

## Research Article

<sup>†</sup>These authors have contributed equally to this paper and share first authorship.

**Cite this article:** Valenzuela-Moreno LF et al. (2025) New genetic variants of *Toxoplasma gondii* isolates and a novel *ROP5* allele in free-range chickens from Tabasco, México. *Parasitology*, 1–9. <https://doi.org/10.1017/S0031182025100401>

Received: 1 March 2025

Revised: 16 May 2025

Accepted: 5 June 2025


### Keywords:

free-range chickens (*Gallus gallus domesticus*); genotyping; isolation; México; *Toxoplasma gondii*; virulence prediction

### Corresponding author:

Heriberto Caballero-Ortega;  
Email: [hcaballero\\_2000@yahoo.com.mx](mailto:hcaballero_2000@yahoo.com.mx)

# New genetic variants of *Toxoplasma gondii* isolates and a novel *ROP5* allele in free-range chickens from Tabasco, México

Luis Fernando Valenzuela-Moreno<sup>1,†</sup>, Carlos Cedillo-Peláez<sup>1,†</sup>,  
Claudia Patricia Rico-Torres<sup>1</sup>, Lluvia Guadalupe Moreno-Pérez<sup>2</sup>,  
Claudia Virginia Zaragoza-Vera<sup>2</sup>, Luz Belinda Ortiz-Alegria<sup>1</sup>,  
Irma Cañedo-Solares<sup>1</sup>, Lizbeth Xicoténcatl-García<sup>3</sup>, Fernando García-Lacy<sup>4</sup> and  
Heriberto Caballero-Ortega<sup>1,5</sup> 

<sup>1</sup>Laboratorio de Inmunología Experimental, Subdirección de Medicina Experimental, Instituto Nacional de Pediatría, CDMX, México, México; <sup>2</sup>División Académica de Ciencias Agropecuarias, Universidad Juárez Autónoma de Tabasco, Tabasco, México; <sup>3</sup>Bioterio, Subdirección de Medicina Experimental, Instituto Nacional de Pediatría, CDMX, México, México; <sup>4</sup>Departamento de Medicina, Cirugía y Zootecnia para Equinos, FMVZ, UNAM, CDMX, México, México and <sup>5</sup>Programa de Doctorado en Ciencias de la Producción y de la Salud Animal, Facultad de Medicina Veterinaria y Zootecnia, Universidad Nacional Autónoma de México, México, México

## Abstract

*Toxoplasma gondii* is a protozoan parasite that causes infection in birds and mammals (terrestrial and marine), both domestic and wild. The state of Tabasco has favourable climatic and ecological conditions for the replication and dissemination of this parasite. Therefore, the aim of this study was to isolate and genotype *T. gondii* from free-range chickens in this region of México by PCR-RFLP of 15 genetic markers. A total of 12 chickens were obtained from 7 municipalities. Serological survey by the modified agglutination test (MAT) of chicken serum samples revealed that 9 out of the 12 chickens (75%) tested had antibodies against *T. gondii* (titres  $\geq 1:10$ ). *Toxoplasma gondii* DNA was detected by PCR in tissues from 8 out of the 12 chickens. Twelve viable strains of *T. gondii* were isolated from the heart and brain samples of eight chickens by bioassay in mice. Genetic characterization of tachyzoite-derived DNA was performed using 10 multilocus RFLP-PCR genotyping markers (*SAG1*, *SAG2*, *SAG3*, *BTUB*, *GRA6*, *c22-8*, *c29-2*, *L358*, *PK1* and *Apico*) and five virulence-related markers (*CS3*, *ROP5*, *ROP16*, *ROP17* and *ROP18*). A total of 6 ToxoDB PCR-RFLP genotypes were identified, including #8 (also known as Type BrIII), #28, #38, and 3 new genotypes designated as #344, #345 and #346. Combination of *ROP18/ROP5* alleles were 1/3, 3/3 and 4/3. ToxoDB #344 and #345 genotypes also had a new allele at the *ROP5* locus. These results suggest high genetic diversity of *T. gondii* in southeastern México.

## Introduction

Toxoplasmosis is a zoonotic disease caused by *Toxoplasma gondii*, an obligate intracellular parasite that infects a wide variety of homeothermic animals, including humans. It can cause spontaneous abortions, stillbirths and congenital malformations in both humans and several animal species (Buxton, 1990; Tenter et al., 2000). It is estimated that approximately 30–50% of the world's population is chronically infected by this protozoan, which is more common in warm and humid areas (Montoya and Liesenfeld, 2004; Dubey, 2010; Flegr et al., 2014). In México, approximately 44% of the human population is seropositive, with the Gulf and Pacific coasts being the southeastern regions with the highest seroprevalence of *T. gondii* infection (Caballero-Ortega et al., 2012). The Mexican southeast is a tropical region made up of warm-humid states such as Quintana Roo, Yucatán, Campeche, Chiapas and Tabasco, where the climatic and biotic conditions required for the perpetuation of *T. gondii* are ideal, along with the presence of wild felines, which benefit the spread and high genetic variability of the parasite (Comisión Nacional para el Conocimiento y Uso de la Biodiversidad (CONABIO) y Gobierno del Estado de Tabasco, 2021). Sentinel animals such as stray dogs, feral cats and free-range chickens are good indicators of the presence of this protozoan in the environment because they can consume water and food contaminated with *T. gondii* oocysts. Furthermore, most of the *T. gondii* isolates and genotypes described in the literature have been obtained from these animal species (Shwab et al., 2014; Calero-Bernal et al., 2022).

In addition to archetypal genotypes I, II and III, 343 different genotypes obtained from various animal species worldwide, including humans, are grouped into 6 different clades (Lorenzi et al., 2016; Gennari et al., 2024). Therefore, in areas with a high population density of definitive

hosts (wild, feral or domestic cats), a wide variety of genotypes is expected due to sexual genetic recombination. We looked for the circulating genotypes of this parasite in the southeastern region of México because its genetic characteristics can directly influence the clinical presentation and severity of toxoplasmosis. We obtained *T. gondii* isolates (ToxoDB #3, #8, #10, #28, #38, #116, #164, #182, #225 genotypes) from feral cats from Quintana Roo, stray dogs from Chiapas and stray dogs and free-range chickens from Campeche, as well as several partial genotypes, mixed infections and the existence of up to 18 possible new genotypes of this parasite (Valenzuela-Moreno *et al.*, 2019, 2020; Rico-Torres *et al.*, 2024). These results suggest a considerable presence of different genotypes of *T. gondii*, including nonarchetypal strains that could be endemic to this region, and high infective pressure to which several intermediate hosts, including humans, are exposed. In addition to the classic genotyping panel, we have reported the genetic characterization of *CS3*, *ROP16*, *ROP17*, *ROP18* and *ROP5* markers that help predict the virulence of this parasite in mice. These loci play important roles in the parasite's phenotype in murine models, as they modulate the host immune response through the ability to phosphorylate and activate transcription factors during invasion (Weilhammer and Rasley, 2011; Zhang *et al.*, 2019). In particular, *ROP18/ROP5* allele combinations are highly predictive of virulence in a murine model (Gazzinelli *et al.*, 2014; Shwab *et al.*, 2016).

Continuing the genetic mapping of this parasite, we sampled free-range chickens from Tabasco to isolate and genotype *T. gondii* by Mn-PCR-RFLP, using 10 classical polymorphic markers and five specific genetic markers to predict its virulence in a murine model. Tabasco is located in the southeastern México that borders the Gulf of México, Veracruz, Chiapas, Campeche and Guatemala (Comisión Nacional para el Conocimiento y Uso de la Biodiversidad (CONABIO) y Gobierno del Estado de Tabasco, 2021), and to date there are no reports of *T. gondii* genotyping in this state.

## Materials and methods

### Tissue samples from free-range chickens

Twelve chickens were randomly sampled in the municipalities of Centro, Huimanguillo, Jalapa, Jalpa de Méndez, Macuspana, Nacajuca and Teapa in the State of Tabasco, México. The animals were euthanized with an intracelomic overdose of sodium pentobarbital (390 mg/kg, Doletal<sup>®</sup>, Vetoquinol, France) following the guidelines of the Mexican Official Standard NOM-033-ZOO-1995. After confirming the loss of muscle activity and respiratory and cardiac arrest, necropsies were performed and representative sections of tissues (brain, lung, heart, liver and striated muscle) were collected and immersed in 10% buffered formalin (Schuneman and Constantino, 2002). Half of the heart and brain were kept at 4 °C and used for parasite isolation via a mouse bioassay.

### Serology

To determine the serologic status of the free-range chickens, the modified agglutination test (MAT) was standardized according to the methodology described by Dubey and Desmonts (1987) with slight modifications. *Toxoplasma gondii* GT1 tachyzoites were proliferated in NIH/3T3 mouse fibroblasts (ATCC CRL-1658) in 24-well plates (Costar, cat. 3524, Corning, NY, USA) and subsequently fixed in 6% formalin for 72 h, after which 0.2% Evans

blue was added to the alkaline buffer. Sera from all chickens were diluted 1:5 in filtered PBS and serially diluted to 1:40. The MAT assay was carried out twice. Negative and positive serum controls were included in each plate (sera from naive and inoculated with Me49 strain mice, respectively). The sediment at the bottom of the plate was considered negative, and samples without sediment were considered positive.

### Bioassay and isolation of *T. gondii*

Heart and brain samples from each chicken were digested with pepsin to purify and concentrate the *T. gondii* tissue cysts following the methodology described by Dubey (1998). Briefly, the tissues were cut into small pieces, macerated in a mortar with saline solution and digested with acidic pepsin solution in a water bath at 37 °C for 1 h. The homogenate was neutralized with 1.2% sodium bicarbonate (pH 8.3) and then suspended in saline solution with 1000 IU of penicillin and 100 µg/mL streptomycin (Sigma-Merck KGaA, Darmstadt, Germany). Finally, 1 mL of the homogenate from each tissue sample (heart and brain) was aseptically inoculated intraperitoneally into two BALB/c mice (one for each tissue). One week after inoculation, the peritoneal lavage of all of the mice was performed with 2 mL of sterile PBS to obtain tachyzoites. The obtained lavages were seeded on NIH/3T3 mouse fibroblasts (ATCC CRL-1658) in 24-well plates. After 3–7 days, the supernatant was transferred to NIH/373 cells growing at 70% confluency in 25 cm<sup>2</sup> flasks (Falcon, cat. 353014, Corning, NY, USA), cultured in DMEM (ATCC, Washington, DC, USA) supplemented with 10% calf bovine serum (ATCC, Washington, DC, USA) and 1× antibiotic–antimycotic mixture (Gibco, MA, USA), and monitored daily for the appearance of tachyzoites. All of the isolated parasites were proliferated in cell culture and cryopreserved in 90% calf bovine serum (ATCC cat. 30-2020) and 10% dimethyl sulfoxide (DMSO, Sigma-Merck KGaA, cat. D2650, Darmstadt, Germany) in liquid nitrogen at –196 °C until use.

### DNA extraction

The genomic DNA from free tachyzoites collected from cell culture as well as from chicken tissues used to obtain *T. gondii* isolates was extracted using the Qiagen Gentra<sup>®</sup> Puregene<sup>®</sup> Tissue Kit (Hilden, Germany) following the manufacturer's instructions and as previously reported (Cedillo-Peláez, 2009). DNA was quantified with a spectrophotometer (Thermo Scientific Nanodrop<sup>™</sup> 1000, MA, USA) and stored at –20 °C until use.

### *Toxoplasma gondii* PCR, genotyping and parasite load

The DNA of all of the viable parasite-like isolates was analysed for the noncoding repetitive sequence 529RE of *T. gondii* by convectional PCR using previously described primers (Homan *et al.*, 2000). Positive *T. gondii* isolates were processed by multilocus-nested PCR-RFLP (Mn-PCR-RFLP), which amplified 10 polymorphic markers (*SAG1*, 5'3' *SAG2*, Alt. *SAG2*, *SAG3*, *BTUB*, *GRA6*, *c22-8*, *c29-2*, *L358*, *PK1* and *APICO*) to determine the genotype of each isolate and five additional polymorphic markers that predict its virulence in the murine model (*CS3*, *ROP5*, *ROP16*, *ROP17* and *ROP18*) (Pena *et al.*, 2008; Su *et al.*, 2010; Shwab *et al.*, 2016). All of the molecular assays were carried out with AmpliTaq Gold<sup>™</sup> polymerase (Thermo Fisher Scientific, cat.

4311806, MA, USA) as previously described (Su et al., 2010; Rico-Torres et al., 2022). All of the assays included sterile water as a negative control and DNA from reference strains of Types I, II and III (RH, Me49 and VEG, *Toxoplasma gondii* ATCC<sup>®</sup> 50611<sup>TM</sup> and 50861<sup>TM</sup>) as positive controls. Additional negative and positive reamplification controls were included during nested assays to rule out cross-contamination. The PCR products were resolved on 1.5% TBE-agarose gels stained with ethidium bromide (EtBr) and then visualized and digitalized under UV illumination (Bio-DocIt, UVP<sup>™</sup>). RFLP assays were performed with restriction enzymes from New England Biolabs and Thermo Scientific (*Fsp*BI) using 3.5 U of each enzyme as previously described (Pena et al., 2008; Su et al., 2010; Shwab et al., 2016). The products obtained by RFLP were visualized on 3% TBE-agarose gels, stained with EtBr and digitalized. Restriction fragments that showed a different pattern than those of the reference strains (*ROP5* and *ROP17*) were electrophoresed on a 4–15% polyacrylamide gel stained with EtBr to improve the resolution. Parasite load was determined in chicken tissues by TaqMan-based quantitative PCR assays using the *T. gondii* *B1* gene as the genetic target. Standard curves were created by spiking between 50 and  $5 \times 10^6$  tachyzoites of the RH strain into brain and heart samples from uninfected BALB/c mice (Kompalic-Cristo et al., 2007; Cedillo-Peláez et al., 2011; Vargas-Villavicencio et al., 2016). Each sample was analysed in triplicate using a 48-well StepOne thermocycler (Thermo Fisher Scientific, MA, USA). The number of tachyzoites per microgram of tissue was calculated by interpolating the threshold cycle (Ct) value of each tissue sample within the standard curve and the fluorescence data were collected at the end of the PCR process using Step One v2.3 software to obtain the linear regression coefficient ( $R^2$ ) and amplification efficiency ( $\text{Eff}\% \pm 10\%$ ).

## Sequencing

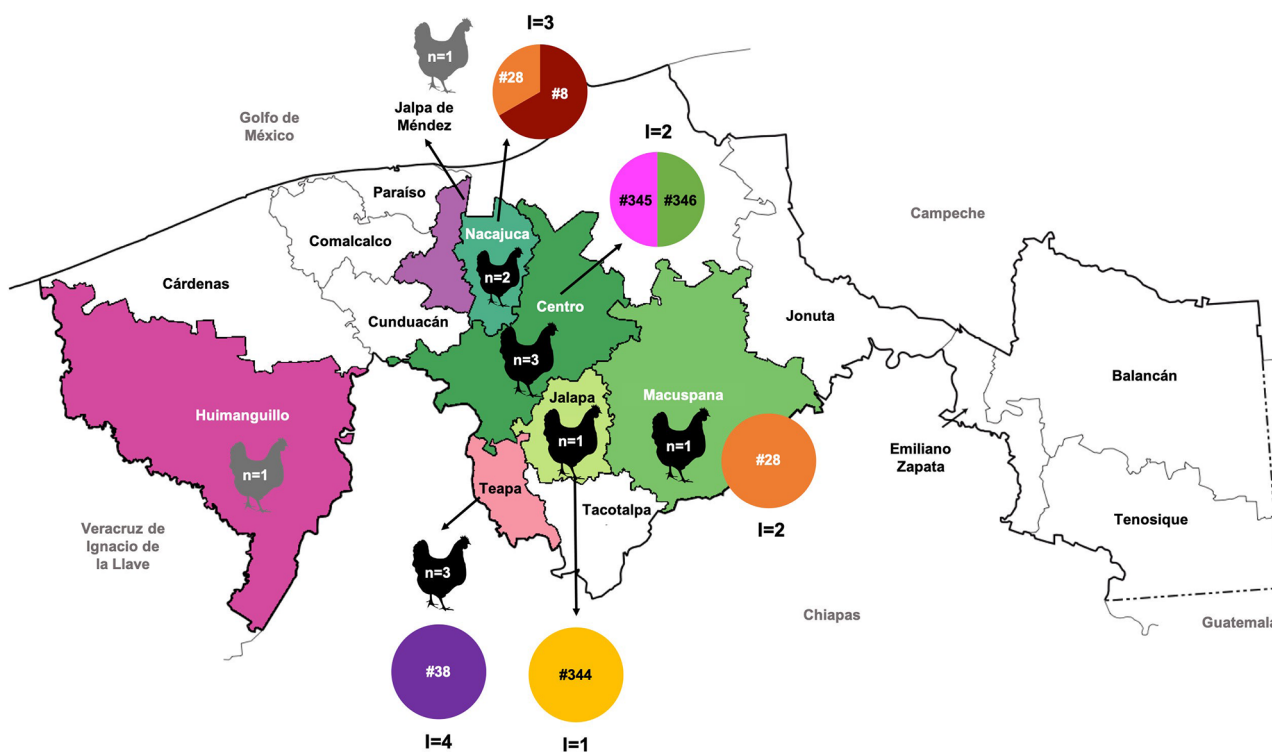
The samples that presented a different RFLP pattern than the RH, Me49 or VEG reference strains were amplified again from the original DNA of each *T. gondii* isolate using internal primers. The amplicons obtained were sequenced both forward and reverse at the Instituto de Biología-UNAM, México, using standard Sanger methodology, with a 3730xl 96-capillary DNA Analyzer (Thermo Fisher Scientific, MA, USA). Chromatograms were analysed with SnapGene viewer<sup>®</sup> v 5.2, and the overall Phred quality was above 30.

## Phylogenetic network

To characterize the evolutionary relationships between the *T. gondii* isolates identified in this study and the representative strains of each haplogroup, a phylogenetic network was constructed. The genotyping marker data were analysed using SplitsTree4 software (Huson, 1998; Huson and Bryant, 2006). To determine the phylogenetic relationships, representative strains of *T. gondii* from each haplogroup were included: RH, GT1 (1); Me49, PTG, PRU (2); VEG, CTG (3); MAS (4); GUY-RUB (5); FOU, Wiktor (6); CAST (7); TgCtBr05 (8); P89 (9); GUY-VAND (10); Cougar (11); Type12, Type 12-X-A (12); Chinese1 (13); GAB2–2007-GAL-DOM2 (14); and TgCtCo05 (15), as well as Mexican strains found in different parts of México, including those isolated in this study.

## Results

Twelve free-range chickens were obtained from 7 municipalities of Tabasco, México (Figure 1). All of the captured animals were adults



**Figure 1.** Sampling of chickens and genotype of *Toxoplasma gondii* isolates in the state of Tabasco, México. The municipalities from which biological samples of free-range chickens were obtained are shown. Grey chickens show individuals from which no isolates were obtained. n: number of chickens included per municipality. I: isolates obtained from chickens included in that municipality.



and ranged from 6 months to 5 years old. During *post mortem* analyses, no lesions compatible with acute or chronic *T. gondii* infection were detected. Nine out of 12 chickens were positive for *T. gondii* according to the MAT test (titres from 1:10 to 1:40; Table 1).

### Isolates, molecular identification and parasite load

Twelve viable isolates were obtained from the tissues of 8 chickens. One isolate was obtained from chickens 2, 6, 7 and 8, while 2 isolates each (one from the heart and one from the brain) were obtained from chickens 1, 3, 4 and 5 (Table 1). The four chickens from which no heart nor brain isolate was obtained were also negative for the presence of *T. gondii* DNA by PCR (data not shown). All of the isolates obtained were positive for the *T. gondii* marker 529RE by conventional PCR and were included in the multilocus nested PCR to obtain their genotype, as well as the genetic characterization of their virulence markers. Among the 12 *T. gondii* isolates, six different ToxoDB genotypes were identified: two #8 Type BrIII, three #28, four #38 and three new genotypes that were identified as #344, #345 and #346. The parasite load was determined in the tissues of origin and was equivalent to < 50 and more than 18 000 tachyzoites/mg of tissue (Table 1). The virulence marker CS3 showed Type I and III alleles, whereas in *ROP16*, *ROP17*, *ROP18* and *ROP5*, the Type 1 alleles were more frequent than Type 3 and these more frequent than Type 4. Interestingly, in the TgCkMxTab6 and TgCkMxTab7 isolates, the *ROP5* restriction pattern was different from that of the archetypal Type I, II and III strains (RH, Me49 and VEG, respectively), as well as five other restriction patterns previously reported for this locus. Three *ROP18/ROP5* virulence patterns were identified in the 12 *T. gondii* isolates: three 4/3 (TgCkMxTab1a, 1b and 2), four 1/3 (TgCkMxTab4a, 4b, 5a and 5b) and three 3/3 (TgCkMxTab3a, 3b and 8). Two restriction patterns (TgCkMxTab6 and 7) could not be associated with virulence because of the new RFLP pattern at the *ROP5* locus (Figure 2 and Table 2). The *ROP17* RFLP digestion products of isolates TgCkMxTab7 and TgCkMxTab8 had a restriction pattern compatible with allele 4, which was confirmed by electrophoresis of the digestion products on polyacrylamide gels (Suppl Figure 1).

### Sequencing

The *ROP17* locus of isolates TgCkMxTab7 and TgCkMxTab8 presented a different restriction pattern from that of the archetypal alleles (RH type 1, Me49 and VEG type 2). Therefore, they were amplified again using the external primers and then sequenced. The obtained sequence was subjected to virtual digestion ([www.benchling.com](http://www.benchling.com)), which replicated the restriction pattern of two known *T. gondii* strains with type 4 alleles of *ROP17*, MAS and TgCatBr5. The sequences obtained from these two isolates were subsequently aligned and compared with the *ROP17* sequences of the archetypal strains GT1, Me49 and VEG and of the non-archetypal strains MAS and TgCatBr5. The two *ROP17* sequences obtained from *T. gondii* strains in chickens had 100% identity with the TgCatBr5 sequence and 99.9% (796/797) identity with the MAS *T. gondii* strain, which has a SNP at position 77 of the alignment (Suppl Figure 2). The obtained and analysed sequences were deposited in GenBank® with accession numbers PP983038 and PP983039.

### Phylogenetic network

The *Toxoplasma gondii* isolates obtained in this study were grouped mainly into Type I and Type III branches or close to them. The isolates TgCkMxTab1a, 1b and 2 shared a branch with other isolates of *T. gondii* genotype #28 obtained in México and with the CAST strain, which is representative of haplogroup 7. TgCkMxTab3a and TgCkMxTab3b were placed on the same branch as a *T. gondii* isolate obtained from a dog from the State of Chiapas (genotype #8) and P89 strain in haplogroup 9, along with another Mexican isolate (TgCatMxHgo1, genotype #48) in the branch of genotypes related to Type III strains. The TgCkMxTab4a, 4b, 5a and 5b isolates clustered with *T. gondii* isolates from the State of Campeche near the TgCatBr05 branch and between haplogroups 3 and 8. Finally, the TgCkMxTab6, 7 and 8 isolates (new genotypes) do not share a branch with any included strain but are close to haplogroups 3, 9 and 14, respectively (Figure 3).

### Discussion

México is located in a climatic transition region between North America and Central and South America. Tabasco is a southeastern state with a tropical climate and favourable conditions for the perpetuation and dissemination of *T. gondii*. Therefore, it is interesting to determine if the circulating genotypes of this region are the same as those described in neighbouring states and other tropical regions of the continent. Free-range chickens are ideal sentinel animals for ground contamination because they feed on soil organic matter and can come in contact with oocysts released by felines (Dubey et al., 2007; Shwab et al., 2014; Kurth et al., 2021). The high frequency of anti-*T. gondii* antibodies in chickens was 75% (9/12) is consistent with the frequency reported in stray dogs from Chiapas and Campeche and in humans from Yucatan Peninsula, which suggests that peridomestic animals are good indicators of the infective pressure of *T. gondii* in the environment (Caballero-Ortega et al., 2012; Valenzuela-Moreno et al., 2020; Rico-Torres et al., 2024).

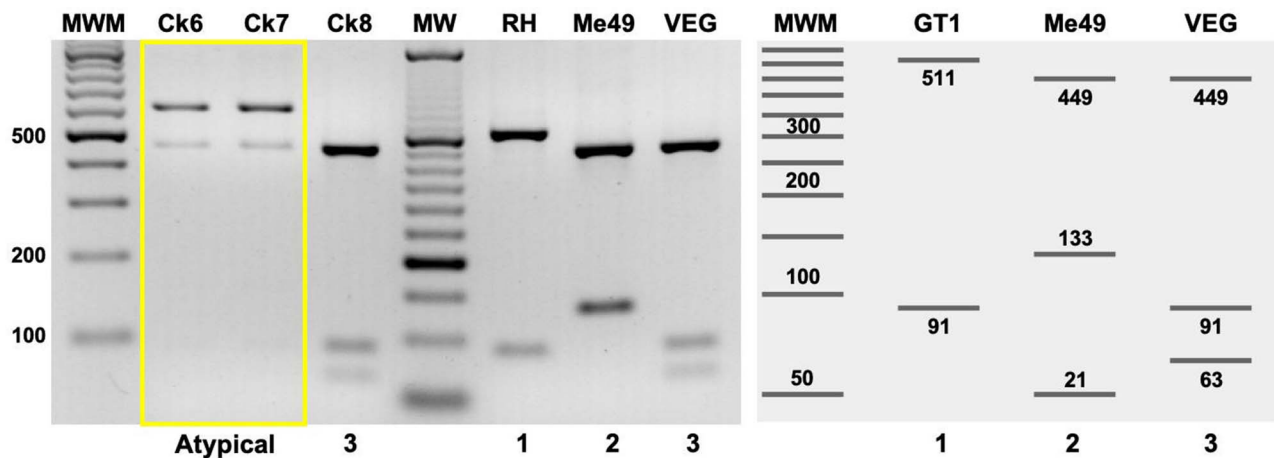
Twelve isolates were obtained from eight chickens that resulted in 6 different genotypes of *T. gondii*, three that had already been reported in México and other countries on the American continent and three previously unpublished genotypes. The ToxoDB #8 genotype (Type BrIII) was first isolated in México from a stray dog from Chiapas, and it is widely spread in Brazil and other American countries, such as the USA and Venezuela (Pena et al., 2008; Shwab et al., 2014; Valenzuela-Moreno et al., 2020). This is the second time that ToxoDB #38 has been isolated in México, and it has otherwise only previously been reported in Colombia, but it could be distributed throughout the tropical regions of southern México and Central and South America (Rajendran et al., 2012; Rico-Torres et al., 2024). The ToxoDB #28 genotype has been isolated several times in states on the Pacific coast and central and southeastern México; thus, it could be a regional genotype similar to BrI-BrIV and China I of Brazil and China, respectively (Shwab et al., 2014; Dardé et al., 2020; Rico-Torres et al., 2023).

The three new genotypes identified in this study are ToxoDB #344, #345 and #346. The first is a recombinant strain I-III with an even number of Type I and III alleles, whereas the remaining two genotypes are recombinant strains I-II-III with a majority of Type III alleles. Notably, genotype #345 is almost identical to genotype #8, except for the Type I allele at the *Apico* locus instead of Type III.

**Table 1.** PCR-RFLP genotypes of *Toxoplasma gondii* isolates obtained from free-range chickens in the state of Tabasco, México

ID	Municipality	MAT	Isolate <sup>a</sup>	Tissue of isolation	<i>B1</i> qPCR <sup>b</sup>	Tachyzoites per mg tissue	Genetic markers											ToxoDB PCR-RFLP genotype
							<i>SAG1</i>	5'3' <i>SAG2</i>	<i>Alt. SAG2</i>	<i>SAG3</i>	<i>BTUB</i>	<i>GRA6</i>	<i>c22</i> – 8	<i>c29</i> – 2	<i>L358</i>	<i>PK1</i>	<i>APICO</i>	
Chicken 1	Macuspana	1:40	TgCkMxTab1a	Heart	+	11 583	I	I	I	I	I	I	II	I	III	I	III	#28
			TgCkMxTab1b	Brain	+	55	I	I	I	I	I	I	II	I	III	I	III	#28
Chicken 2	Nacajuca	Neg	TgCkMxTab2	Heart	+	<50	I	I	I	I	I	I	II	I	III	I	III	#28
Chicken 3	Nacajuca	1:20	TgCkMxTab3a	Heart	+	4503	I	III	III	III	III	III	II	III	III	III	III	#8 (BrIII)
			TgCkMxTab3b	Brain	+	920	I	III	III	III	III	III	II	III	III	III	III	#8 (BrIII)
Chicken 4	Teapa	1:40	TgCkMxTab4a	Heart	+	374	I	III	III	III	I	I	I	III	I	I	III	#38
			TgCkMxTab4b	Brain	+	1579	I	III	III	III	I	I	I	III	I	I	III	#38
Chicken 5	Teapa	1:40	TgCkMxTab5a	Heart	+	18 026	I	III	III	III	I	I	I	III	I	I	III	#38
			TgCkMxTab5b	Brain	+	14 581	I	III	III	III	I	I	I	III	I	I	III	#38
Chicken 6	Jalapa	1:40	TgCkMxTab6	Heart	+	2024	I	III	III	III	I	I	III	III	III	I	I	NEW #344
Chicken 7	Centro	1:40	TgCkMxTab7	Brain	+	1052	I	III	III	III	III	III	II	III	III	III	I	NEW #345
Chicken 8	Centro	1:10	TgCkMxTab8	Heart	+	2301	I	I	I	III	I	III	II	III	III	III	III	NEW #346

<sup>a</sup>*Toxoplasma gondii* strains were designated 'a' and 'b' when two isolates were obtained from the same chicken. <sup>b</sup>A 62 bp fragment of the *B1* gene of *T. gondii* was detected in chicken tissues by qualitative PCR.



**Figure 2.** New *ROP5* allele identified in *Toxoplasma gondii* isolates from chickens in México. The restriction patterns of the Ck6 and Ck7 isolates (TgCkMxTab6 and TgCkMxTab7, respectively) were different from those of the reference strains RH, me49 and VEG, Types I, II and III, respectively (yellow box). The restriction products were resolved on 3% agarose gels stained with EtBr. MWM: molecular weight marker of 100 (agarose) and 50 (*in silico*) bp. Virtual digestion of the *ROP5* locus sequences of the archetypal strains GT1, Me49 and VEG, available at [www.Toxodb.Org](http://www.Toxodb.Org). Virtual digestion was performed with the benchling digestion tool ([www.Benchling.Com](http://www.Benchling.Com)). The image was converted to negative using GIMP v2.8 software.

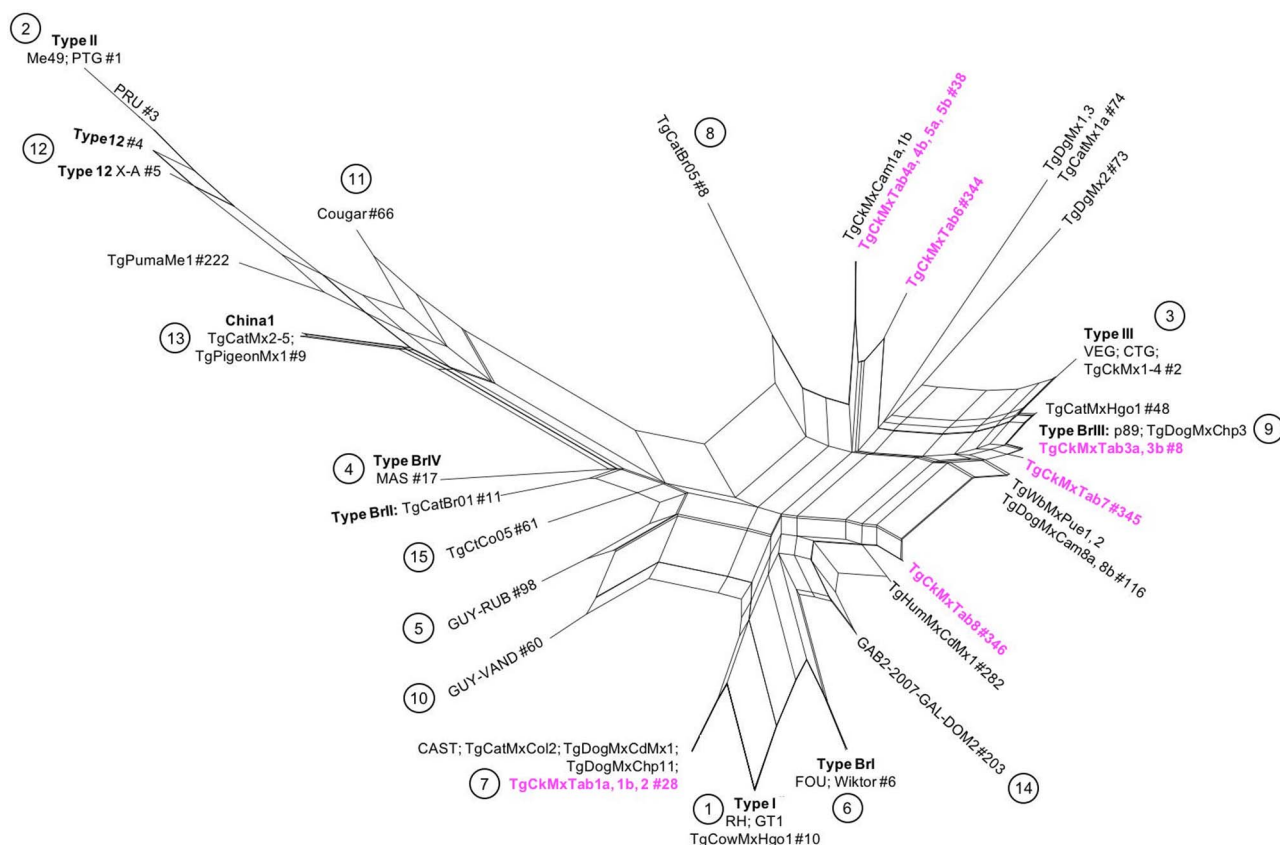
**Table 2.** Virulence multilocus PCR-RFLP typing *Toxoplasma gondii* isolates from free-range chickens in the state of Tabasco, México

Isolate ID	ToxoDB PCR-RFLP	Virulence genetic markers					Virulence prediction in mouse bioassay <sup>+</sup>
		CS3	<i>ROP16</i> <sup>+</sup>	<i>ROP17</i> <sup>^</sup>	<i>ROP18</i>	<i>ROP5</i>	
RH	#10	I	1	1	1	1	High virulence
Me49	#1	II	2	2	2	2	Low virulence
VEG	#2	III	1	2	3	3	Low virulence
<b>Present study</b>							
TgCkMxTab1a	#28	I	1	1	4	3	High virulence
TgCkMxTab1b	#28	I	1	1	4	3	High virulence
TgCkMxTab2	#28	I	1	1	4	3	High virulence
TgCkMxTab3a	#8 BrIII	III	1	1	3	3	Low virulence
TgCkMxTab3b	#8 BrIII	III	1	1	3	3	Low virulence
TgCkMxTab4a	#38	I	1	1	1	3	High virulence
TgCkMxTab4b	#38	I	1	1	1	3	High virulence
TgCkMxTab5a	#38	I	1	1	1	3	High virulence
TgCkMxTab5b	#38	I	1	1	1	3	High virulence
TgCkMxTab6	#344	III	1	1	3	<b>Atypical</b>	<b>Unknown<sup>o</sup></b>
TgCkMxTab7	#345	III	1	4	3	<b>Atypical</b>	<b>Unknown<sup>o</sup></b>
TgCkMxTab8	#346	III	1	4	3	3	Low virulence

<sup>\*</sup>Strains type I and III have allele 1. <sup>^</sup>Strains type II and III have allele 2. <sup>+</sup> According to Shwab et al. (2016). <sup>o</sup>There is no virulence profile for this combination.

According to Shwab et al. (2016), there is a correlation between the combination of *ROP18/ROP5* genetic markers and the outcome of infection in a murine model. At least six genetic profiles have been identified that predict high virulence (mortality > 80%), two with intermediate virulence (31–79%) and five with low virulence (<30%). Among the *T. gondii* isolates obtained in Tabasco, two with high virulence (4/3 and 1/3) and one with low virulence (3/3) were identified. Additionally, an atypical profile could not be associated with virulence because the *ROP5* restriction pattern was different from those previously reported (Shwab et al., 2016). The resulting restriction pattern is similar to that of a type III (faint)

allele, but most of the product remained undigested (Figure 2). The *ROP5* locus is composed of three paralogous genes arranged in tandem (*ROP5a*, *ROP5b* and *ROP5c*). The primers designed for the *ROP5* locus amplify these three paralogous genes; therefore, the isolates TgCkMxTab6 and TgCkMxTab7 may have some copies of an *ROP5* isoform with restriction sites for the *FspBI* enzyme that shows a faint Type III pattern, and the other paralogues may have lost their restriction sites, leaving them uncut (Reese et al., 2011; Shwab et al., 2016). Furthermore, this is the second time we found a different restriction pattern for this particular locus, the first being a recombinant locus 1/3 identified in two *T. gondii* isolates obtained





#38, which has been predicted to be highly virulent due to the *ROP18/ROP5* allele combination. However, its impact on animals and humans in the region remains uncertain. Tabasco is home of species considered highly susceptible to toxoplasmosis, such as the howler monkey (*Alouatta palliata* and *Alouatta pigra*) and the spider monkey (*Ateles geoffroyi*), which are listed as endangered by the Mexican Official Standard NOM-059-SEMARNAT-2010 and could be at risk of developing clinical problems and a fatal outcome if infected with these strains (Ceballos and Oliva, 2005; Dubey, 2010).

In conclusion, the *T. gondii* isolates TgCkMxTab6, TgCkMxTab7 and TgCkMxTab8 have the new genotypes ToxoDB#344, #345 and #346, respectively. In addition, we found *T. gondii* genotypes #8, #28 and #38, which have been previously described in other neighbouring states, suggesting that they are circulating in the tropical areas of southeastern México. Genotype #28 is becoming a frequent finding in different states of México. The ToxoDB#344 and #345 genotypes had an atypical allele at the *ROP5* locus, and this could be the ninth variant found for this paralogous gene. The combination of *ROP18/ROP5* alleles predicts high virulence in most of the *T. gondii* isolates obtained in the present study. However, this prediction must be confirmed by mouse bioassay. This study provides evidence of endemic *T. gondii* strains found in intermediate hosts to which other hosts, including humans, may be exposed.

**Supplementary material.** The supplementary material for this article can be found at <https://doi.org/10.1017/S0031182025100401>.

**Data availability.** Reference strain sequences of GT1, Me49 and VEG (*ROP5* and *ROP17*) are available at: [www.toxodb.org](http://www.toxodb.org). TgCkMxTab7-*ROP17* PP983038 and TgCkMxTab8-*ROP17* PP983039 are available at: <https://www.ncbi.nlm.nih.gov/genbank/>. ToxoDB genotypes are available at: <http://web.utk.edu/~csu1/ListToxoDB-PCR-RFLPgenotypes.html>.

**Acknowledgements.** The authors are very grateful to Dr. Chunlei Su for carefully reviewing the novel genotypes obtained from the *T. gondii* isolates and assigning the ToxoDB genotypes. We are grateful to Héctor Luna-Pastén and José Antonio Vargas-Villavicencio for their excellent technical assistance. Additionally, we thank Dr. Rafael Saavedra from the Institute of Biomedical Research-UNAM (México) for generously providing us with the RH *T. gondii* strain.

**Author contribution.** L.F.V.M., C.C.P. and H.C.O. conceived and designed this study. L.F.V.M., C.P.R.T. and H.C.O. directed and supervised the study. C.P.R.T., L.F.V.M., and H.C.O. performed the data analysis and drafting of the article. L.F.V.M., C.C.P., L.G.M.P., C.V.Z.V., and F.G.L. performed the fieldwork, necropsies and material acquisition. C.P.R.T., L.B.O.A., I.C.S. and L.X.G. performed laboratory procedures. All of the authors have read and approved the final manuscript.

**Financial support.** This work was partially supported by the Consejo Nacional de Humanidades, Ciencias y Tecnologías, CONHACyT, México (grant A1-S-21955) and by Programa E022, Investigación y Desarrollo Tecnológico en Salud (grant 2020/039) from the Instituto Nacional de Pediatría.

**Competing interests.** None declared.

**Ethical standards.** This study was performed at the Instituto Nacional de Pediatría of the Ministry of Health of México and was approved (protocol number 2020/039) by the Institutional Review Board (organization number IORG0011533, which includes the Research and Animal Care Committee). All experiments were performed in accordance with relevant guidelines and regulations. This study is reported in compliance with the ARRIVE guidelines (<https://arriveguidelines.org>).

## References

- Buxton D (1990) Ovine toxoplasmosis: a review. *Journal of the Royal Society of Medicine* **83**, 509–511. doi:10.1177/014107689008300813
- Caballero-Ortega H, Uribe-Salas FJ, Conde-Glez CJ, Cedillo-Pelaez C, Vargas-Villavicencio JA, Luna-Pastén H, Cañedo-Solares I, Ortiz-Alegría LB and Correa D (2012) Seroprevalence and national distribution of human toxoplasmosis in México: Analysis of the 2000 and 2006 National Health Surveys. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **106**, 653–659. doi:10.1016/j.trstmh.2012.08.004
- Calero-Bernal R, Fernández-Escobar M, Katzer F, Su C and Ortega-Mora LM (2022) Unifying virulence evaluation in *Toxoplasma gondii*: A timely task. *Frontiers in Cellular & Infection Microbiology* **12**, 1–17. doi:10.3389/fcimb.2022.868727
- Ceballos G and Oliva G (2005) Mamíferos silvestres de México. Conabio (ed.), *Primera Edición*. FCE, Ciudad de México. México: Comisión Nacional para el Conocimiento y Uso de la Biodiversidad. Fondo de Cultura Económica.
- Cedillo-Peláez C (2009) Determinación de genotipos de *Toxoplasma gondii* en fauna silvestre en México. MSc Dissertation, Universidad Nacional Autónoma de México.
- Cedillo-Peláez C, Rico-Torres CB, Salas-Garrido CG and Correa D (2011) Acute toxoplasmosis in squirrel monkeys (*Saimiri sciureus*) in Mexico. *Veterinary Parasitology* **180**, 368–371. doi:10.1016/j.vetpar.2011.03.012
- Comisión Nacional para el Conocimiento y Uso de la Biodiversidad (CONABIO) y Gobierno del Estado de Tabasco (2021) La biodiversidad en Tabasco: Estudio de Estado. CONABIO 1. Available at: [https://www.biodiversidad.gob.mx/media/1/region/eeb/files/TABASCO\\_resumen.pdf](https://www.biodiversidad.gob.mx/media/1/region/eeb/files/TABASCO_resumen.pdf) (accessed 14 May 2025).
- Dardé ML, Mercier A, Su C, Khan A and Grigg ME (2020) Molecular epidemiology and population structure of *Toxoplasma gondii*. In Weiss LM and Kim K (eds.), *Toxoplasma Gondii*. Amsterdam: Academic Press, 63–116.
- Dubey JP (1998) Refinement of pepsin digestion method for isolation of *Toxoplasma gondii* from infected tissues. *Veterinary Parasitology* **74**, 75–77. doi:10.1016/S0304-4017(97)00135-0
- Dubey JP (2010) *Toxoplasmosis of Animals and Humans, Second Edition* Boca Raton, Florida: CRC Press.
- Dubey JP and Desmonts G (1987) Serological responses of equids fed *Toxoplasma gondii* oocysts. *Equine Veterinary Journal* **19**, 337–339. doi:10.1111/j.2042-3306.1987.tb01426.x
- Dubey JP, Sundar N, Gennari SM, Minervino AHH, Farias Na da R, Ruas JL, Dos Santos Trb, Cavalcante GT, Kwok OCH and Su C (2007) Biologic and genetic comparison of *Toxoplasma gondii* isolates in free-range chickens from the northern Pará state and the southern state Rio Grande do Sul, Brazil revealed highly diverse and distinct parasite populations. *Veterinary Parasitology* **143**, 182–188. doi:10.1016/j.vetpar.2006.08.024
- Flegr J, Prandota J, Sovičková M and Israili ZH (2014) Toxoplasmosis – A global threat. Correlation of latent toxoplasmosis with specific disease burden in a set of 88 countries. *PLoS ONE* **9**, e90203. doi:10.1371/journal.pone.0090203
- Gazzinelli RT, Mendonça-Neto R, Lilue J, Howard J and Sher A (2014) Innate resistance against *Toxoplasma gondii*: An evolutionary tale of mice, cats, and men. *Cell Host & Microbe* **15**, 132–138. doi:10.1016/j.chom.2014.01.004
- Gennari SM, Pena HFJ, Soares HS, Minervino AHH, de Assis Ffv, Alves BF, Oliveira S, Aizawa J, Dias RA and Su C (2024) A population study of *Toxoplasma gondii* in the Amazon region expands current knowledge of the genetic diversity in South America. *PLOS Neglected Tropical Diseases* **18**, e0012737. doi:10.1371/journal.pntd.0012737
- Homan W, Vercammen M, De Braekeleer J and Verschuere H (2000) Identification of a 200- to 300-fold repetitive 529 bp DNA fragment in *Toxoplasma gondii*, and its use for diagnostic and quantitative PCR. *International Journal for Parasitology* **30**, 69–75. doi:10.1016/S0020-7519(99)00170-8
- Huson DH (1998) SplitsTree: Analyzing and visualizing evolutionary data. *Bioinformatics* **14**, 68–73. doi:10.1093/bioinformatics/14.1.68
- Huson DH and Bryant D (2006) Application of Phylogenetic Networks in Evolutionary Studies. *Molecular Biology and Evolution* **23**, 254–267. doi:10.1093/molbev/msj030



- Kompalic-Cristo A, Frotta C, Suárez-Mutis M, Fernandes O and Britto C (2007) Evaluation of a real-time PCR assay based on the repetitive *B1* gene for the detection of *Toxoplasma gondii* in human peripheral blood. *Parasitology Research* **101**, 619–625. doi:10.1007/s00436-007-0524-9
- Kurth K, Jiang T, Muller L, Su C and Gerhold RW (2021) *Toxoplasma gondii* contamination at an animal agriculture facility: Environmental, agricultural animal, and wildlife contamination indicator evaluation. *International Journal for Parasitology: Parasites and Wildlife* **16**, 191–198. doi:10.1016/j.ijppaw.2021.09.009
- Lorenzi H, Khan A, Behnke MS, Namasivayam S, Swapna LS, Hadjithomas M, Karamycheva S, Pinney D, Brunk BP, Ajioka JW, Ajzenberg D, Boothroyd JC, Boyle JP, Dardé ML, Diaz-Miranda MA, Dubey JP, Fritz HM, Gennari SM, Gregory BD, Kim K, Saeij JPI, Su C, White MW, Zhu XQ, Howe DK, Rosenthal BM, Grigg ME, Parkinson J, Liu L, Kissinger JC, Roos DS and David Sibley L (2016) Local admixture of amplified and diversified secreted pathogenesis determinants shapes mosaic *Toxoplasma gondii* genomes. *Nature Communications* **7**, 10147. doi:10.1038/ncomms10147
- Montoya JG and Liesenfeld O (2004) Toxoplasmosis. *The Lancet* **363**, 1965–1976. doi:10.1016/S0140-6736(04)16412-X
- Morrison DA (2005) Networks in phylogenetic analysis: New tools for population biology. *International Journal for Parasitology* **35**, 567–582. doi:10.1016/j.ijpara.2005.02.007
- Murillo-Léon M, Bastidas-Quintero AM and Steinfeldt T (2024) Decoding *Toxoplasma gondii* virulence: The mechanisms of IRG protein inactivation. *Trends in Parasitology* **40**, 805–819. doi:10.1016/j.pt.2024.07.009
- Pena HFJ, Gennari SM, Dubey JP and Su C (2008) Population structure and mouse-virulence of *Toxoplasma gondii* in Brazil. *International Journal for Parasitology* **38**, 561–569. doi:10.1016/j.ijpara.2007.09.004
- Rajendran C, Su C and Dubey JP (2012) Molecular genotyping of *Toxoplasma gondii* from Central and South America revealed high diversity within and between populations. *Infection Genetics & Evolution* **12**, 359–368. doi:10.1016/j.meegid.2011.12.010
- Reese ML, Zeiner GM, Saeij JPI, Boothroyd JC and Boyle JP (2011) Polymorphic family of injected pseudokinases is paramount in *Toxoplasma virulence*. *Proceedings of the National Academy of Sciences* **108**, 9625–9630. doi:10.1073/pnas.1015980108
- Rico-Torres CP, Valenzuela-Moreno LF, Luna-Pastén H, Cedillo-Peláez C, Correa D, Morales-Salinas E, Martínez-Maya JJ, Alves BF, Pena HFJ and Caballero-Ortega H (2023) Genotyping of *Toxoplasma gondii* isolates from México reveals non-archetypal and potentially virulent strains for mice. *Infection Genetics & Evolution* **113**, 105473. doi:10.1016/j.meegid.2023.105473
- Rico-Torres CP, Valenzuela-Moreno LF, Luna-Pastén H, Figueroa-Damián R, Gómez-Toscano V, Hernández-Delgado L, Escobedo-Torres MP and Correa D (2018) High heterogeneity, mixed infections and new genotypes in human congenital toxoplasmosis cases in the megametropolis of Central México. *Acta Tropica* **178**, 124–129. doi:10.1016/j.actatropica.2017.11.008
- Rico-Torres CP, Valenzuela-Moreno LF, Méndez-Cruz ST, Cedillo-Peláez C and Caballero-Ortega H (2022) Can cloning and sequencing help to genotype positive *Toxoplasma gondii* clinical samples? Results and validation using SAG3 as a model. *Infection Genetics & Evolution* **101**, 105283. doi:10.1016/j.meegid.2022.105283
- Rico-Torres CP, Valenzuela-Moreno LF, Robles-González E, Cruz-Tamayo AA, Huchin-Cab M, Pérez-Flores J, Xicoténcatl-García L, Luna-Pastén H, Ortiz-Alegria LB, Cañedo-Solares I, Cedillo-Peláez C, García-Lacy F and Caballero-Ortega H (2024) Genotyping of *Toxoplasma gondii* in domestic animals from Campeche, México, reveals virulent genotypes and a recombinant *ROP5* allele. *Parasitology* **151**, 363–369. doi:10.1017/S0031182024000106
- Schuneman MA, and Constantino CF (2002) *Técnicas de Necropsia En Animales Domésticos - 2: edición (Spanish Edition)*. Aldrete Velasco J (ed), Manual Moderno. Distrito Federal, México: Editorial Manual Moderno.
- Shwab EK (2015) Genetic diversity, population structure, and evolution of virulence in *Toxoplasma gondii*. PhD Dissertation, University of Tennessee.
- Shwab EK, Jiang T, Pena HFJ, Gennari SM, Dubey JP and Su C (2016) The *ROP18* and *ROP5* gene allele types are highly predictive of virulence in mice across globally distributed strains of *Toxoplasma gondii*. *International Journal for Parasitology* **46**, 141–146. doi:10.1016/j.ijpara.2015.10.005
- Shwab EK, Zhu XQ, Majumdar D, Pena HFJ, Gennari SM, Dubey JP and Su C (2014) Geographical patterns of *Toxoplasma gondii* genetic diversity revealed by multilocus PCR-RFLP genotyping. *Parasitology* **141**, 453–461. doi:10.1017/S0031182013001844
- Su C, Schwab EK, Zhou P, Zhu XQ and Dubey JP (2010) Moving towards an integrated approach to molecular detection and identification of *Toxoplasma gondii*. *Parasitology* **137**, 1–11. doi:10.1017/S0031182009991065
- Tenter AM, Heckeroth AR and Weiss LM (2000) *Toxoplasma gondii*: From animals to humans. *International Journal for Parasitology* **30**, 1217–1258. doi:10.1016/S0020-7519(00)00124-7
- Valenzuela-Moreno LF, Rico-Torres CP, Cedillo-Peláez C, Luna-Pastén H, Méndez-Cruz ST, Lara-Martínez G, Correa D and Caballero-Ortega H (2019) Mixed *Toxoplasma gondii* infection and new genotypes in feral cats of Quintana Roo, México. *Acta Tropica* **193**, 199–205. doi:10.1016/j.actatropica.2019.03.006
- Valenzuela-Moreno LF, Rico-Torres CP, Cedillo-Peláez C, Luna-Pastén H, Méndez-Cruz ST, Reyes-García ME, Correa D, Alves BF, Pena HFJ and Caballero-Ortega H (2020) Stray dogs in the tropical state of Chiapas, México, harbour atypical and novel genotypes of *Toxoplasma gondii*. *International Journal for Parasitology* **50**, 85–90. doi:10.1016/j.ijpara.2019.12.001
- Vargas-Villavicencio JA, Cedillo-Peláez C, Rico-Torres CP, Besné-Mérida A, García-Vázquez F, Saldaña JI and Correa D (2016) Mouse model of congenital infection with a non-virulent *Toxoplasma gondii* strain: Vertical transmission, “sterile” fetal damage, or both? *Exp. Parasitol* **166**, 116–123. doi:10.1016/j.exppara.2016.04.002
- Weilhammer D and Rasley A (2011) Genetic approaches for understanding virulence in *Toxoplasma gondii*. *Briefings in Functional Genomics and Proteomics* **10**, 365–373. doi:10.1093/bfpgp/blr028
- Zhang Y, Lai BS, Juhas M and Zhang Y (2019) *Toxoplasma gondii* secretory proteins and their role in invasion and pathogenesis. *Microbiological Research* **227**, 126293. doi:10.1016/j.micres.2019.06.003