

## REVIEW ARTICLE

# The Q fever epidemic in The Netherlands: history, onset, response and reflection

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## SUMMARY

The 2007–2009 human Q fever epidemic in The Netherlands attracted attention due to its magnitude and duration. The current epidemic and the historical background of Q fever in The Netherlands are reviewed according to national and international publications. Seroprevalence studies suggest that Q fever was endemic in The Netherlands several decades before the disease was diagnosed in dairy goats and dairy sheep. This was in 2005 and the increase in humans started in 2007. Q fever abortions were registered on 30 dairy goat and dairy sheep farms between 2005 and 2009. A total of 3523 human cases were notified between 2007 and 2009. Proximity to aborting small ruminants and high numbers of susceptible humans are probably the main causes of the human Q fever outbreak in The Netherlands. In general good monitoring and surveillance systems are necessary to assess the real magnitude of Q fever.

**Key words:** Control, *Coxiella*, outbreaks, Q fever, zoonoses.

## INTRODUCTION

Q fever is a zoonosis caused by *Coxiella burnetii*, an intracellular Gram-negative bacterium that is prevalent throughout the world [1]. Domestic ruminants are considered to be the main reservoir for Q fever in humans [2], although other animal species, including pet animals, birds and reptiles, may also be responsible for human cases. Transmission to humans is mainly accomplished through inhalation of contaminated aerosols. The main clinical symptom of Q fever

in goats and sheep is abortion and in cattle reduced fertility. With abortion, 1 000 000 000 *C. burnetii*/g placenta can be excreted [3]. Duration of shedding of *C. burnetii* by infected livestock varies depending on the excretion route and species. In milk, *C. burnetii* can be excreted for 8 days in ewes and up to 13 months in cattle. In faeces, *C. burnetii* can be excreted up to 8 days after lambing in ewes and up to 20 days in goats [4]. Goats may shed *C. burnetii* in two successive kidding periods [5]. Most animal species carrying *C. burnetii* show no symptoms at all [4]. In humans infection with *C. burnetii* remains asymptomatic in ~60% of infected persons. In symptomatic patients, acute Q fever usually presents as a flu-like, self-limiting

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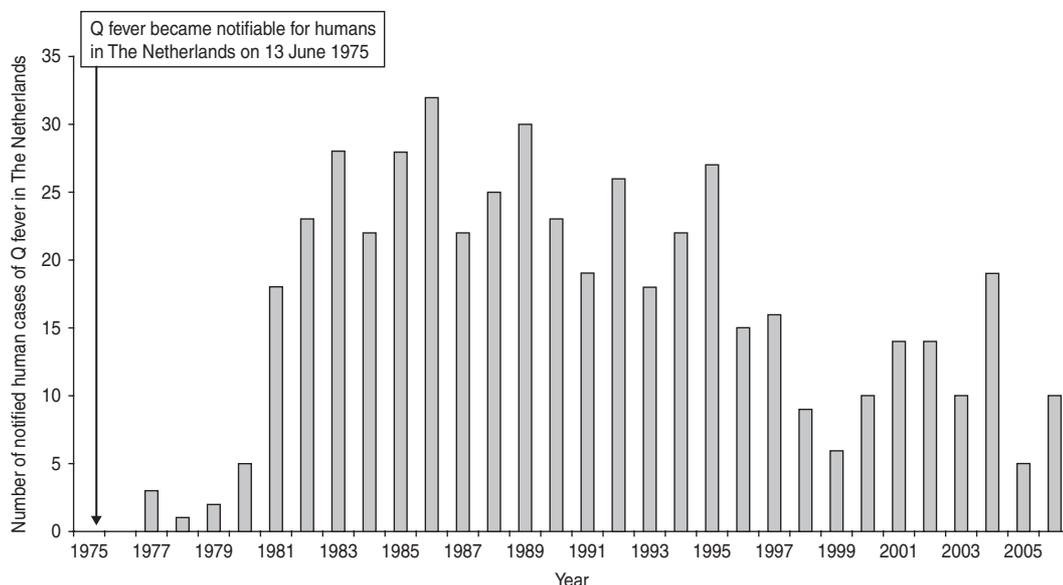


Fig. 1. Number of notified human cases in The Netherlands between 1975 and 2006.

disease, atypical pneumonia or hepatitis. Infection in pregnancy may lead to adverse pregnancy outcomes, such as spontaneous abortion or premature delivery. About 1–5% of all Q fever cases may progress into a chronic infection, often leading to life-threatening endocarditis [4, 6–9].

Since 2007, a Q fever outbreak has been ongoing in The Netherlands and this is referred to as the largest outbreak of Q fever ever reported in the literature [10]. However, Q fever is not an entirely new disease in The Netherlands. In this review we give an overview of the history of Q fever in The Netherlands, both in animals and in humans, the emergence of the disease and the control measures that have recently been taken. In conclusion, we put the outbreak in an international perspective.

### Q fever becomes endemic, 1956–2005

#### *Situation of Q fever in humans between 1956 and 2005*

In the 1950s a comprehensive survey of the global distribution of Q fever was commissioned by the World Health Organization. In The Netherlands, between 1951 and 1954, almost 10 000 human sera were tested for Q fever with a complement-fixation test and all were negative [11, 12]. In 1956, the first three human cases were reported in The Netherlands [13]. Two patients might have been linked to imported cattle or a visit abroad, while in the third patient there was no obvious cause. In 1958 and 1967 two human Q fever cases were described associated with the handling of imported wool [14, 15].

The notification of infectious diseases started in The Netherlands in 1865. The law changed several times and the list of notifiable diseases became longer with every change. Q fever was made notifiable in 1975, although it was a very rare disease at that time [16]. Between 1975 and 2006 the annual number of cases increased from 0 to 32 per year (Fig. 1). Thirty-three of the cases notified between 1979 and 1983 were investigated more thoroughly. Twenty-two (67%) of these patients were probably infected in The Netherlands through contact with animals or animal products [17]. A survey in 1982/1983 among 432 persons, considered to be at high risk because of close contact with animals and animal products, showed high percentages (58% in taxidermists to 84% in veterinarians) of seropositive responders with mainly IgG antibodies against *C. burnetii* phase-II antigen, indicating that Q fever had become endemic in The Netherlands [18].

The increase in human cases between 1977 and 1983 (Fig. 1) was assessed by Richardus *et al.* [18] by testing serum samples from persons, not considered to be at high risk, taken in 1968, 1975, 1979, and 1983 with an indirect immunofluorescence test (IFT) for specific IgG antibodies against *C. burnetii* phase-II antigen. In adults an average seroprevalence of 46% in 1968 and of 48% in 1983 was found. In children seroprevalences varied from on average 54% in 1975 to 28% in 1979 and 1983. These results showed no significant increase in the percentage of infected persons over the years 1968–1983. The increased number of notified cases since 1980 was explained by the

introduction of a sensitive indirect immunofluorescence test for IgM antibodies against *C. burnetii*. In the same study [18] occupational groups with a high risk of infection showed a significantly higher proportion of seropositives compared to the low-risk control group, in 1983 84% of 221 veterinarians and 68% of 94 residents of dairy farms were positive, whereas in the control group on average 29% tested positive for IgG antibodies against phase II of *C. burnetii*. The age distribution suggested an early onset of infection as antibody percentages over all age groups between 1 and 64 years were comparable [18]. It remains unclear why seroprevalence in the non-high-risk groups, varying between 30% and 50% in the 1980s, was high while on average 23 clinical Q fever cases were detected. Protective immunity from childhood [19] and under-diagnosis [20] could have played a role, but the serological test methods that were used in the 1980s were developed in-house and are no longer available, so the sensitivity and specificity of these tests can no longer be verified.

#### *Situation of Q fever in animals between 1956 and 2005*

Surveys in the early 1950s indicated that the bovine population was free of Q fever at that time [11, 12]. Between 1981 and 1987 the endemic state of Q fever in farm animals was confirmed by seroprevalence studies in cattle, sheep and goats [21, 22]. In 1981, 55% of 20 sampled cattle on one farm were tested positive. In 1987, 10% of 1320 non-dairy cattle, 21% of 1160 dairy cattle and 3% of a total of 494 dairy heifers were seropositive. At the herd level, about 36% were found positive with an average of 35% seropositive animals per herd. The occurrence of *C. burnetii* in cattle herds seemed to be associated with abortions, as 77% of the herds with abortions had a mean seroprevalence for Q fever of 39%. For the 31% of the herds without abortions, the mean seroprevalence was only 17%. In sheep 3.5% (127/3603 sheep sera from 191 flocks) of the animals were found positive with a herd prevalence of 27%. In goats 2/594 sera from individual goats from 54 flocks were positive. In 1992 prevalence in 220 representative cat sera was 10% and in 400 representative dog sera 13% (D. J. Houwers, *et al.* unpublished data).

#### **Transition period, 2005–2007**

##### *Changing situation of Q fever in animals since 2005*

Clinical Q fever in animals was diagnosed in The Netherlands for the first time in 2005 [23]. In two

Table 1. *Number of dairy goat and dairy sheep farms with confirmed Q fever abortions*

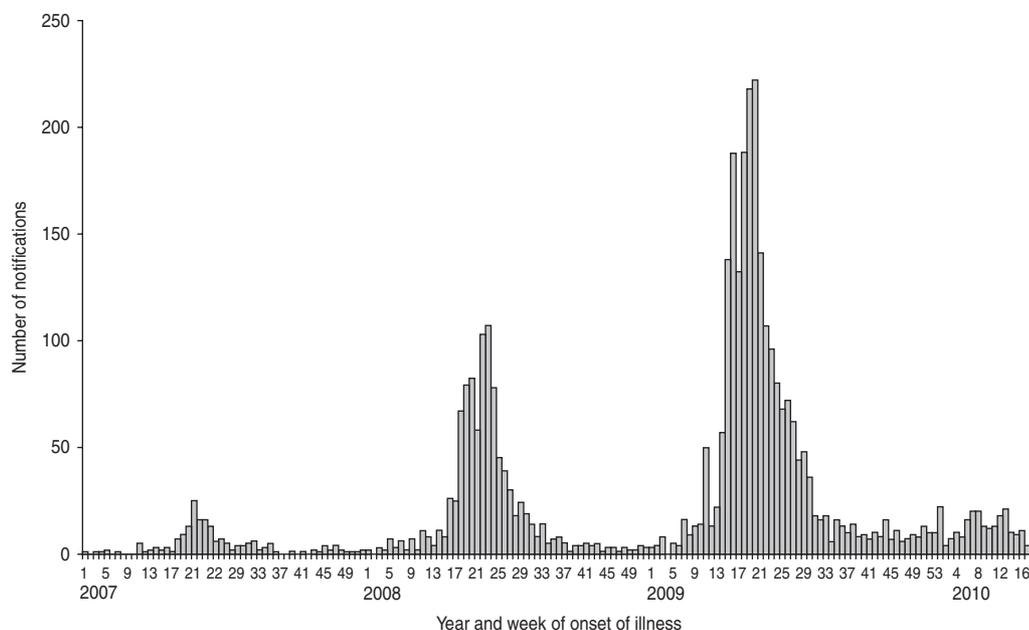
	2005	2006	2007	2008	2009	Total
Dairy sheep farms	—	1	—	1	—	2
Dairy goat farms	2	6	7	7*	6	28

\* Including one farm with animals at two locations.

dairy goat herds with abortion problems *C. burnetii* was detected by immunohistochemistry (IHC) on placentas [24]. From 2005 to 2007 Q fever abortions were diagnosed on 15 dairy goat farms and one dairy sheep farm (Table 1). Abortions, with herd rates up to 60%, were seen mainly in the final month of pregnancy without signs of general illness, although some goats were temporarily a little sluggish with reduced appetite. After abortion some goats showed symptoms of endometritis. Full-term kids were weak, with low body weight and high mortality. In several apparently healthy kids the rearing period was complicated by respiratory and digestive tract disorders. Treatment of pregnant goats with oxytetracyclines did not reduce the abortion rate [24]. Retrospectively, *C. burnetii* could be detected by IHC in one preserved dairy goat's placenta from a farm with abortion problems in 2001. In dairy cattle herds *C. burnetii* antibodies were detected on 57% of 344 farms using ELISA on bulk tank milk (BTM) samples during 2005–2006. It was calculated that on 35% of the farms at least 30% of the cattle could be positive [25].

##### *Changing situation in humans since 2007*

In 2007, individual human Q fever cases were reported, occurring after visits to dairy goat farms with abortion problems [24, 26]. The first documented outbreak of Q fever in The Netherlands was described by Karagiannis *et al.* [27] and Van Steenberghe *et al.* [28]. A total of 168 human cases were notified in 2007 (Fig. 2). The epidemic became apparent by a number of indicators: first, a medical microbiologist reported two patients admitted to hospital with severe pneumonia, who did not respond to standard antibiotic therapy. Four days later, a general practitioner informed the Municipal Health Service (MHS) of an unusual number of ten patients with atypical pneumonia in his practice. A similar report by another general practitioner from the same area was received 2 weeks later. Finally, an increase in the number of notifications for Q fever was noticed in the national registration system of the National Institute for



**Fig. 2.** Number of notified human Q fever cases with a known first day of illness according to the week of onset of symptoms, from 1 January 2007 to 11 May 2010. 2007 ( $n = 168$ ), 2008 ( $n = 1000$ ), 2009 ( $n = 2355$ ), 2010 ( $n = 208$ ). Total number of human cases in 2007–2009 ( $n = 3523$ ) (compiled by F. Dijkstra).

Public Health and the Environment (RIVM). No specific source could be identified, but it was postulated that the high number of abortions on the surrounding dairy goat farms might be the source of the human Q fever cases in the province of Noord-Brabant. Airborne transmission of contaminated dust particles could have been facilitated by the unusually hot and dry weather in the spring of 2007.

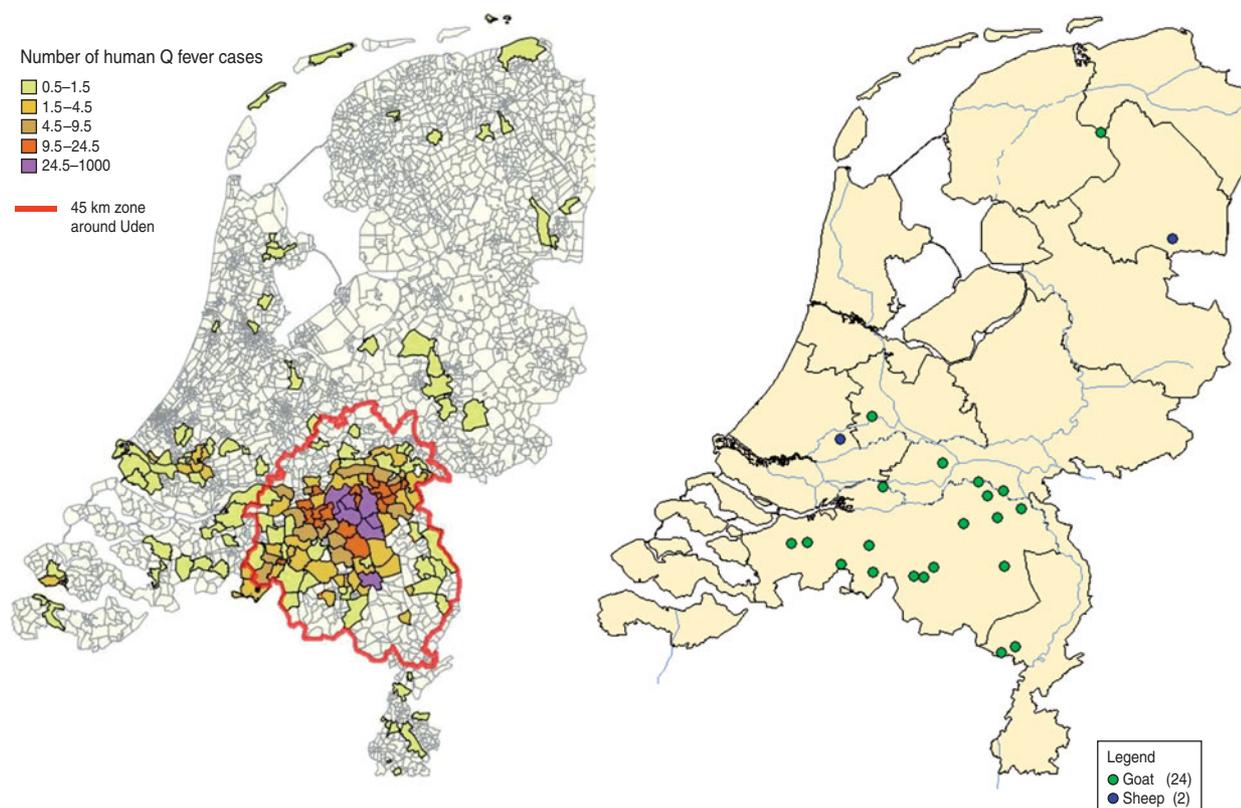
In 2007, the outbreak was concentrated around a single village and in this village a case-control study was performed [29]. Contact with manure, hay, and straw proved to be a risk factor. Moreover, people living in the eastern part of the village close to ruminant farms, of which one dairy goat farm had a recent history of abortion problems, were at higher risk than people living in other parts of the village. Contact with animals and consumption of raw milk products were not significant risk factors in the multivariable analysis.

Notification criteria for a confirmed human Q fever case were a clinical presentation with fever or pneumonia or hepatitis and confirmation of the diagnosis in the laboratory by at least a fourfold rise in IgG antibody titre against *C. burnetii* in paired sera or the presence of IgM antibodies against phase II or antibodies against *C. burnetii* phase I [30]. A probable case was defined as clinical signs with a single high antibody titre [31].

## Response phase, 2008–2010

### *Continuation of the human outbreak*

In 2008, it soon became clear that the 2007 outbreak was not an isolated incident. In May 2008 an outbreak of Q fever occurred in a psychiatric care institution in Nijmegen, province of Gelderland, ~15 km from the 2007 outbreak area [32]. At least 28 in-patients, employees, and visitors had laboratory-confirmed Q fever illness and several patients of the institution developed atypical pneumonia. A small flock of sheep without clinical symptoms of Q fever was present at the location. Patients had close contact with lambs, including cuddling as part of the patients' therapy sessions. Furthermore, on a dairy goat farm close to the city of Nijmegen, a large number of goats unexpectedly aborted their offspring. Q fever was confirmed by PCR on vaginal swabs. The farmer showed no clinical symptoms indicative for Q fever. The farmer's wife suffered from only moderate flu-like symptoms, including fever and coughing, and for both persons Q fever was confirmed by PCR on serum, throat swabs, urine and faecal samples. To determine the genetic relatedness between the human and animal clinical samples from both locations multiple-locus variable number tandem repeat analysis (MLVA) was carried out. Clinical samples from other patients from different locations in the same high-risk area were



**Fig. 3.** Map of The Netherlands. *Left:* Number of human cases in 2007 and 2008. The red line shows the dairy goat and dairy sheep voluntary vaccination area in 2008. *Right:* Dairy goat farms and dairy sheep farms with Q fever abortion history between 2005 and 2008. [Compiled by National Institute for Public Health and the Environment (RIVM) and Animal Health Service (GD).]

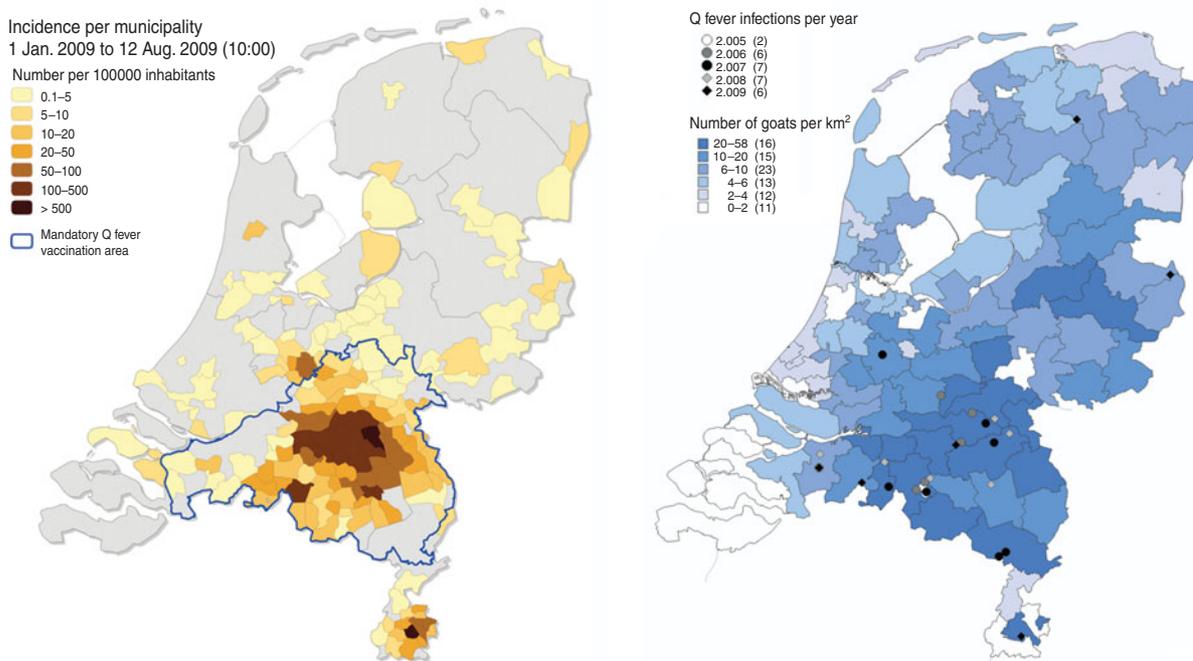
included in the study. All genotypes showed a high degree of similarity with most genotypes differing from each other by only a single marker, suggesting a clonal origin (J. J. H. C. Tilburg & C. H. W. Klaassen, unpublished observations) [33, 34].

Eventually, 1000 human Q fever cases were notified in 2008 (Fig. 2). The centre of the outbreak and the area with the most human cases were the same as in 2007, but there was a clear geographical spread to adjacent areas (Fig. 3). The age distribution (range 7–87 years, average 51 years) was similar to 2007, but the hospitalization rate decreased from 50% in 2007 to 21% in 2008 [10, 35]. The high hospitalization rate of 50% in 2007 might be biased by active case-finding in a retrospective survey among hospitalized cases [35]. The overall gender breakdown was the same in 2008 as in 2007: the female to male ratio was 1:1.7 [10].

In April 2009 a sharp increase in human cases was observed again resulting in a total number of 2355 cases (Fig. 2). Again most of the human cases were from the same area as in 2007 and 2008, with yet again a wider geographical spread (Fig. 4). Pneumonia

is the predominant presentation of Q fever in The Netherlands. For patients notified in 2008 for whom clinical details were available, 545 were diagnosed with pneumonia, 33 with hepatitis and 115 with other febrile illness [31]. The age median of 50 years in 2009 did not differ from that in 2008, neither did the hospitalization rate of 20% nor the gender breakdown of 1:1.7 female to male ratio [35].

The overall outbreak in humans consisted of at least 10 separate clusters with multiple sources of exposure. One cluster became apparent in 2008 with a strong connection to one goat farm with abortion problems. Patients were living downwind of the goat farm. Living within 2 km of the goat farm was associated with a higher risk of Q fever infection compared to living >5 km from the farm [36]. In 2009, no abortions were notified on this farm, nor in this area, and veterinary measures such as the handling of manure, hygiene measures and a visitors ban were implemented (measures are clarified in the section ‘Response in the veterinary field’), but the number of human cases still increased. To date, the source of this



**Fig. 4.** Map of The Netherlands. *Left:* Human Q fever incidence/100 000 inhabitants per municipality in 2009. The blue line shows the dairy goat and dairy sheep mandatory vaccination area in 2009. *Right:* Dairy goat farms with Q fever abortion history between 2005 and 2009. Darker blue indicates more goats/km<sup>2</sup>. [Compiled by National Institute for Public Health and the Environment (RIVM) and Animal Health Service (GD).]

increase in 2009 remains unclear [31]. In general, 59% of the notified human cases in 2009 lived within a 5-km zone around a notified dairy goat or dairy sheep farm, while 12% of the Dutch population live within such zones [35].

In 2010 criteria for detection of *C. burnetii* in blood, serum or a sample from the respiratory tract were added to the notification criteria for human Q fever cases. In addition, only acute cases with the day of first illness within 90 days of notification are recorded in the national infectious disease notification databases [37].

During the period January to May 2010, 208 human cases were registered which indicates a decrease in the number of notified human cases compared to 2009 (Fig. 2). Although weather conditions may have been unfavourable for transmission, it is hoped that the decrease in the number of human Q fever cases was caused by reduced exposure due to veterinary interventions.

#### *Dairy goat industry and prevalence of Q fever in small ruminants*

The dairy goat industry in The Netherlands is concentrated in the province of Noord-Brabant, with

farm sizes ranging from 300 to 7000 goats with an average of at least 600 animals in 2007. Goat density was 38.1 goats/km<sup>2</sup> [38]. These farms often bordered close to villages and cities. The total number of registered small ruminant farms in The Netherlands in 2008 was 52 000. The number of professional dairy goat farms with more than 200 adult goats was 350 and the professional dairy sheep industry consisted of 40 farms [39]. Dairy goat farming in The Netherlands started after the introduction of the European milk quotation system for dairy cattle in 1984 and increased after the outbreaks of classical swine fever in 1997 and foot-and-mouth disease in 2001. The total number of goats increased from 7415 in 1983 to 178 571 in 2000 and to 374 184 in 2009. The total number of dairy goats aged >1 year increased from 98 077 in 2000 to 231 090 in 2009 (Table 2) [40, 41].

In 2008, nationwide Q fever seroprevalence in all small ruminants was low; only 7.8% of goats and 17.8% of goat farms were positive, and 2.4% of sheep and 14.5% of sheep farms were positive. In 26% of the BTM samples from 306 dairy goat and dairy sheep farms *C. burnetii* DNA could be detected. Analysis of the first 13 goat farms with abortions showed an average number of goats per farm of 900 of which

Table 2. Number of goats in The Netherlands [40, 41]

Year	Total no. of goats	Total no. of dairy goats aged >1 year
1983	7415	n.r.
1995	76 063	n.r.
2000	178 571	98 077
2009	374 184	231 090

n.r., No registration.

20% aborted. The average number of sheep on the two affected dairy sheep farms was 400 with an abortion rate of 5% [39].

#### Response in the veterinary field

In June 2008, Q fever became notifiable for small ruminants kept for milk production following the advice of experts (Table 3). Additional measures were taken to reduce the assumed risk associated with the spread of manure and to restrict the number of visitors to infected farms. In October 2008, voluntary vaccination of goats was made possible by the Ministry of Agriculture in the high-risk Q fever area in Noord-Brabant with the so far unregistered phase-I Q fever vaccine for ruminants [Coxevac<sup>®</sup>, Ceva Santé Animale, France; Fig. 3 (red line), Table 3] [42]. A total of 36 000 goats were vaccinated in an area within a radius of 45 km around the village of Uden. This was the first time a Q fever vaccine had been used with the ultimate goal of reducing the number of human Q fever cases. In order to reduce the number of human cases the exposure of humans to *C. burnetii* should be reduced. To achieve this, excretion of *C. burnetii* from the animal host should be minimized, particularly by the prevention of abortion due to Q fever. Phase-I Q fever vaccines, contrary to phase-II vaccines, strongly reduce the number of abortions and excretion of *C. burnetii* in challenged pregnant goats that were initially Q fever-negative [43]. In clinically Q fever-infected goat herds vaccination with a phase-I vaccine should reduce the excretion of *C. burnetii*, especially in young animals vaccinated before the breeding season [44]. In general, vaccination with a phase-I Q fever vaccine is expected to be effective in non-infected goats. The effect of vaccination in Q fever-infected goats is not clear and may imply a continuation of the risk of shedding *C. burnetii* and exposure to humans. According to the summary of product characteristics (SPC), the phase-I Q fever vaccine is not indicated for use in pregnant sheep.

Assessment of the efficacy of the phase-I vaccine in pregnant cattle showed a similar probability of the animals becoming shedders when vaccinated while pregnant compared to non-vaccinated animals [45].

In February 2009, measures taken by the government were tightened with a stringent hygiene protocol made mandatory for all professional dairy goat and dairy sheep farms in The Netherlands, independent of their Q fever status. The hygiene protocol included vermin control, measures for handling manure (farmers were not allowed to remove manure from their deep litter stables for at least 1 month after the kidding season, were obligated to cover manure during storage and transport, and had to underplough manure immediately when spreading on farming land or had to store it for at least 3 months), compulsory rendering of aborted fetuses and placentas and improved general farm hygiene (such as the prevention of dust and aerosol formation, protective industrial clothing, clean delivery equipment and the use of sufficient high-quality bedding material). In addition, farmers were advised to submit aborted fetuses for pathological examination [39, 46]. Vaccination became mandatory for all dairy goats and dairy sheep, and the vaccination area was extended from the 45-km zone around Uden in 2008 (red line in Fig. 3) to the whole province of Noord-Brabant and small neighbouring areas in 2009 (indicated by the blue line in Fig. 4).

In response to the increasing number of human cases in 2009, additional measures were implemented in October 2009: PCR positivity of BTM on dairy goat and dairy sheep farms became a notification criterion of Q fever in small ruminants in addition to unexpectedly high abortion rates (>5%); a transport ban of animals from Q fever-positive farms and a visitors ban at Q fever-positive farms (Table 3). Improved monitoring of farm infection status with BTM monitoring was advised by experts. The background of this advice was that the abortion rate is difficult to measure in a flock with over 600 animals, and on infected farms *C. burnetii* can also be excreted in large quantities during normal birth. In large infected herds with many animals having normal deliveries, the total amount of excreted bacteria can also be very high with a subsequent risk for public health. BTM monitoring could give insight to the number of farms where *C. burnetii* is present. Initially, BTM monitoring was take place every 2 months, but with the increased awareness of the risk of Q fever and the possible risk that Q fever-positive farms might pose

Table 3. Overview of legislation concerning *Q* fever in small ruminants in The Netherlands [42]

Date of implementation	Document code	Measure
12 June 2008	TRCJZ/2008/1622	Q fever notifiable in dairy goats and dairy sheep; notification when over 5% abortions within 30 days at farms with more than 100 animals and when over 3% abortions within 30 days at farms with fewer than 100 animals (abortion rates up to 5% are considered to be more or less normal)
12 June 2008	TRCJZ/2008/1645	Prohibited from removing manure from the stable for 90 days after notification Visitors ban in place for 90 days after notification
16 October 2008	TRCJZ/2008/2817	Special dispensation for Coxevac (CEVA) Q fever vaccine to be used in The Netherlands Voluntary vaccination in dairy sheep and dairy goats at farms with more than 50 sheep or goats, petting zoos and nursing farms in the restricted 45-km zone (Fig. 3)
2 February 2009	TRCJZ/2009/244	Prohibited from farming more than 50 dairy goats and dairy sheep if certain hygienic measures are not implemented, such as vermin control, manure measures, rendering foetuses and placentas (see text)
20 April 2009	TRCJZ/2009/1142	Mandatory vaccination of dairy sheep and dairy goats on farms with more than 50 animals, on care farms, petting zoos and zoos in the extended area (Fig. 4) before 1 January 2010
1 October 2009	Regulation 40823	Mandatory bulk tank milk monitoring for Q fever every 2 months Prohibited from transporting dairy sheep and dairy goats from a positive farm. Vaccinated animals may be transported to positive farms Visitors ban in place at positive farms
9 December 2009	Regulation 96744	Ban on increase of numbers of dairy goats and dairy sheep on a farm Ban on reproduction of goats
1 January 2010	Regulation 72246	Mandatory vaccination of dairy sheep and dairy goats, on care farms, petting zoos, zoos, on farms open to the public, mobile sheep flocks, and in natural reserves nationwide before 2011
14 December 2009	Regulation 98748	Mandatory bulk tank milk monitoring for Q fever every 2 weeks
16 December 2009	Regulation 99604	Prohibited from removing manure from the stable within 30 days after ending of lambing season If manure has to be removed from the stable, it should be stored on the farm for 90 days
16 December 2009	Letter to Parliament; VDC 09.2695/CPM	Culling of all pregnant goats and sheep on Q fever-positive dairy goat and dairy sheep farms
18 December 2009	Regulation 101785	Prohibited from adding sheep or goats to a farm

during kidding season, the frequency of monitoring was increased to every 2 weeks. Due to BTM monitoring the number of Q fever-positive dairy goat and dairy sheep farms had increased to 88 on 17 April 2010 (Fig. 5).

With changing public and political awareness of Q fever, the Outbreak Management Team or expert panels advised the Ministries of Health and Agriculture about additional risk reduction measures. At a meeting in December 2009, experts stated that they expected vaccination would not be sufficiently effective in reducing the excretion of *C. burnetii* in the 2010 kidding season, due to the fact that not all dairy goats and dairy sheep had been vaccinated before the 2009 breeding season. A more considerable reduction in the possible excretion of *C. burnetii* and thus,

environmental contamination, thereby attempting to reduce human exposure and potential risk to public health in 2010, might be achieved by preventing pregnant Q fever-positive goats on Q fever-positive farms from kidding. These animals were identified as high-risk animals. Veterinary experts from the national veterinary reference institute (the Central Veterinary Institute of Wageningen UR), taking consideration of the findings of Rousset *et al.* [47], believed it impossible to distinguish infected pregnant animals from non-infected pregnant animals by laboratory testing prior to the start of the kidding season. This was the basis of the decision to cull all pregnant animals on Q fever-positive farms. In addition, breeding dairy goats and dairy sheep was prohibited until at least June 2010. The culling started in late



**Fig. 5.** Map of The Netherlands with all 88 bulk tank milk-positive dairy goat and dairy sheep farms known on 17 April 2010. The red zone is the 5-km zone around a positive farm. [Compiled by Dutch Ministry of Agriculture, Nature and Food Quality (LNV).]

December 2009 and was completed in June 2010, when a number of temporary animal measures imposed by the government ended and now require revision.

The measures in the veterinary field were taken to identify risk farms and to reduce the excretion of *C. burnetii* from these farms. Although the effectiveness of certain measures is still unclear and the lag time of the measures and the contribution of environmental contamination with *C. burnetii* to the

exposure of humans is unknown, the decreasing numbers of human cases in 2010 thus far might indicate that the measures taken since 2008 have been effective.

### The Dutch Q fever epidemic in perspective

Q fever was present in The Netherlands long before it became a problem in public health and animal husbandry. In this period, Q fever appeared to be neither

a major health problem for humans, or for domestic animals. This situation, with high seroprevalence in animal populations but few human cases, exists in most European countries [48]. In the years 2005 and 2006, Q fever became a problem in the dairy goat and dairy sheep industry. Abortion storms in small ruminants were not confirmed prior to 2005, although abortions with unknown aetiology are part of small ruminant husbandry with abortion rates up to 5%. It is not clear why Q fever became a major problem in The Netherlands but not elsewhere. Several factors could have facilitated a change in epidemiology in goats: first, an increase in goat density in specific areas of The Netherlands and second, extension of the farms over the years. These two factors could have affected in-herd and between-herd dynamics of Q fever, resulting in outbreaks. Third, there could be pathogen-related factors with circulation of a highly virulent *C. burnetii* strain. In the years 2007 and 2008, it became clear that Q fever also posed a problem to public health. The connection between Q fever problems in the dairy goat and dairy sheep industry and in the human population was made by several authors based on epidemiological and *C. burnetii* typing-based findings [28, 33, 35]. The increase in goat density took place in the highly populated province of Noord-Brabant (average population density in Noord-Brabant is 497 inhabitants/km<sup>2</sup>, compared to an average of 398 inhabitants/km<sup>2</sup> for The Netherlands). This proximity to a source excreting high numbers of *C. burnetii* during abortion, with transmission facilitated by dry weather and high numbers of susceptible humans is probably the main cause of the human Q fever outbreak in The Netherlands. The relationship between the change in epidemiology of Q fever in humans and changes in animal husbandry has been shown earlier. After the collapse of the large state farms in Bulgaria in the 1990s individual farmers started to raise goats, which resulted in a more than doubling of the number of goats to 1 million in 7 years [49]. This increase, together with a change to a more extensive husbandry system, was believed responsible for the increase of human cases. The increase in human Q fever in Germany was also probably related to socio-geographical factors associated with urbanization of rural areas [50]. Despite these examples no general conclusions can be drawn, as *C. burnetii* is endemic in domestic animals throughout Europe and infection can be maintained in a wide range of husbandry systems. Although risk factors, such as an association between human infection and small

ruminants, the proximity of animals (especially during parturition) and human populations, and specific weather conditions are clear, there is still an incomplete understanding of transmission pathways with regard to the maintenance of Q fever within the animal reservoir and its transmission to humans [48]. In addition, to date there are no indications of Dutch Q fever spreading to adjacent areas in Belgium and Germany [35].

The question can be raised if the Dutch outbreak is indeed the largest of its kind. *A posteriori*, it would be difficult to establish if the current epidemic in The Netherlands, with 3523 human cases within three consecutive years, represents a unique phenomenon. In Paragurische in Bulgaria, more than 2000 cases that were probably due to Q fever were diagnosed in a 6-month episode in 1993. Although confirmation of Q fever was hampered, these numbers of patients were comparable to the 2355 in 2009 in The Netherlands. The Q fever outbreak of the Balkans, named 'Balkangrippe', during the Second World War was at least comparable in size. In 1941 over 1000 cases were reported among German troops and during the years 1942–1945 comparable outbreaks were reported in general terms [51]. However, in these epidemics, no systematic investigations were performed. The importance of this in combination with the availability of diagnostic tests was also shown in the epidemic in a Swiss Alpine valley in 1983 [52]. The hospitalization of seven patients with atypical pneumonia was able to be diagnosed as due to Q fever as a result of newly available diagnostic tests. This detection was followed by a very large retrospective study in which infection was identified in nearly 15% of the inhabitants of the valley, probably correlated with migration of sheep. This showed that the proportion of patients presenting sufficiently severe symptoms to be hospitalized accounted for only 2%, and that 56% were completely asymptomatic. This means that for each patient being diagnosed, an additional 50 patients probably remain undiagnosed. This is solely based on systematic testing of the hospitalized patients with fever. Under these conditions, it is very difficult to evaluate the true incidence of Q fever, and it is highly likely that some epidemics have gone completely unnoticed. Thus, the reported incidence of Q fever depends on three distinct elements. First, it depends on the true incidence of the disease including existence of epidemics. Second, the reported incidence depends on sensitivity and specificity of the diagnostic tools used. In the Dutch epidemic the use of serological tests,

which are now commercially available and the use of real-time PCR on various clinical specimens allowed a better assessment of the incidence. Finally, the interest of the clinicians and the awareness of the general public incontestably reinforces the quality of detection and the percentage of detected patients. With monitoring systems more cases might be detected, but few data exist on large-scale monitoring of Q fever in the human population. In France, preliminary data showed a regular increase in the number of diagnosed cases of acute Q fever which probably testifies to the growing interest and the diagnostic capacities for this disease. Retrospective research in The Netherlands also shows that real-time syndrome surveillance might have detected clusters of human Q fever cases up to 2 years earlier than 2007 [53].

## CONCLUSION

The Q fever epidemic in humans in The Netherlands arose from an endemic state and was preceded by severe Q fever abortion problems in dairy goats and dairy sheep. Dairy goats and dairy sheep are considered to be the main source of human outbreaks and control measures were focused on these animal species. Although it is too early to evaluate the magnitude of the human Q fever epidemic in 2010, the decrease in the number of human cases compared to the previous year is promising. The proximity to small ruminants excreting high numbers of *C. burnetii* during abortion, with transmission facilitated by dry weather and high numbers of susceptible humans is probably the main cause of the human Q fever outbreak in The Netherlands. Q fever outbreaks can easily be missed in the human field as well in the veterinary field. In general good monitoring and surveillance systems are necessary to assess the real magnitude of Q fever.

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## DECLARATION OF INTEREST

None.

## REFERENCES

1. **Raoult D, Marrie TJ, Mege JL.** Natural history and pathophysiology of Q fever. *Lancet Infectious Disease* 2005; **5**: 219–226.
2. **Woldehiwet Z.** Q fever (coxiellosis): epidemiology and pathogenesis. *Research in Veterinary Science* 2004; **77**: 93–100.
3. **Arricau Bouvery N, et al.** Experimental *Coxiella burnetii* infection in pregnant goats: excretion routes. *Veterinary Research* 2003; **34**: 423–433.
4. **Arricau-Bouvery N, Rodolakis A.** Is Q fever an emerging or re-emerging zoonosis? *Veterinary Research* 2005; **36**: 327–349.
5. **Berri M, et al.** Goats may experience reproductive failures and shed *Coxiella burnetii* at two successive parturitions after a Q fever infection. *Research in Veterinary Science* 2007; **83**: 47–52.
6. **Fenollar F, Fournier PE, Raoult D.** Molecular detection of *Coxiella burnetii* in the sera of patients with Q fever endocarditis or vascular infection. *Journal of Clinical Microbiology* 2004; **42**: 4919–4924.
7. **Klee SR, et al.** Highly sensitive real-time PCR for specific detection and quantification of *Coxiella burnetii*. *BMC Microbiology* 2006; **6**: 2.
8. **McQuiston JH, Childs JE, Thompson HA.** Q fever. *Journal of the American Veterinary Medical Association* 2002; **221**: 796–799.
9. **Musso D, Raoult D.** *Coxiella burnetii* blood cultures from acute and chronic Q-fever patients. *Journal of Clinical Microbiology* 1995; **33**: 3129–3132.
10. **Schimmer B, et al.** Large ongoing Q fever outbreak in the south of The Netherlands, 2008. *Eurosurveillance* 2008; **13**: 18939.
11. **Kaplan MM, Bertagna P.** The geographical distribution of Q fever. *Bulletin of the World Health Organization* 1955; **13**: 829–860.
12. **Wolff JW, Kouwenaar W.** Investigation on occurrence of Q fever in Netherlands [in Dutch]. *Nederlands Tijdschrift voor Geneeskunde* 1954; **98**: 2726–2732.
13. **Westra SA, Lopes Cardozo E, Ten Berg J.** The first cases of Q-fever in the Netherlands [in Dutch]. *Nederlands Tijdschrift voor Geneeskunde* 1958; **102**: 69–72.
14. **Jordans GH.** A case of Q fever [in Dutch]. *Nederlands Tijdschrift voor Geneeskunde* 1958; **102**: 1548–1549.
15. **Terwindt VA, van Holten JW.** Q fever [in Dutch]. *Nederlands Tijdschrift voor Geneeskunde* 1967; **111**: 1951–1959.
16. **van Vliet JA.** History of notification [in Dutch]. *Tijdschrift voor Infectieziekten* 2009; **2**: 51–60.
17. **Richardus JH, et al.** Q fever in the Netherlands; a description of 33 case reports observed between 1979 and 1983 [in Dutch]. *Nederlands Tijdschrift voor Geneeskunde* 1984; **128**: 2253–2258.
18. **Richardus JH, et al.** Q fever in the Netherlands: a sero-epidemiological survey among human population groups from 1968 to 1983. *Epidemiology and Infection* 1987; **98**: 211–219.
19. **Richardus JH.** Q fever in the Netherlands – largely unknown [in Dutch]. *Infectieziekten Bulletin* 1998; **9**: 3–8.

20. **Gageldonk-Lafeber vAB, et al.** Occurrence of Q fever in the Netherlands [in Dutch]. *Infectieziekten Bulletin* 2003; **14**: 173–177.
21. **Houwers DJ, Richardus JH.** Infections with *Coxiella burnetii* in man and animals in The Netherlands. *Zentralblatt fuer Bakteriologie, Mikrobiologie und Hygiene, Series A* 1987; **267**: 30–36.
22. **Anon.** Is Q fever present in the Netherlands? [in Dutch]. *Tijdschrift voor Diergeneeskunde* 1981; **106**: 41–42.
23. **Vellema P.** New diseases Q fever and pasteurellosis appear [in Dutch]. *Geitenhouderij* 2006; July, pp. 14–15.
24. **Wouda W, Dercksen DP.** Abortion and stillbirth among dairy goats as a consequence of *Coxiella burnetii* [in Dutch]. *Tijdschrift voor Diergeneeskunde* 2007; **132**: 908–911.
25. **Muskens J, Mars MH, Franken P.** Q fever: an overview [in Dutch]. *Tijdschrift voor Diergeneeskunde* 2007; **132**: 912–917.
26. **Weevers A, et al.** Pneumonia after visiting a goat farm [in Dutch]. *Nederlands Tijdschrift voor Medische Microbiologie* 2007; **15**: 80–81.
27. **Karagiannis I, et al.** Q fever outbreak in the Netherlands: a preliminary report. *Eurosurveillance* 2007; **12**: 3247.
28. **Van Steenberghe JE, et al.** An outbreak of Q fever in The Netherlands – possible link to goats [in Dutch]. *Nederlands Tijdschrift voor Geneeskunde* 2007; **151**: 1998–2003.
29. **Karagiannis I, et al.** Investigation of a Q fever outbreak in a rural area of The Netherlands. *Epidemiology and Infection* 2009; **137**: 1283–1294.
30. **Centre for Infectious Disease Control, National Institute for Public Health and the Environment.** ([http://www.rivm.nl/cib/infectieziekten-A-Z/infectieziekten/Q\\_koorts/index.jsp](http://www.rivm.nl/cib/infectieziekten-A-Z/infectieziekten/Q_koorts/index.jsp)). Accessed 1 August 2009.
31. **Schimmer B, et al.** Sustained intensive transmission of Q fever in the south of the Netherlands, 2009. *Eurosurveillance* 2009; **14**: 19210.
32. **Koene RP, et al.** A Q fever outbreak in a psychiatric care institution in The Netherlands. *Epidemiology and Infection*. Published online: 9 February 2010. doi:10.1017/S095026881000021X.
33. **Klaassen CH, et al.** Multigenotype Q fever outbreak, the Netherlands [Letter]. *Emerging Infectious Diseases* 2009; **15**: 613–614.
34. **Tilburg JJHC, et al.** Molecular typing of *Coxiella burnetii* directly in various clinical samples shows a possible clonal origin of the ongoing Q-fever outbreak [Abstract]. *Antonie van Leeuwenhoek International Journal of General and Molecular Microbiology* 2009; **95**: 22.
35. **van der Hoek W, et al.** Q fever in the Netherlands: an update on the epidemiology and control measures. *Eurosurveillance* 2010; **15**: 19520.
36. **Schimmer B, et al.** The use of a geographic information system to identify a dairy goat farm as the most likely source of an urban Q-fever outbreak. *BMC Infectious Diseases* 2010; **10**: 69.
37. **Centre for Infectious Disease Control, National Institute for Public Health and the Environment.** ([http://www.rivm.nl/cib/infectieziekten-A-Z/infectieziekten/Q\\_koorts/index.jsp](http://www.rivm.nl/cib/infectieziekten-A-Z/infectieziekten/Q_koorts/index.jsp)). Accessed 2 April 2010.
38. **Roest HJ, et al.** Q fever in 2008 in the Netherlands and the expectations of 2009 [in Dutch]. *Tijdschrift voor Diergeneeskunde* 2009; **134**: 300–303.
39. **Van den Brom R, Vellema P.** Q fever outbreaks in small ruminants and people in the Netherlands. *Small Ruminant Research* 2009; **86**: 74–79.
40. **Statline, database Statistics Netherlands (CBS).** (<http://statline.cbs.nl/StatWeb/publication/?VW=T&DM=SLNL&PA=7316SLLB&D1=939,941,964,971,1009,1013,1020,1023-1024,1026-1028,1030-1033,1035-1038,1042&D2=0&D3=0-19&HD=090508-1703&HDR=T&STB=G1,G2>). Accessed 3 June 2010.
41. **Statline, database Statistics Netherlands (CBS).** ([http://statline.cbs.nl/StatWeb/publication/?VW=T&DM=SLNL&PA=71284NED&D1=443-451,483-491&D2=0&D3=0,5,\(1-2\),\(1-1\),1&HD=090602-1429&HDR=G1,G2&STB=T](http://statline.cbs.nl/StatWeb/publication/?VW=T&DM=SLNL&PA=71284NED&D1=443-451,483-491&D2=0&D3=0,5,(1-2),(1-1),1&HD=090602-1429&HDR=G1,G2&STB=T)). Accessed 3 June 2010.
42. **Dutch Ministry of Agriculture Nature and Food Quality (LNV).** ([http://www.minlnv.nl/portal/page?\\_pageid=116,3169803&\\_dad=portal&\\_schema=PORTAL](http://www.minlnv.nl/portal/page?_pageid=116,3169803&_dad=portal&_schema=PORTAL)). Accessed 28 March 2010.
43. **Arricau-Bouvery N, et al.** Effect of vaccination with phase I and phase II *Coxiella burnetii* vaccines in pregnant goats. *Vaccine* 2005; **23**: 4392–4402.
44. **Rousset E, et al.** Efficiency of a phase I vaccine for the reduction of vaginal *Coxiella burnetii* shedding in a clinically affected goat herd. *Clinical Microbiology and Infection* 2009; **15**: 188–189.
45. **Guatteo R, et al.** Assessing the within-herd prevalence of *Coxiella burnetii* milk-shedder cows using a real-time PCR applied to bulk tank milk. *Zoonoses and Public Health* 2007; **54**: 191–194.
46. **Dutch Ministry of Agriculture Nature and Food Quality (LNV).** Hygiene plan for dairy goat and dairy sheep farms [in Dutch] ([http://www.minlnv.nl/portal/page?\\_pageid=116,3169799&\\_dad=portal&\\_schema=PORTAL&p\\_document\\_id=2124959&p\\_node\\_id=7609889&p\\_mode=](http://www.minlnv.nl/portal/page?_pageid=116,3169799&_dad=portal&_schema=PORTAL&p_document_id=2124959&p_node_id=7609889&p_mode=)). Accessed 6 February 2009.
47. **Rousset E, et al.** *Coxiella burnetii* shedding routes and antibody response after outbreaks of Q fever-induced abortion in dairy goat herds. *Applied Environmental Microbiology* 2009; **75**: 428–433.
48. **EFSA Panel on Animal Health and Welfare.** Scientific Opinion on Q fever. *EFSA Journal* 2010; **8**: 1595.
49. **Serbezov VS, et al.** Q fever in Bulgaria and Slovakia. *Emerging Infectious Diseases* 1999; **5**: 388–394.
50. **Hellenbrand W, et al.** Changing epidemiology of Q fever in Germany, 1947–1999. *Emerging Infectious Diseases* 2001; **7**: 789–796.
51. **Spicer AJ.** Military significance of Q fever: a review. *Journal of the Royal Society of Medicine* 1978; **71**: 762–767.
52. **Dupuis G, et al.** An important outbreak of human Q fever in a Swiss Alpine valley. *International Journal of Epidemiology* 1987; **16**: 282–287.
53. **van den Wijngaard CC, et al.** In search of hidden Q-fever outbreaks: linking syndromic hospital clusters to infected goat farms. *Epidemiology and Infection*. Published online: 18 May 2010. doi:10.1017/S0950268810001032.