residue effects on aggregate stability is consistent with two other recent studies where BDM and PE residues in soil did not affect aggregate stability (Qi *et al.*, 2020a; Gao *et al.*, 2021).

Crop yields

Sweet corn yield at SBF in the first season following mulch use was influenced by the interacting effects of compost and incorporation (Table 5). In the absence of compost, mulch residue incorporation reduced yield by 16% ± 5% compared to removal; however, there was no difference between incorporation and

Table 3. Soil sorptivity, aggregate tensile strength, penetration resistance and macroaggregate stability sampled approximately 6 and 18 months after mulch residues (a starch-polyester bioplastic [STP] or a polylactic acid biofabric with embedded wood particles [PLA]) were removed or incorporated in soil in Lincoln, NE

| | Lincoln | | | | | | | |
|---------------------------------|---------------------------------------|------------------------------|------------------------------------|----------------------------|---------------------------------------|------------------------------|------------------------------------|----------------------------|
| | 6 months | | | | 18 months | | | |
| | Sorptivity (cm/s ^{-1/2}) | Tensile strength (kPa) | Penetration resistance (MPa) | Aggregate stability (%) | Sorptivity (cm/s ^{-1/2}) | Tensile strength (kPa) | Penetration resistance (MPa) | Aggregate stability (%) |
| <i>STP</i> | | | | | | | | |
| Removed | | | | | | | | |
| Compost | – | 132.8 | 0.71 | 45.9 | – | 121.5 | 2.5 | – |
| None | 0.20 | 139.6 | 0.71 | 52.2 | 0.12 | 137.7 | 2.6 | 38.0 |
| Incorporated | | | | | | | | |
| Compost | – | 126.1 | 0.78 | 47.6 | – | 105.8 | 2.6 | – |
| None | 0.21 | 128.5 | 0.68 | 45.5 | 0.13 | 136.4 | 2.5 | 50.0 |
| <i>PLA</i> | | | | | | | | |
| Removed | | | | | | | | |
| Compost | – | 140.8 | 0.52 | 49.3 | – | 135.3 | 2.8 | – |
| None | 0.21 | 117.6 | 0.70 | 47.0 | 0.11 | 191.5 | 2.6 | 44.3 |
| Incorporated | | | | | | | | |
| Compost | – | 111.5 | 0.64 | 51.4 | – | 148.4 | 2.7 | – |
| None | 0.17 | 105.4 | 0.71 | 46.0 | 0.13 | 177.2 | 2.6 | 39.1 |
| Standard error | 0.02 | 21.1 | 0.07 | 4.0 | 0.01 | 44.4 | 0.2 | 5.4 |
| <i>Significance</i> | | | | | | | | |
| Mulch | ns | ns | ns | ns | ns | ns | ns | ns |
| Incorporation | ns | ns | ns | ns | ns | ns | ns | ns |
| Mulch × incorporation | ns | ns | ns | ns | ns | ns | ns | ns |
| Compost | – | ns | ns | ns | – | * | ns | – |
| Mulch × compost | – | ns | ns | ns | – | ns | ns | – |
| Incorporation × compost | – | ns | ns | ns | – | ns | ns | – |
| Mulch × incorporation × compost | – | ns | ns | ns | – | ns | ns | – |

Results of analysis of variance for effects of mulch (STP or PLA), incorporation (removed or incorporated), and compost for each soil property are included at the bottom of the table and significance of each factor is indicated as: 'ns' = not significant; * = $P < 0.05$; ** = $P < 0.01$, *** = $P < 0.001$.

removal when the soil was amended with compost (Fig. 2). Sweet corn yield at LNK was not different among treatments. Cabbage yield in 2019 was improved by compost at LNK, not SBF, but mulch type and incorporation had no effects on yield at either location (Table 5).

The soil nitrate results at SBF at 6 months after mulch incorporation provide the most likely explanation for the sweet corn yield differences in 2018 at SBF. Mulch incorporation, particularly PLA residues, immobilized soil nitrate at SBF due to the large mass of carbon-rich substrate added to an already low-nitrogen soil (Table 2). However, the addition of nitrogen from the compost soil amendment seemed to alleviate the nitrogen-limiting conditions caused by the incorporation of mulch residues. The lack of negative yield effects from BDM residues on cabbage in 2019 at SBF suggests nitrate was no longer limiting in residue incorporated plots, which is consistent with the data (Table 2).

Results from some greenhouse-scale, pot studies suggest that BDM residues in the soil can inhibit crop root growth and yield (Qi *et al.*, 2018; Samuelson *et al.*, 2019), but these effects are less commonly observed under field conditions (Hu *et al.*, 2020). Residues of PE, which are more persistent than most BDMs, are more likely to drive reductions in crop yields (Gao *et al.*, 2021). While polylactic acid is persistent in soil like PE, the effects of polylactic acid on soil chemical and physical properties are different, which in turn seems to influence crop yield differently. Any observed yield reductions from PE or BDM residue are often attributed to reduced soil quality, including reduced porosity and water use efficiency (Hu *et al.*, 2020; Gao *et al.*, 2021). However, the only yield reduction detected in this study was likely driven by nitrogen immobilization. While similar research was designed to study the impacts of repeated mulch use and soil incorporation on soil properties and crop yield

Table 4. Soil sorptivity, aggregate tensile strength, penetration resistance and macroaggregate stability sampled approximately 6 and 18 months after mulch residues (a starch-polyester bioplastic [STP] or a polylactic acid biofabric with embedded wood particles [PLA]) were removed or incorporated in soil in Scottsbluff, NE.

| | Scottsbluff | | | | | | | |
|---------------------------------|---------------------------------------|------------------------------|------------------------------------|----------------------------|---------------------------------------|------------------------------|------------------------------------|----------------------------|
| | 6 months | | | | 18 months | | | |
| | Sorptivity (cm/s ^{-1/2}) | Tensile strength (kPa) | Penetration resistance (MPa) | Aggregate stability (%) | Sorptivity (cm/s ^{-1/2}) | Tensile strength (kPa) | Penetration resistance (MPa) | Aggregate stability (%) |
| <i>STP</i> | | | | | | | | |
| Removed | | | | | | | | |
| Compost | – | 63.3 | 0.68 | 5.3 | – | 218.3 | 1.4 | – |
| None | 0.095 | 62.5 | 0.80 | 4.0 | 0.097 | 160.9 | 1.5 | 8.1 |
| Incorporated | | | | | | | | |
| Compost | – | 47.7 | 0.71 | 4.4 | – | 108.9 | 1.4 | – |
| None | 0.085 | 62.7 | 0.81 | 5.2 | 0.097 | 137.8 | 1.6 | 8.6 |
| <i>PLA</i> | | | | | | | | |
| Removed | | | | | | | | |
| Compost | – | 65.2 | 0.86 | 4.3 | – | 158.9 | 1.4 | – |
| None | 0.083 | 71.8 | 0.69 | 6.3 | 0.093 | 166.3 | 1.5 | 8.6 |
| Incorporated | | | | | | | | |
| Compost | – | 74.4 | 0.74 | 4.3 | – | 137.0 | 1.6 | – |
| None | 0.092 | 71.8 | 0.74 | 4.0 | 0.083 | 151.3 | 1.6 | 11.4 |
| Standard error | 0.0059 | 10.5 | 0.07 | 1.2 | 0.0088 | 39.9 | 0.2 | 1.6 |
| <i>Significance</i> | | | | | | | | |
| Mulch | ns | ns | ns | ns | ns | ns | ns | ns |
| Incorporation | ns | ns | ns | ns | ns | ns | ns | ns |
| Mulch × incorporation | ns | ns | ns | ns | ns | ns | ns | ns |
| Compost | – | ns | ns | ns | – | ns | ns | – |
| Mulch × compost | – | ns | * | ns | – | ns | ns | – |
| Incorporation × compost | – | ns | ns | ns | – | ns | ns | – |
| Mulch × incorporation × compost | – | ns | ns | ns | – | ns | ns | – |

Results of analysis of variance for effects of mulch (STP or PLA), incorporation (removed or incorporated), and compost for each soil property are included at the bottom of the table and significance of each factor is indicated as: 'ns' = not significant; * = $P < 0.05$; ** = $P < 0.01$, *** = $P < 0.001$.

(Moore and Wszelaki, 2019; Ghimire *et al.*, 2020; Sintim *et al.*, 2021), the unique value of this study is that management of the subsequent crops was identical within locations and years (i.e., no continued or additional treatments were imposed and tested in the original experimental area). Thus, estimates of yield were not confounded by continued use of the mulches and differences (or lack thereof) can be attributed to the BDM residues alone.

The BDM residues in this study had relatively few impacts on soil chemical or physical properties for two years following incorporation in soil, which is consistent with Sintim *et al.* (2021). However, yield loss in a low fertility soil environment (i.e., SBF, no compost) in the season immediately following incorporation of BDM residue in soil (regardless of mulch type) should inform future BDM residue management. Specifically,

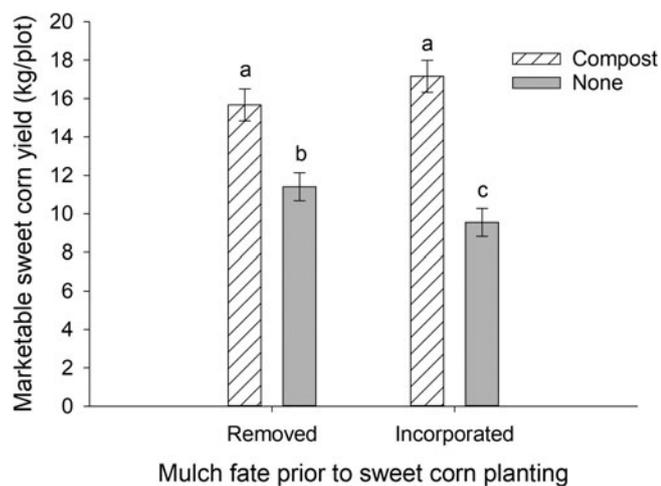
growers should manage BDM residues like other carbon-rich crop residues and supplement with nitrogen fertilizer to mitigate potentially negative effects of nitrogen immobilization (Gao *et al.*, 2009).

Overall, we would not recommend the annual incorporation of the PLA mulch in soil. The potential for soil nitrogen immobilization and reduced yield combined with the high initial mass (1744 kg ha⁻¹) and persistence in soil (29 to 33% initial mass remaining after 22 months in soil; Samuelson *et al.*, 2022) will lead to the accumulation of residues at unsustainable rates that could eventually degrade soil productivity and disrupt agronomic logistics like planting. In contrast, the STP mulch degrades more readily in soil and is unlikely to accumulate at unsustainable rates (Samuelson *et al.*, 2022; Sintim *et al.*, 2021); thus, we recommend the use

Table 5. Marketable yield of sweet corn (2018) and cabbage (2019) harvested at Lincoln or Scottsbluff, NE after mulch residues (a starch-polyester bioplastic [STP] or a polylactic acid biofabric with embedded wood particles [PLA]) were removed or incorporated in soil in 2017.

| | 2018 | | 2019 | |
|---------------------------------|--|-------------|---|-------------|
| | Marketable sweet corn yield (kg plot ⁻¹) | | Marketable cabbage yield (kg plot ⁻¹) | |
| | Lincoln | Scottsbluff | Lincoln | Scottsbluff |
| <i>STP</i> | | | | |
| Removed | | | | |
| Compost | 17.2 | 15.4 | 8.4 | 3.5 |
| None | 18.7 | 11.4 | 7.8 | 4.2 |
| Incorporated | | | | |
| Compost | 20.4 | 17.3 | 10.4 | 4.7 |
| None | 19.4 | 8.8 | 8.0 | 3.7 |
| <i>PLA</i> | | | | |
| Removed | | | | |
| Compost | 20.3 | 16.0 | 11.4 | 5.0 |
| None | 18.1 | 11.5 | 8.3 | 4.3 |
| Incorporated | | | | |
| Compost | 18.0 | 17.0 | 11.2 | 3.4 |
| None | 17.7 | 10.3 | 8.7 | 3.6 |
| <i>Standard error</i> | 1.3 | 1.0 | 1.4 | 0.7 |
| <i>Significance</i> | | | | |
| Mulch | ns | ns | ns | ns |
| Incorporation | ns | ns | ns | ns |
| Mulch × incorporation | ns | ns | ns | ns |
| Compost | ns | *** | * | ns |
| Mulch × compost | ns | ns | ns | ns |
| Incorporation × compost | ns | ** | ns | ns |
| Mulch × incorporation × compost | ns | ns | ns | ns |

Results of analysis of variance for effects of mulch (STP or PLA), incorporation (removed or incorporated), and compost for each soil property are included at the bottom of the table and significance of each factor is indicated as: 'ns' = not significant; * = $P < 0.05$; ** = $P < 0.01$, *** = $P < 0.001$.

**Fig. 2.** Marketable sweet corn yield (kg plot⁻¹) at Scottsbluff, NE in 2018 as influenced by mulch fate in the previous season (removal or incorporation in soil) and compost soil amendment. Error bars represent \pm one standard error of the means and different letters above columns indicate significant differences among treatments ($\alpha = 0.05$).

and incorporation of STP residues except in particularly low fertility soils. The soil in this study was not mechanically disturbed again following the incorporation of BDM residues in 2017, whereas most vegetable growers (especially those using PE and BDM) will till soils at least one time per year in preparation for planting. Tillage may facilitate greater mechanical shredding and biodegradation (Ghimire *et al.*, 2020) of STP residues than observed here (Samuelson *et al.*, 2022); and faster degradation may help to mitigate any potentially negative effects of BDM residues on soil nutrient availability or yield.

While not suitable for biodegradation in soil, the experimental PLA mulch studied here has promising potential for weed suppression in high-density plantings of carrots (*Daucus carota* subsp. *sativus*), spinach (*Spinacia oleracea*), lettuce (*Lactuca sativa*) and strawberry (*Fragaria × ananassa*) (Tofanelli *et al.*, 2021) and in establishing fruit tree orchards. In these production scenarios, PLA could likely be used for at least two growing seasons (personal observations) and then removed from the field and composted at temperatures greater than 60°C and adequate

moisture (Chamas *et al.*, 2020). As with any agricultural mulch, complete removal from the field is impractical, and some residues will remain. However, the results of this study, where the entire mulch was incorporated in soil, suggest that any smaller proportion of PLA residues left after field removal (approximately 11%; He *et al.*, 2018) are unlikely to have deleterious effects on soil quality or subsequent crop yield.

While PLA is persistent in soil, Thompson *et al.* (2019) demonstrated depolymerization and reduced molecular weight of polylactic acid in soil over time, even as the mass loss of mulch slowed. Moreover, depolymerization was accelerated by the addition of plant residues in the mulch matrix like the wood particles included in the PLA mulch of this study. The PLA residues will not degrade fast enough to be certified as biodegradable in soil but are unlikely to persist as long as most PE residues (Chamas *et al.*, 2020). However, further research is needed to determine the long-term fate of PLA residues in soil and their impacts on soil health, particularly when repeated applications are made in the same field where the accumulation of residues beyond the levels studied here is possible.

Conclusions

Biodegradable and biobased mulches are a potentially sustainable alternative to PE because they can be incorporated into the soil or removed and composted at the end of their useful life. While biodegradation in soil or at a commercial composting facility is a preferred environmental outcome compared to burning or landfilling PE, the results of this study suggest that incorporating BDM residues in soil carries some short-term risk to soil productivity. In the season immediately following initial BDM use, sweet corn yield was reduced by 16% when BDM residues were incorporated instead of removed from the soil. However, this negative effect was only observed at SBF in the absence of compost soil amendment where SOM (1.5%) and pre-plant soil nitrate (3.4 mg kg^{-1}) were relatively low. Cabbage yields were not different in the second year following BDM use at SBF and neither sweet corn nor cabbage yields were affected by mulch use or incorporation at LNK. Incorporation of BDM residues may immobilize nitrogen in the short-term in low fertility soils, but no additional negative effects of BDM residues on soil chemical and physical properties were observed. Results contribute to a growing body of evidence that together suggests biobased and biodegradable mulch residue can be safely incorporated in the soil so long as growers take steps to minimize the potential for nitrogen immobilization.

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Conflict of interest. Author I. Kadoma is employed at 3M Company. There are no other competing interests to declare.

References

- Amoakwah E, Frimpong KA and Arthur E (2017) Corn Cob biochar improves aggregate characteristics of a tropical sandy loam. *Soil Science Society of America Journal* **81**, 1054–1063.
- Ashton K, Holmes L and Turner A (2010) Association of metals with plastic production pellets in the marine environment. *Marine Pollution Bulletin* **60**, 2050–2055.
- Bandopadhyay S, Martin-Closas L, Pelacho AM and DeBruyn JM (2018) Biodegradable plastic mulch films: impacts on soil microbial communities and ecosystem functions. *Frontiers in Microbiology* **9**, 1–7. <https://doi.org/10.3389/fmicb.2018.00819>
- Beare MH, Parmelee RW, Hendrix PF, Cheng W, Coleman DC and Crossley Jr. DA (1992) Microbial and faunal interactions and effects on litter nitrogen and decomposition in agroecosystems. *Ecological Monographs* **62**, 569–591.
- Bossuyt H, Deneff K, Six J, Frey SD, Merckx R and Paustian K (2001) Influence of microbial populations and residue quality on aggregate stability. *Applied Soil Ecology* **16**, 195–208.
- Briassoulis D, Babou E, Hiskakis M and Kyrikou I (2015) Analysis of long-term degradation behaviour of polyethylene mulching films with pro-oxidants under real cultivation and soil burial conditions. *Environmental Science and Pollution Research International* **22**, 2584–2598.
- Carrizo ME, Alesso CA, Soares Franco HH, Bernabé Ferreira CJ and Imhoff S (2018) Tensile strength of mollisols of contrasting texture under influence of plant growth and crop residues addition. *Geoderma* **329**, 1–10.
- Chamas A, Moon H, Zheng J, Qiu Y, Tabassum T, Jang JH, Abu-Omar M, Scott SL and Suh S (2020) Degradation rates of plastics in the environment. *ACS Sustainable Chemistry & Engineering* **8**, 3494–3511.
- de Souza Machado AA, Lau CW, Till J, Kloas W, Lehmann A, Becker R and Rillig MC (2018) Impacts of microplastics on the soil biophysical environment. *Environmental Science & Technology* **52**, 9656–9665.
- Dexter AR and Kroesbergen B (1985) Methodology for determination of tensile strength of soil aggregates. *Journal of Agricultural Engineering Research* **31**, 139–147.
- Dharmalingam S, Hayes DG, Wadsworth LC and Dunlap RN (2016) Analysis of the time course of degradation for fully biobased nonwoven agricultural mulches in compost-enriched soil. *Textile Research Journal* **86**, 1343–1355.
- Döring TF, Brandt M, Heß J, Finckh MR and Saucke H (2005) Effects of straw mulch on soil nitrate dynamics, weeds, yield and soil erosion in organically grown potatoes. *Field Crops Research* **94**, 238–249.
- Feuilloloy P, César G, Benguigui L, Grohens Y, Pillin I, Bewa H, Lefaux S and Jamal M (2005) Degradation of polyethylene designed for agricultural purposes. *Journal of Polymers and the Environment* **13**, 349–355.
- Gao Y, Li Y, Zhang J, Liu W, Dang Z, Cao W and Qiang Q (2009) Effects of mulch, N fertilizer, and plant density on wheat yield, wheat nitrogen uptake, and residual soil nitrate in a dryland area of China. *Nutrient Cycling in Agroecosystems* **85**, 109–121.
- Gao X, Xie D and Yang C (2021) Effects of a PLA/PBAT biodegradable film mulch as a replacement of polyethylene film and their residues on crop and soil environment. *Agricultural Water Management* **255**, 107053.
- Ghimire S, Saxton AM, Wszelaki AL, Moore JC and Miles CA (2017) Reliability of soil sampling method to assess visible biodegradable mulch fragments remaining in the field after soil incorporation. *HortTechnology* **27**, 650–658.
- Ghimire S, Scheenstra E and Miles CA (2020) Soil-biodegradable mulches for growth, yield, and quality of sweet corn in a Mediterranean-type climate. *HortScience* **55**, 317–325.
- Goldberger JR, Jones RE, Miles CA, Wallace RW and Inglis DA (2015) Barriers and bridges to the adoption of biodegradable plastic mulches for US specialty crop production. *Renewable Agriculture and Food Systems* **30**, 143–153.
- Greenly KM and Rakow DA (1995) The effect of wood mulch type and depth on weed and tree growth and certain soil parameters. *Journal of Arboriculture* **21**, 225–232.
- Hayes DG, Wadsworth LC, Sintim HY, Flury M, English M, Schaeffer S and Saxton AM (2017) Effect of diverse weathering conditions on the physico-chemical properties of biodegradable plastic mulches. *Polymer Testing* **62**, 454–467.
- He G, Wang Z, Li S and Malhi SS (2018) Plastic mulch: tradeoffs between productivity and greenhouse gas emissions. *Journal of Cleaner Production* **172**, 1311–1318.
- Hoagland L, Carpenter-Boggs L, Granatstein D, Mazzola M, Smith J, Peryea F and Reganold JP (2008) Orchard floor management effects on

- nitrogen fertility and soil biological activity in a newly established organic apple orchard. *Biology and Fertility of Soils* **45**, 11.
- Hu Q, Li X, Gonçalves JM, Shi H, Tian T and Chen N (2020) Effects of residual plastic-film mulch on field corn growth and productivity. *Science of The Total Environment* **729**, 138901.
- Irmak S, Payero JO, VanDeWalle B, Rees J, Zoubek G, Martin DL, Kranz WL, Eisenhauer DE and Leininger D (2016) Principles and operational characteristics of Watermark granular matrix sensor to measure soil water status and its practical applications for irrigation management in various soil textures, University of Nebraska Extension EC783. <https://extensionpublications.unl.edu/assets/pdf/ec783.pdf>.
- Janczak K, Hryniewicz K, Znajewska Z and Dąbrowska G (2018) Use of rhizosphere microorganisms in the biodegradation of PLA and PET polymers in compost soil. *International Biodeterioration & Biodegradation* **130**, 65–75.
- Jiang XJ, Liu W, Wang E, Zhou T and Xin P (2017) Residual plastic mulch fragments effects on soil physical properties and water flow behavior in the minqin oasis, northwestern China. *Soil and Tillage Research* **166**, 100–107.
- Karamanlioglu M and Robson GD (2013) The influence of biotic and abiotic factors on the rate of degradation of poly(lactic acid) (PLA) coupons buried in compost and soil. *Polymer Degradation and Stability* **98**, 2063–2071.
- Kasirajan S and Ngouajio M (2012) Polyethylene and biodegradable mulches for agricultural applications: a review. *Agronomy for Sustainable Development* **32**, 501–529.
- Li C, Moore-Kucera J, Miles C, Leonas K, Lee J, Corbin A and Inglis D (2014a) Degradation of potentially biodegradable plastic mulch films at three diverse U.S. locations. *Agroecology and Sustainable Food Systems* **38**, 861–889.
- Li C, Moore-Kucera J, Lee J, Corbin A, Brodhagen M, Miles C and Inglis D (2014b) Effects of biodegradable mulch on soil quality. *Applied Soil Ecology* **79**, 59–69.
- Li Z, Schneider RL, Morreale SJ, Xie Y, Li C and Li J (2018) Woody organic amendments for retaining soil water, improving soil properties and enhancing plant growth in desertified soils of ningxia, China. *Geoderma* **310**, 143–152.
- Liu Y, Huang Q, Hu W, Qin J, Zheng Y, Wang J, Wang Q, Xu Y, Guo G, Hu S and Xu L (2021) Effects of plastic mulch film residues on soil-microbe-plant systems under different soil pH conditions. *Chemosphere* **267**, 128901.
- Masunga RH, Uzokwe VN, Mlay PD, Odeh I, Singh A, Buchan D and De Neve S (2016) Nitrogen mineralization dynamics of different valuable organic amendments commonly used in agriculture. *Applied Soil Ecology* **101**, 185–193.
- Miles C, DeVetter L, Ghimire S and Hayes DG (2017) Suitability of biodegradable plastic mulches for organic and sustainable agricultural production systems. *HortScience* **52**, 10–15.
- Minasny B and Cook F (2011) Sorptivity of soils. In Gliński J, Horabik J and Lipiec J (eds), *Encyclopedia of Agrophysics*. Dordrecht: Springer Netherlands, pp. 824–826.
- Moore JC and Wszelaki AL (2019) The use of biodegradable mulches in pepper production in the southeastern United States. *HortScience* **54**, 1031–1038.
- Mooshammer M, Wanek W, Hämmerle I, Fuchslueger L, Hofhansl F, Knoltsch A, Schneckner J, Takriti M, Watzka M, Wild B, Keiblinger KM, Zechmeister-Boltenstern S and Richter A (2014) Adjustment of microbial nitrogen use efficiency to carbon:nitrogen imbalances regulates soil nitrogen cycling. *Nature Communications* **5**, 3694.
- Murphy BW and Murphy BW (2015) Impact of soil organic matter on soil properties—a review with emphasis on Australian soils. *Soil Research* **53**, 605–635.
- Nimmo JR and Perkins KS (2002) 2.6 Aggregate stability and size distribution. In Dane JH and Topp GC (eds), *Methods of Soil Analysis*. Hoboken, New Jersey, USA: John Wiley & Sons, Ltd., pp. 317–328.
- Osipitan OA, Dille JA, Assefa Y, Radicetti E, Ayeni A and Knezevic SZ (2019) Impact of cover crop management on level of weed suppression: a meta-analysis. *Crop Science* **59**, 833–842.
- Öztaş T, Canpolat MY and Sönmez K (1999) Strength of individual soil aggregates against crushing forces II. Influence of selected soil properties. *Turkish Journal of Agriculture and Forestry* **23**, 573–577.
- Powelson DS, Riche AB, Coleman K, Glendining MJ and Whitmore AP (2008) Carbon sequestration in European soils through straw incorporation: limitations and alternatives. *Waste Management* **28**, 741–746.
- Qi Y, Yang X, Pelaez AM, Huerta Lwanga E, Beriot N, Gertsen H, Garbeva P and Geissen V (2018) Macro- and micro- plastics in soil-plant system: effects of plastic mulch film residues on wheat (*Triticum aestivum*) growth. *Science of the Total Environment* **645**, 1048–1056.
- Qi Y, Beriot N, Gort G, Huerta Lwanga E, Gooren H, Yang X and Geissen V (2020a) Impact of plastic mulch film debris on soil physicochemical and hydrological properties. *Environmental Pollution* **266**, 115097.
- Qi Y, Ossowicki A, Yang X, Huerta Lwanga E, Dini-Andreote F, Geissen V and Garbeva P (2020b) Effects of plastic mulch film residues on wheat rhizosphere and soil properties. *Journal of Hazardous Materials* **387**, 121711.
- Qi J, Jensen JL, Christensen BT and Munkholm LJ (2022) Soil structural stability following decades of straw incorporation and use of ryegrass cover crops. *Geoderma* **406**, 115463.
- Rakkar MK, Blanco-Canqui H, Drijber RA, Drewnoski ME, MacDonald JC and Klopfenstein T (2017) Impacts of cattle grazing of corn residues on soil properties after 16 years. *Soil Science Society of America Journal* **81**, 414–424.
- Reichel R, Wei J, Islam MS, Schmid C, Wissel H, Schröder P, Schloter M and Brüggemann N (2018) Potential of wheat straw, spruce sawdust, and lignin as high organic carbon soil amendments to improve agricultural nitrogen retention capacity: an incubation study. *Frontiers in Plant Science* **28**, 1–13. <https://doi.org/10.3389/fpls.2018.00900>
- Samuelson MB (2019) *Microbial Response to Biodegradable Mulch: Can Degradation Rate Be Accelerated by Management?* (MS thesis). University of Nebraska, Lincoln.
- Samuelson MB, Drijber R and Wortman SE (2019) Assessing the quality and possible functions of compost extracts in organic systems. *HortScience* **54**, S29–S30.
- Samuelson MB, Reid EV, Drijber R, Jeske E, Blanco-Canqui H, Mamo M, Kadoma I and Wortman SE (2022) Effects of compost, cover crops, and local conditions on degradation of two agricultural mulches in soil. *Renewable Agriculture and Food Systems* **37**, 128–141.
- Sintim HY, Bandopadhyay S, English ME, Bary A, Liquey y González JE, DeBruyn JM, Schaeffer SM, Miles CA and Flury M (2021) Four years of continuous use of soil-biodegradable plastic mulch: impact on soil and groundwater quality. *Geoderma* **381**, 114665.
- Six J, Paustian K, Elliott ET and Combrink C (2000) Soil structure and organic matter I. distribution of aggregate-size classes and aggregate-associated carbon. *Soil Science Society of America Journal* **64**, 681–689.
- Smith RE (1999) Technical note: rapid measurement of soil sorptivity. *Soil Science Society of America Journal* **63**, 55–57.
- Tahboub MB, Lindemann WC and Murray L (2008) Chemical and physical properties of soil amended with pecan wood chips. *HortScience* **43**, 891–896.
- Thompson AA, Samuelson MB, Kadoma I, Soto-Cantu E, Drijber R and Wortman SE (2019) Degradation rate of Bio-based agricultural mulch is influenced by mulch composition and biostimulant application. *Journal of Polymers and the Environment* **27**, 498–509.
- Tofanelli MBD and Wortman SE (2020) Benchmarking the agronomic performance of biodegradable mulches against polyethylene mulch film: a meta-analysis. *Agronomy* **10**, 1618.
- Tofanelli MBD, Kadoma I and Wortman SE (2021) Proof of concept for growing lettuce and carrot in a biobased mulch membrane. *Renewable Agriculture and Food Systems* **36**, 121–125. doi: doi:10.1017/S1742170520000162
- Wang J, Luo Y, Teng Y, Ma W, Christie P and Li Z (2013) Soil contamination by phthalate esters in Chinese intensive vegetable production systems with different modes of use of plastic film. *Environmental Pollution* **180**, 265–273.
- Ward RC (2021) *Ward Guide: Guiding Producers Today to Feed the World Tomorrow*. Kearney, Nebraska, USA: Ward Laboratories, Inc., Available at <https://www.wardlab.com/wp-content/uploads/2021/03/WARDGUIDE-Master-Updated-3.25.21.pdf>.

- Wortman SE, Kadoma I and Crandall MD** (2015) Assessing the potential for spunbond, nonwoven biodegradable fabric as mulches for tomato and bell pepper crops. *Scientia Horticulturae* **193**, 209–217.
- Wortman SE, Holmes AA, Miernicki E, Knoche K and Pittelkow CM** (2017) First-season crop yield response to organic soil amendments: a meta-analysis. *Agronomy Journal* **109**, 1210–1217.
- Yu Y, Griffin-LaHue DE, Miles CA, Hayes DG and Flury M** (2021) Are micro- and nanoplastics from soil-biodegradable plastic mulches an environmental concern? *Journal of Hazardous Materials Advances* **4**, 100024.
- Zhou J, Wen Y, Marshall MR, Zhao J, Gui H, Yang Y, Zeng Z, Jones DL and Zang H** (2021) Microplastics as an emerging threat to plant and soil health in agroecosystems. *Science of The Total Environment* **787**, 147444.