

# Fossil Lagerstätten and the enigma of anactulistic fossil preservation

Robert R. Gaines<sup>1</sup>  and Mary L. Droser<sup>2</sup><sup>1</sup>Geology Department, Pomona College, Claremont, California 91711, U.S.A.<sup>2</sup>Department of Earth Sciences, University of California–Riverside, Riverside, California 91711, U.S.A.

## Invited Article

**Cite this article:** Gaines, R. R., and M. L. Droser (2025). Fossil Lagerstätten and the enigma of anactulistic fossil preservation. *Paleobiology* **51**, 29–43.

<https://doi.org/10.1017/pab.2024.38>

Received: 04 December 2023

Revised: 29 July 2024

Accepted: 14 August 2024

### Handling Editor:

Mark Patzkowsky

### Corresponding author:

Robert R. Gaines;

Email: [robert.gaines@pomona.edu](mailto:robert.gaines@pomona.edu)

## Abstract

Over the last 50 years, paleobiology has made great strides in illuminating organisms and ecosystems in deep time through study of the often-curious nature of the fossil record itself. Among fossil deposits, none are as enigmatic or as important to our understanding of the history of life as Konservat-Lagerstätten, deposits that preserve soft-bodied fossils and thereby retain disproportionately large amounts of paleobiological information. While Konservat-Lagerstätten are often viewed as curiosities of the fossil record, decades of study have led to a better understanding of the environments and circumstances of exceptional fossilization. Whereas most types of exceptional preservation require very specific sets of conditions, which are rare but can occur at any time, Seilacher noted the problem of “anactulistic” modes of exceptional preservation, defined as modes of fossilization that are restricted in time and that no longer occur. Here, we focus on anactulistic preservation and the widely recognized overrepresentation of Konservat-Lagerstätten in the Ediacaran and early Paleozoic. While exceptional fossil deposits of Ediacaran, Cambrian, and Early Ordovician age encompass a number of modes of fossilization, the signal of exceptional preservation is driven by only two modes, Ediacara-type and Burgess Shale-type preservation. Both are “extinct” modes of fossilization that are no longer present in marine environments. We consider the controls that promoted widespread anactulistic preservation in the Ediacaran and early Paleozoic and their implications for the environmental conditions in which complex life first proliferated in the oceans.

## Non-technical Summary

Over the last 50 years, paleobiology has made great strides in illuminating organisms and ecosystems in deep time. Sometimes, these advances have come by interrogating the actual nature of the fossil record itself, specifically, the factors that govern how and why fossils are preserved. Among fossil deposits, none are as enigmatic or as important to our understanding of the history of life as deposits that preserve soft-bodied fossils and thereby retain unusually large amounts of paleobiological information. The great German paleontologist Adolf Seilacher called such deposits “Lagerstätten,” a term now taken to signify paleontological “mother lodes.” These fossil deposits typically represent precise sets of conditions that occur extraordinarily rarely but throughout the geologic record. Some types of these extraordinary deposits, however, represent essentially extinct modes of fossilization, which no longer occur within marine environments. Here, we consider these “anactulistic” styles of fossilization that once were widespread in Earth’s oceans, but only for a geologically brief period of time. We conclude that the circumstances that caused these lost pathways of fossil preservation resulted from specific suites of conditions that dominated ancient oceans during the rise of animal life. The same conditions that promoted anactulistic fossilization may have important implications for the circumstances in which complex life first proliferated in the oceans.

© The Author(s), 2025. Published by Cambridge University Press on behalf of Paleontological Society. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.

**PALEOBIOLOGY**  
A PUBLICATION OF THE  
 **Paleontological SOCIETY**

 **CAMBRIDGE**  
UNIVERSITY PRESS

## Introduction

Fifty years ago, Seilacher (1970) introduced the seminal concept of fossil Lagerstätten to paleontology. Defined as “fossil deposits unusually rich in paleontological information,” fossil Lagerstätten comprise two distinct types of deposits: Konzentrat-Lagerstätten, which are unusual concentrations of fossils, and Konservat-Lagerstätten, which are characterized by unusual fidelity of preservation and include extraordinary preservation of non-biomineralized soft tissues. While the former group holds potential to reveal anatomical and ecological information, unavailable from the “typical” fossil record, the latter group includes some clades of organisms that are entirely soft-bodied and, hence, are otherwise absent from the normal fossil record (e.g., Ediacara and Burgess Shale biotas). Study of fossils and assemblages of soft-bodied organisms thus offers unique insights into the history of life. In this paper, we focus on Konservat-Lagerstätten, here referred to simply as Lagerstätten (singular = Lagerstätte). For the purposes of this review, we use a basic definition of Lagerstätten to refer to deposits that preserve originally non-biomineralized tissues of fossil organisms.



Over the last half century of research, study of Lagerstätten has brought transformative new data to bear in three critical areas: (1) the paleobiology of individual organisms and the implications of key anatomical attributes for animal phylogeny; (2) a fuller picture of the diversity of life present at different intervals of geologic time and the structure of ancient ecosystems; and (3) novel insights into critical transitions in the history of life. Individual fossils often exhibit detailed preservation of soft-bodied morphology and may retain aspects of internal anatomy, including digestive, sensory, circulatory, and nervous systems (e.g., McCoy et al. 2016; Bicknell et al. 2021; Aria et al. 2023). In some cases, structural information at a cellular level is preserved (Strullu-Derrien et al. 2019; Demaris 2000; McNamara et al. 2016, 2018; Strullu-Derrien et al. 2019; Decombeix et al. 2023). Due to the extraordinary level of detail retained, fossils of soft-bodied organisms have yielded phenomenal insights for paleobiology by greatly extending the temporal ranges of many lineages, by revealing the existence of high-ranking taxonomic groups (i.e., phyla, classes) that have become extinct (Erwin et al. 2011), and through preservation of extraordinary anatomical information that informs phylogenetic study and continues to refine our understanding of the fundamental structure of the tree of animal life. In addition, Lagerstätten include organisms lacking biomineralized tissues and thus preserve components of diversity that are absent from typical fossil deposits, which are composed of shells, teeth, and bones. As such, Lagerstätten have contributed greatly to understanding ancient ecosystems from fossil records that, in many cases, are far more complete than typical fossil deposits. The position of some Lagerstätten in time and in space has furthermore helped to frame the modern understanding of many critical transitions in the history of life, including the origins of complex ecosystems (Droser and Gehling 2015), the initial colonization of land by invertebrates (Strullu-Derrien et al. 2019), and the theropod–avian transition (Chiappe 2009).

As originally recognized by Seilacher (1970) and Seilacher et al. (1985), Lagerstätten encompass many modes of exceptional preservation and represent many disparate environments and mechanisms of formation. Seilacher et al. (1985) considered three key environmental factors to be important in the genesis of exceptional fossil deposits: obrution, stagnation (anoxia), and microbial sealing. Subsequent research has demonstrated that each of these attributes are critically important at various points along a continuum of exceptional preservation. Intensive research, however, has also demonstrated that these three factors—alone or in combination—are insufficient to trigger the preservation of soft tissues and that other causal mechanisms are therefore required. Indeed, the mechanisms of preservation underpinning many types of Lagerstätten remain enigmatic and are the subject of intensive study and debate. For this reason, as well as the great rarity of these deposits, early research tended to view Lagerstätten as curiosities of the fossil record, outliers that could represent rare, isolated environmental settings, in contrast to more normal habitats represented in the “typical” fossil record (Briggs 2023).

While emphasizing the great rarity of Lagerstätten in the fossil record, Seilacher and colleagues (1985) also noted that these deposits are not distributed evenly in geologic time but exhibit a concentration in late Neoproterozoic and earliest Phanerozoic open-marine deposits, with a second, smaller concentration of exceptional preservation in restricted marine settings of the Jurassic (see also Allison and Briggs 1993). More recent work has revealed a third concentration of deposits from terrestrial settings of Paleogene and Neogene age (Muscente et al. 2019). In the last several decades of research, these patterns have become more robust as

the discovery and documentation of new Lagerstätten and understanding of their occurrence and distribution have become priorities for paleobiology (Allison and Briggs 1993; Butterfield 1995; Muscente et al. 2017). In particular, the long-standing problem of the Ediacaran–early Phanerozoic “taphonomic window” (Allison and Briggs 1993; Orr 2014) in open-marine settings has remained a topic of much debate.

In this paper, we focus on the marine record of fossil Lagerstätten in order to specifically address the issue of widespread exceptional preservation during the early history of complex life. We briefly review some of the major types of marine Konservat-Lagerstätten, with emphasis on the geologic circumstances of their occurrences, and consider the problem of the concentration of these deposits in open-marine settings of the Ediacaran and early Paleozoic. We argue that the circumstances that promoted the exceptional fossil record during the Ediacaran–Early Ordovician were widespread in global marine environments, a condition fundamentally unlike those of other types of Lagerstätten, and that the “anactulistic” circumstances (e.g., absent from similar environments today) responsible for exceptional preservation during this interval would have had an impact on the trajectory of the early diversification of animals.

### Large-Scale Controls on Exceptional Preservation

Exceptional preservation of non-biomineralized soft tissues occurs across a spectrum of environments represented in the rock record (Seilacher 1970; Seilacher et al. 1985). As a result, the taphonomic mode of soft tissue preservation, that is, the manner in which nonmineralized tissues are preserved—replicated by mineral phases (e.g., pyrite, calcium phosphate) or conserved as carbonaceous organic remains (e.g., in mudstones)—exhibits considerable variation (Clements and Gabbott 2022). Similarly, biostratinomic factors play a leading role in facilitating exceptional preservation, but they too vary widely among different settings where Lagerstätten form (Brett and Baird 1986; Behrensmeier et al. 2000). Accordingly, in their original classification, Seilacher and colleagues recognized that Lagerstätten are most meaningfully classified based on circumstances of their geologic occurrence, which reflect both pre- and postburial factors (biostratinomy and fossil diagenesis) that led to exceptional preservation in each case.

Seilacher et al. (1985) further classified the deposits according to the relative influences of three physical, chemical, and biological factors considered important in their genesis: obrution, stagnation (anoxia), and microbial sealing. Over the last 50 years, research has made great strides in understanding each of these factors. The rise of experimental taphonomy in the investigation of decay (Murdoch et al. 2014; Sansom 2014; Clements et al. 2022) and exceptional preservation (Briggs and Kear 1993b; Briggs 1995; McNamara 2013; McNamara et al. 2013; Briggs and McMahon 2016; Alleon et al. 2017; Muscente et al. 2017; Purnell et al. 2018; Slater et al. 2023), as well as a focus on intensive, fine-scale sedimentologic and geochemical investigation of Lagerstätten (Raiswell et al. 2008; Gaines et al. 2012c; McNamara et al. 2012; Farrell et al. 2013; Cotroneo et al. 2016; Tarhan et al. 2016), have brought new understanding to the genesis of these deposits and thus has helped to resolve the context and ecological significance of exceptional fossil assemblages.

Obrution, the rapid burial of fossils in sediments or volcanic ash, is an essential first step in a great majority of cases of exceptional preservation. Not only does rapid burial halt physical processes of disarticulation (Brett and Baird 1986), but it can also promote

changes in chemical environment that slow the processes of decomposition (Schiffbauer et al. 2014), and it can create isolated environments for fossil diagenesis and for the mineralization, molding, or stabilization of organic tissues (Darroch et al. 2012; McCoy 2014).

The role of anoxia (stagnation) in exceptional preservation in subaqueous environments has been the subject of intensive study. Experimental work quickly established that anoxia alone is insufficient to cause soft-bodied preservation, as microbial decomposition via anaerobic pathways is capable of complete destruction of soft tissues (Allison 1988a). Although anoxia precludes bioturbation and scavenging of buried carcasses and slows rates of microbial decomposition, the major contribution of anoxia to exceptional preservation is that it allows for the activity of anaerobic pathways of microbial respiration that can promote the rapid growth of authigenic minerals that may replicate soft tissues with high fidelity (Allison 1988b; Briggs 2003). The specific circumstances that favor mineralization of soft tissues, as well as those that favor the preservation of primary organic remains, are described later. Research in recent decades has also made clear that, while some types of aqueous soft-bodied preservation require anoxia in the benthic environment (Butterfield 1995; Gaines and Droser 2010), others may occur in anoxic burial environments beneath weakly oxygenated bottom waters (e.g., Farrell et al. 2011, 2013; Muscente et al. 2019, 2023). At a larger scale, research in recent decades has shown that atmospheric and oceanic oxygen levels did not reach modern concentrations until after the colonization of land by plants in the mid-Paleozoic (Dahl et al. 2010). Relatively low oxygen concentrations during the Neoproterozoic and early Paleozoic led to widespread anoxic and suboxic marine settings on the continental shelves (Gill et al. 2011; Saltzman et al. 2015), a factor now broadly recognized as significant in promoting exceptional preservation during this critical interval in the history of life (Gaines 2014; Muscente et al. 2017). It is also widely recognized that the majority of the marine record comes from epicontinental seas, which are prone to restriction and oxygen stress, particularly during greenhouse intervals (Peters 2007; Muscente et al. 2019).

“Sealing” by phototrophic microbial mats or by heterotrophic microbial biofilms that may rapidly form over decaying carcasses has also been demonstrated to play a significant role in the preservation of soft tissues, as originally hypothesized (Seilacher et al. 1985; Iniesto et al. 2015). Sealing is particularly important in settings where rapid burial is not favored, and its activity may have multiple chemical effects. Biofilms may serve to isolate the chemical environments around decaying carcasses, establishing and/or maintaining chemical gradients favorable to early diagenetic mineralization or, in some cases, trapping and concentrating ions liberated from carcasses or from sediments, and thus favoring the precipitation of minerals around soft tissues (Briggs and Kear 1993a; Wilby et al. 1996; Varejão et al. 2019).

### Fossil Lagerstätten in the Marine Realm

In the following sections, we briefly review some of the primary marine settings in which exceptional preservation occurs (Fig. 1, Table 1).

#### Deep Marine

Within the marine realm, Lagerstätten are particularly well represented in deep-marine settings characterized by oxygen deficiency,

including outer shelf, shelf break, and restricted/silled basin environments (Fig. 1C,D). These settings typically lie near but below the storm wave base, are dominated by mudstones, and may be subject to periodic obrution events resulting from storm wave disturbance of sediments higher on the slope. In these settings, three major modes of preservation occur: pyritization, phosphatization, and organic preservation. Molding of soft tissue anatomy, including via the remineralization of volcanic ash as described later, is also known from deep-marine settings.

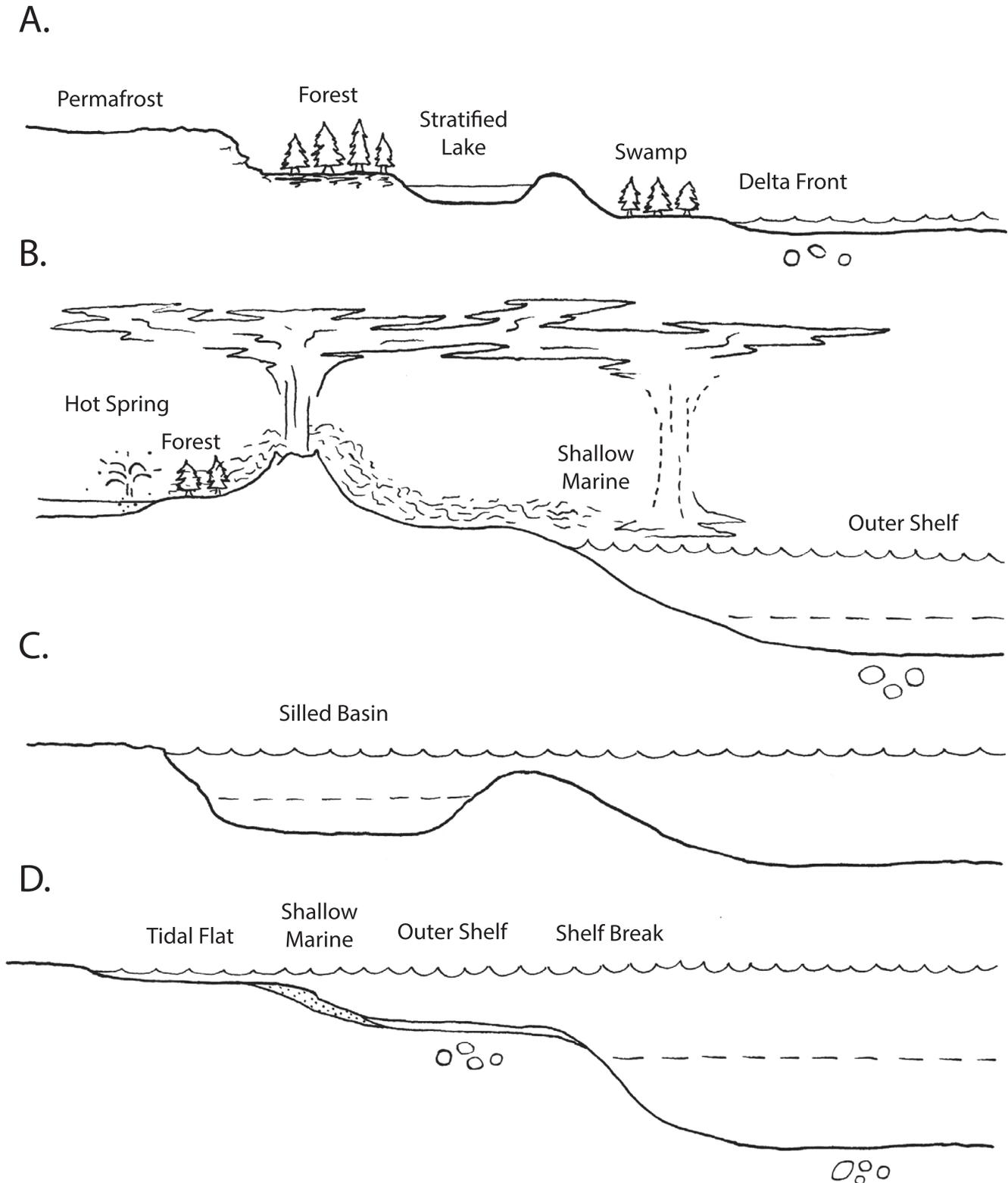
#### Pyritization

Pyritization as a primary mode of fossilization occurs where soft tissues are rapidly replicated in pyrite, as best known from Beecher's Trilobite Bed (Ordovician, New York), and Hunsrück (Devonian, Germany) (Raiswell et al. 2008; Farrell et al. 2013; Fig. 2A). In these deposits, pyrite is the primary agent responsible for capturing gross soft tissue morphology, often in three dimensions, and may also coat or replicate originally biomineralized tissues (e.g., trilobite cuticle; Farrell 2014). Because soft tissues rapidly collapse into two dimensions and are completely lost to decay on a timescale of weeks (Briggs and Kear 1993a), pyritization of soft tissues in three dimensions must occur before collapse.

Remarkable anatomical detail may be captured via pyritization, including the limbs, gills, and eggs of arthropods and the tube feet of asteroids (Farrell 2014; Siveter et al. 2014). Pyrite forms in the early burial environment due to the activity of microbial iron reduction, which liberates  $\text{Fe}^{2+}$  to solution by reduction of Fe oxides from sediments, and microbial sulfate reduction, which transforms aqueous  $\text{SO}_4^{2-}$  from seawater into  $\text{H}_2\text{S}^-$ . These by-products of microbial respiration react to form iron monosulfides, which are subsequently recrystallized to pyrite ( $\text{FeS}_2$ ). The pyritization of soft tissues requires an anoxic burial environment with iron-rich pore-waters and carbon-poor sediments that focus microbial sulfate reduction around freshly buried carcasses (Farrell 2014). As pyritization requires both anoxic pore-water conditions and a steady supply of seawater sulfate during fossilization, it typically occurs in dysoxic outer shelf settings where the diffusion of oxygen into the shallow burial environment is limited by low bottom water concentrations, but the diffusion of sulfate into pore-waters is unimpeded and thus not limiting to sulfate reduction (Farrell et al. 2013). Thus, the processes of pyritization require anoxic pore-waters and shallow burial conditions, but may occur under weakly oxygenated bottom waters (Farrell et al. 2011).

It has recently been demonstrated that, under the constraints described earlier, the conditions for pyritization of soft tissues were optimized in Beecher's Trilobite Bed by the recycling of sedimentary pyrite (Raiswell et al. 2008). There, early diagenetic pyrite-bearing sediments were eroded from the seafloor and entrained in turbidity currents that then swept up and buried soft-bodied animals. During transport, oxidation of some fraction of sedimentary pyrite in seawater formed labile Fe-(oxy)hydroxides, which were readily re-reduced shortly after deposition, resulting in particularly high iron concentrations of pore-waters in the freshly deposited sediment (Raiswell et al. 2008). The localization of sulfate reduction around soft tissues of animals then generated  $\text{H}_2\text{S}^-$ , which rapidly reacted with dissolved  $\text{Fe}^{2+}$  to form pyrite coatings around soft tissues (Briggs et al. 1991).

Soft tissue replication in pyrite also commonly occurs as an auxiliary mode of preservation in many other types of Lagerstätten; in these cases, pyrite replicates specific, limited aspects of soft-



**Figure 1.** Environmental settings of fossil Lagerstätten, after Seilacher et al. (1985). **A**, Terrestrial settings, including delta front transitional setting at right. **B**, Terrestrial and marine settings of volcanic-associated Lagerstätten. **C**, Silled/restricted marine basin environments. **D**, Marine settings of Lagerstätten. The position of the oxycline in **B–D** is indicated by dashed lines. Circles indicate loci of concretion formation.

bodied anatomy in association with another primary mode of preservation (Burgess Shale–type preservation, phosphatization, molding in concretions). Because it is a common early diagenetic product in marine sediments, pyrite may also be found in

association with soft-bodied fossils without capturing any morphological information; however, unless pyrite replicates specific aspects of soft-bodied anatomy, such pyrite should be considered purely incidental to preservation.

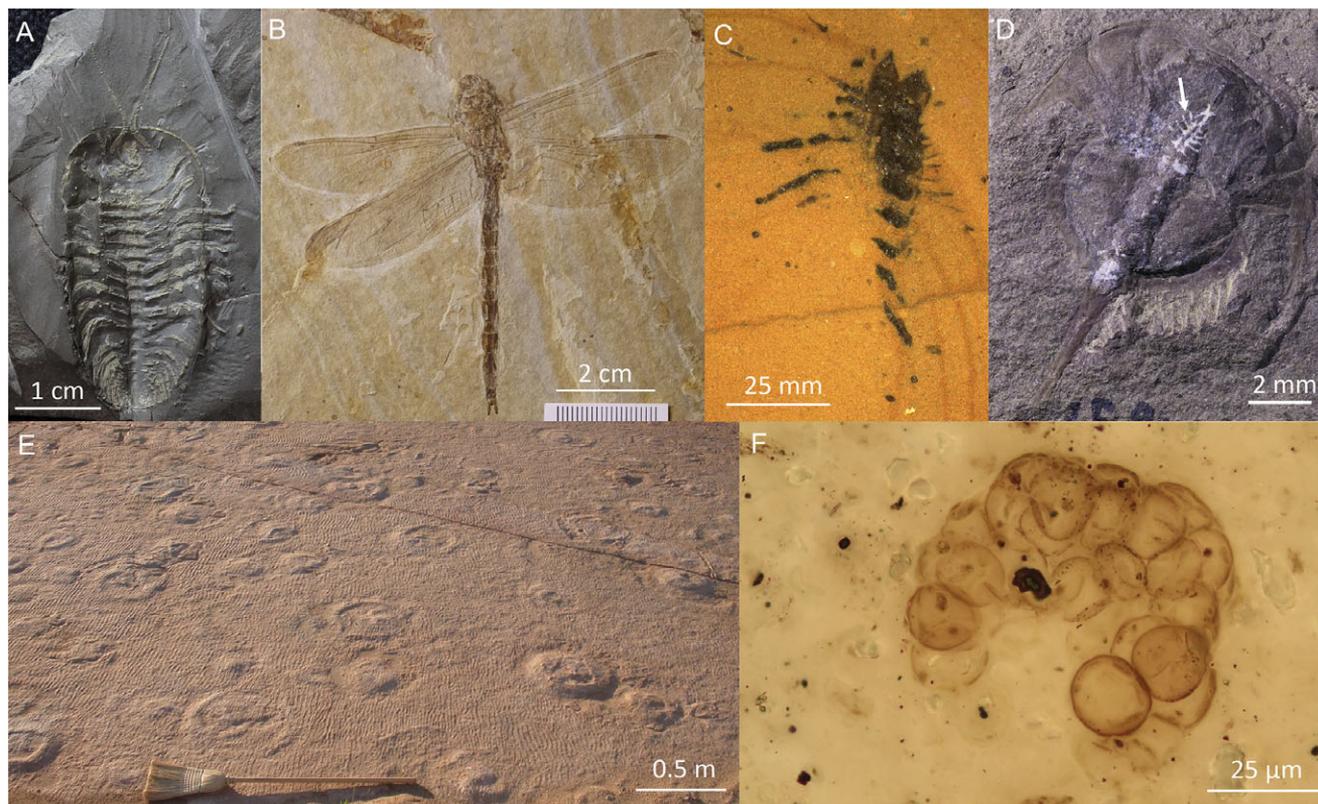
**Table 1.** Marine Konservat-Lagerstätten, arranged by environmental setting and mode of preservation. Blue highlights indicate “anactualistic” modes of preservation (Seilacher et al. 1985).

Environment	Primary mode of preservation	Example	Reference
Shelf break	Carbon	Burgess Shale	Gaines 2014
Outer shelf	Pyrite	Beecher's Trilobite Bed	Farrell 2014
	Mold and cast	Herefordshire	Siveter et al. 2020
	Phosphate	Orsten	Maas et al. 2006
Silled/restricted basin		Ya Ha Tinda	Muscente et al. 2019
Shallow marine	Mold and cast	Ediacara	Tarhan et al. 2016
Delta front	Siderite entombment (concretions)	Mazon Creek	Cotroneo et al. 2016
Evaporative tidal flat	Mold and cast	Mt. Simon/Wonewoc	Hagadorn et al. 2002
	Silica entombment	Bitter Springs	Butterfield 2003
Anactualistic mode of preservation			

### Phosphatization

Exceptional preservation via phosphatization, like pyritization, generally occurs in low-oxygen settings, close to the boundary of anoxic and dysoxic benthic environments. Phosphatized fossils may occur in three dimensions in massive phosphates (e.g., Doushantuo, Neoproterozoic, South China; Xiao and Knoll 2000) or in carbonate concretions (e.g., Orsten, Cambrian, Sweden; Maas et al. 2006), or as two-dimensional compressions (Fig. 2B) in black, organic-rich shales characterized by low net sedimentation rates and periodic obrution events (e.g., Ya Ha Tinda, Triassic, Canada; Muscente et al. 2019). Phosphatized compression fossils may also occur in restricted inter-reef settings in carbonate muds (Wendruff et al. 2020; Anderson et al. 2021; Pulsipher et al. 2022).

Phosphate ( $\text{PO}_4^{3-}$ ) is an essential nutrient in seawater that is rapidly taken up by phytoplankton and supplied to sediments as organic matter. It is also delivered to sediments by sorption onto iron oxides in the water column. In anoxic sediments, microbial reduction reactions liberate phosphate from both these sources to solution (Paytan and McLaughlin 2007). Phosphatization of soft tissues is favored by low-pH settings in the shallow burial environment at the anoxic–dysoxic transition in sediments where the precipitation of calcium phosphates as hydroxyapatite is favored over the precipitation of calcium carbonate (Allison 1988b; Briggs and Wilby 1996; Muscente et al. 2015). Low sediment accumulation rates and elevated primary productivity favor the enrichment of  $\text{PO}_4^{3-}$  in sediments, and restricted marine basins, including silled



**Figure 2.** Examples of exceptional preservation from selected marine and transitional fossil Lagerstätten. **A**, Pyritized trilobite (Holotype of *Triarthrus eatoni*) from Beecher's Trilobite Bed (Ordovician, New York), Yale Peabody Museum specimen number YPM IP 000219. **B**, Phosphatic preservation of a dragonfly (taxon indet.) from the Solnhofen Lagerstätte (Jurassic, Germany), Yale Peabody Museum specimen number YPM IP 428805. **C**, Three-dimensional preservation of the arthropod *Aquilonifer spinosus* as dark-colored calcite against lighter-colored concretion matrix from the Herefordshire Lagerstätte (Silurian, U.K.), Oxford Museum of Natural History specimen number OUMNH C.29695. **D**, Moldic preservation of the horseshoe crab *Euproops danae* in siderite concretion from the Mazon Creek Lagerstätte (Carboniferous, Illinois), including preserved neural tissues (arrow), Yale Peabody Museum specimen number YPM IP 168040. **E**, Bedding surface with multiple scyphozoan medusoids preserved as casts and molds in tidal flat sandstones of the Elk Mound Group (Cambrian, Wisconsin). **F**, Photomicrograph of organic-walled microfossils entombed in silica from cherts of the Fifteenmile Group (Tonian, ca. 800 Ma, Yukon, Canada). Images courtesy of D. Briggs and J. Utrup (**A–C**); R. Bicknell (**D**); J. Hagadorn (**E**); P. Cohen (**F**).

basins or restricted foreland basins (Fig. 1C), can serve as phosphate traps (Muscente et al. 2015). High concentrations of sedimentary phosphate, however, are lacking in many instances of soft tissue phosphatization, suggesting that soft tissues themselves contributed phosphate for mineralization (Briggs and Wilby 1996; McNamara et al. 2009, 2012). Indeed, many instances of phosphatization are restricted to specific tissue types (Butterfield 2002; McNamara et al. 2012). While reduced pH around decomposing carcasses certainly plays an important role in calcium phosphate mineralization (Briggs and Wilby 1996), recent experimental work has shown that tissue-specific phosphatization cannot be explained by pH gradients within decomposing carcasses. Because individual organs do not have unique pH trajectories during decay, tissue-specific phosphatization is more likely related to primary tissue composition (Clements et al. 2022).

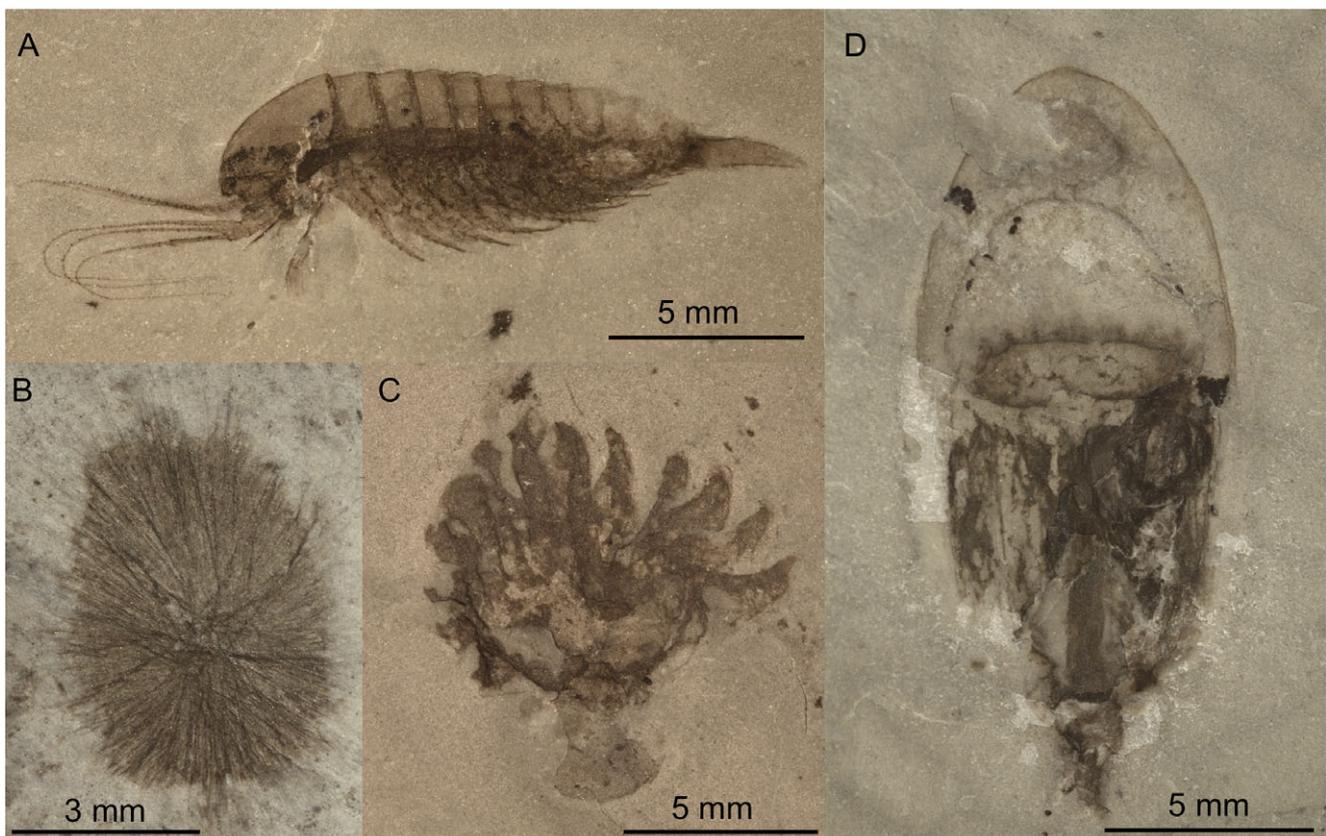
Spectacular examples of phosphatized soft tissues, including the muscle architecture of Devonian fish (Trinajstić et al. 2022) and the internal tissues of Jurassic invertebrates (Wilby et al. 1996), occur within calcium carbonate concretions from deep-marine environments. The formation of concretions may aid in the phosphatization process via sealing (McCoy et al. 2015a,b) or through development of low-pH microenvironments inside fossil carcasses (McNamara et al. 2012; Trinajstić et al. 2022). Such internal microenvironments may also promote phosphatization and may also occur as a secondary mode of preservation inside guts or other internal organs (Butterfield 2002).

#### Organic Preservation: Burgess Shale-type

The conservation of primary organic remains as carbonaceous compression fossils in marine settings is a particularly problematic

feature of the early Paleozoic fossil record. While the fossilization of the relatively recalcitrant organic remains of plants, algae, and graptolites as carbon may be explained by a combination of stagnation and obrution (e.g., Shiffbauer, et al. 2014), the carbonaceous preservation of guts, eyes, gills, and even nervous systems of soft-bodied organisms in open-marine conditions in outer shelf and shelf break settings (Fig. 3), widely known as “Burgess Shale-type” preservation (BST) is a feature unique to the Cambrian and Early Ordovician, and one that requires additional explanation (Butterfield 1995, 2003). While the abbreviation BST is also used to describe Burgess Shale-type fossil assemblages, here, we use BST to refer to a mode of preservation. As this mode of preservation of entire assemblages of soft-bodied organisms is restricted in time, as originally noted by Seilacher et al. (1985), and does not occur in modern environments, BST preservation is considered “anactulistic” (sensu Seilacher et al. 1985).

Burgess Shale-type preservation is widespread in outer shelf and shelf break mudstones of Cambrian–Early Ordovician age, where it occurs in more than 100 deposits at the junction of anaerobic and dysaerobic benthic environments (Gaines 2014; Muscente et al. 2017; Fu et al. 2019). Recent reinterpretation has suggested that the Chengjiang biota occupied a delta front environment (Saleh et al. 2022) where similar redox and biostratinomic conditions prevailed (Gaines et al. 2012b). As long understood from the type area of the Burgess Shale, BST assemblages were entrained in turbid flows; transported from habitable benthic environments to inhospitable, anoxic settings; and buried rapidly in fine-grained mud (Conway Morris 1986; Gaines and Droser 2010). Biostratinomic factors, namely slope angle, the presence of a shelf break, and the power of sediment-gravity flows, largely controlled the diversity of



**Figure 3.** Examples of Burgess Shale-type preservation of metazoan fossils from the early Cambrian Qingjiang biota (Fu et al. 2019), showing typical preservation as carbonaceous films (primary organic remains). **A**, Leanchoilid arthropod. **B**, Sponge belonging to the genus *Choia*. **C**, Cnidarian sea anemone. **D**, Medusoid cnidarian. All images courtesy of D. J. Fu.

BST assemblages as well as the fidelity of preservation of fine anatomical details (Gaines 2014). Although assemblages are transported, some do retain meaningful ecological information (Nanglu et al. 2020). The conservation of primary organic remains in these deposits requires either the special protection of soft-bodied fossils from the normal processes of microbial decomposition in sediments or a large-scale slowdown of microbial activity in the sediments at large (Gaines et al. 2008). Recent evidence supports both types of processes. Experimental evidence indicates that the clay mineral kaolinite, a product of intense chemical weathering, is capable of slowing microbial decay of organic tissues (Wilson and Butterfield 2014; McMahan et al. 2016) and comprised a significant fraction of the primary clay mineralogy of BST sediments (Anderson et al. 2018). In addition, the activity of microbial decomposition in BST sediments was shown to have been severely restricted in the early burial environment by the formation of seafloor crusts of calcite cements, which were emplaced shortly after deposition and restricted the diffusion of oxidants necessary to sustain microbial respiration into the sediments (Gaines et al. 2012c). Together, these effects acted to prematurely curtail microbial activity in sediments, resulting in incomplete decay, the collapse of tissues into two dimensions, and the preservation of primary organic remains as carbonaceous compressions.

#### Mold and Cast- Associated with Volcanic Ash

Exceptional preservation in volcanic ash is known from outer shelf settings in the Fezouata (Ordovician) and Herefordshire (Silurian) biotas. In the Fezouata, giant radiodont arthropods ~2 m in length are preserved in silica-chlorite concretions, inferred to have been derived from the remineralization of volcanic ash at the seafloor (Gaines et al. 2012a). Volcanic ash is comprised of fine metastable glass shards that are subject to rapid alteration upon entering the marine environment, liberating silica, aluminum, and iron (Duggen et al. 2010; Ayris and Delmelle 2012) from which quartz and clay minerals, including chlorite, may precipitate. In the case of Fezouata radiodonts, the formation of concretion-like masses of fine silicate minerals that molded the soft-bodied anatomy in three dimensions was inferred to be driven by low pH around decomposition of the giant carcasses (Gaines et al. 2012a). By contrast, the Herefordshire biota includes a diversity of invertebrate forms preserved as finely detailed external molds within concretions (Fig. 2C) inside an ash bed deposited in an outer shelf setting. The Herefordshire fossil molds occur as calcite-filled voids within carbonate concretions (Orr et al. 2000; Siveter et al. 2020). Because these concretions include a significant silicate component in addition to the carbonate matrix (Orr et al. 2000; Saleh et al. 2023), it is considered likely that the concretions formed within an already remineralized volcanic ash matrix that was responsible for molding the fossils. This possibility would explain both the occurrence of fossils outside the centers of concretions, where fossils usually occur, and the preservation of fossils as voids, which is unlike other examples of soft-bodied preservation in concretions (Orr et al. 2000; Siveter et al. 2020). In this view, the Herefordshire fossils would have been preserved via a chemical process very similar to that which preserved the Fezouata radiodonts, although the giant radiodont bodies are not preserved as voids, but are also comprised of the same silica-chlorite-calcite mineral assemblage (Gaines et al. 2012a). The relatively small (centimeter-scale) Herefordshire fossils were swept up into an ash-dominated sediment-gravity flow and rapidly buried, whereas the Fezouata radiodonts may have died and fallen to the seafloor onto a substrate rich in volcanic ash.

The Ediacara biota from Mistaken Point, Newfoundland, is associated with ash deposits and does not fit with typical “Ediacara-type preservation” described later for shallow-marine settings. Like other Ediacara biotas, the Mistaken Point fauna is preserved as molds and casts in sandstone; however, unlike other examples, the Mistaken Point biota occurs in a deep-marine basin, it was preserved via obrution under volcanic ash deposits, and fossils are preserved on the top surface of clastic beds, rather than on the base of obrution beds, as in the other examples described later (Fedonkin 2007). Volcanic ash is important to the biostratinomy of the Mistaken Point assemblages, but a possible role for ash in fossil diagenesis in that deposit remains unclear.

#### Shallow Marine

Shallow-marine settings, defined here as subtidal settings lying above storm wave base, are rarely sites of exceptional preservation, owing to generally well-oxygenated conditions, higher energy, and coarser sediment grain size relative to muddy outer shelf settings (Fig. 1D). A prominent exception to this pattern is the widespread preservation of the Ediacara biota as relatively three-dimensional molds and casts in sandstones.

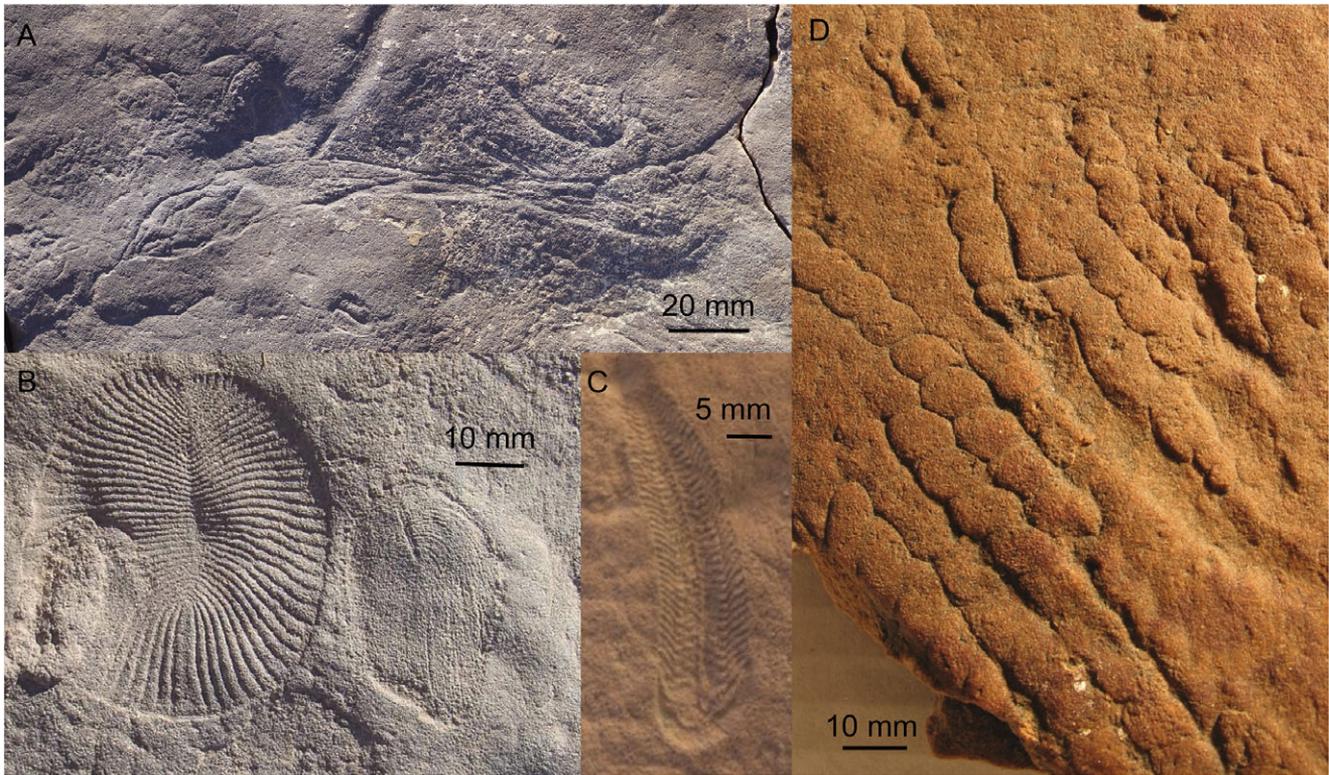
#### Mold and Cast: Ediacara-type Preservation

The Ediacara biota is an entirely soft-bodied suite of fossils, including the oldest known animals, that flourished worldwide during the late Neoproterozoic (Fig. 4). It is overwhelmingly known from enigmatic molds and casts in sandstone (Seilacher 1999), although some examples are known from shale (Xiao et al. 2013) and from carbonates (Xiao et al. 2021). Within sandstones, variation in taphonomic style exists; however, the dominant mode is characterized by the type localities in South Australia, where benthic communities that lived on or floated above a microbial mat were smothered under rapidly deposited sands and are preserved in hyporelief on the soles of the upper, event-deposited beds. Although fine details of external anatomy may be preserved, no internal features are captured by external molds. Like Burgess Shale-type preservation, Ediacara-type preservation (Butterfield 2003) no longer occurs in normal marine settings and therefore was considered an anactualistic mode of preservation by Seilacher.

While the role of phototrophic microbial mats in typical Ediacaran ecosystems was profound (Droser et al. 2022), microbial mats alone could not have induced moldic preservation on the underside of the beds under which they were buried, and thus another mechanism is required. Pyrite veneers were hypothesized to have formed over the surfaces of fossils in the early burial environment and to have acted as a sediment-stabilizing agent that facilitated molding (Gehling 1999). Although dispersed pyrite grains occur in type Ediacara sediments (Liu et al. 2019), they do not form continuous sheets capable of capturing molds (Tarhan et al. 2019). Instead, the anactualistic agent responsible for fossilization is early cementation of the sandstones by silica that precipitated from seawater before the collapse of organic remains, thereby capturing three-dimensional morphology (Tarhan et al. 2016). Experimental evidence further suggests that microbial mats may act as a focus of early diagenetic cementation (Slagter et al. 2022).

#### Molding in Volcanic Ash

A recently discovered example of exceptional preservation in a shallow-marine setting comes from the early Cambrian Tatelt Formation, in which the complete soft-bodied anatomy of trilobites was preserved in volcanic ash, within a graded pyroclastic flow that



**Figure 4.** Fossils of the Ediacara Biota from the Ediacara Member of the Rawnsley Quartzite, Nilpena Ediacara National Park, showing typical preservation of fossils as external molds in sandstones. **A**, Macroscopic alga preserved in negative hyporelief with *Parvancorina* (bottom center). **B**, *Dickinsonia* (left) and *Andiva* (right) preserved in negative hyporelief. **C**, *Spriggina* preserved in negative hyporelief. **D**, *Funisia* preserved largely in positive hyporelief.

entered a nearshore shallow-marine environment (El Albani et al. 2024). Rapid remineralization of the volcanic ash, as described earlier for deep-marine settings, captured the three-dimensional anatomy as hollow external molds, in a mode very similar to that of the Herefordshire biota, described earlier. The expression of these fossils is inconspicuous and provides a target for future exploration.

### Delta Front

Deltas are important transitional environments characterized by the mixing of freshwater with seawater and high sedimentation rates at the locus where terrigenous sediments are introduced to the marine environment. Deltaic environments are favored to form in areas protected from wave energy (Fig. 1A). For all of these reasons, delta front environments provide ecological and taphonomic settings unlike those of other marine settings. Exceptional fossilization may occur in delta front environments through moldic preservation in siderite concretions, which may capture both terrestrial and marine biotas.

### Molding in Siderite Concretions

Early diagenetic siderite forms in shallow burial environments where pore-waters are anoxic and iron oxides occur in excess of calcium and sulfate, favoring the precipitation of iron carbonate (siderite) over calcium carbonate and pyrite (Baird et al. 1986; Vuillemin et al. 2019). Although they may occur in a variety of ferruginous marine settings (e.g., Perrier and Charbonnier 2014), siderite concretions are favored in delta front settings, which are generally dominated by mud delivered from continental environments. Clay minerals formed by oxidative weathering in soils are

commonly associated with particulate iron oxides, rendering deltaic environments particularly iron-rich settings. Particulate iron oxides are readily reduced upon entering an anoxic environment, liberating iron to solution. The physical setting of delta fronts may also favor rapid burial of macrofossils, and localization of decay around carcasses in the shallow burial environment can lead to rapid depletion of oxygen from pore-waters. When optimized, this provides an ideal scenario for the growth of siderite concretions around macrofossils as loci of microbial decomposition, as best known from the Carboniferous Mazon Creek Lagerstätte (Baird et al. 1986; Cotroneo et al. 2016). In that deposit, both terrestrial and marine fossils were fossilized in great detail by rapid molding in siderite (Fig. 2D), with auxiliary preservation by pyrite, calcium phosphate, and calcium carbonate mineralization, and organic preservation of some tissues, including eyes (Clements et al. 2016, 2019; McCoy et al. 2016; Bicknell et al. 2021).

### Tidal Flat

Tidal flat environments are often subject to evaporation, leading to hypersalinity and mineral precipitation (Fig. 1D). In some circumstances, tidal flats can promote exceptional preservation via early mineralization.

### Mold and Cast

Accumulations of cnidarian medusae, interpreted as shoreline strandings, occur in abundance on at least seven bedding surfaces in the late Cambrian Mt. Simon and Wonewoc Formations of Wisconsin (Hagadorn et al. 2002; Fig. 2E), as well as in the early Cambrian of Spain and California (Mayoral et al. 2004; Sappenfield

et al. 2017). These assemblages are preserved as external molds in sandstone and superficially resemble Ediacara-type preservation. While the means of soft-bodied preservation have not been investigated in petrographic or geochemical detail, it is possible that early lithification via evaporative concentration may play a role in sandstone mold and cast-type preservation in tidal flat environments.

### Silica Entombment

Silica is a common evaporatively induced precipitate in carbonate-dominated tidal flat environments of the Proterozoic, where it captures microbial fossils, including those interpreted as cyanobacteria (Fig. 2F). Entombment in silica in this setting was termed “Bitter Springs-type” preservation by Butterfield (2003), who attributed the loss of this mode of fossilization in tidal flats to a secular change in the saturation state of silica in the oceans, driven first by the advent of silica biomineralizing sponges in the early Phanerozoic.

### Anactulistic Fossil Preservation

Seilacher et al. (1985) noted the issue of “anactulistic” preservation from several types of deposits within the rich record of Konservat-Lagerstätten, defining anactulistic preservation as modes of fossilization that no longer occur in comparable settings today. Three of the types of marine preservation described earlier meet this criterion: Bitter Springs-type preservation, Ediacara-type preservation, and Burgess Shale-type preservation. Each of these distinct types has a unique signature in time whereby the means of preservation is lost from the fossil record.

A meaningful counterexample is provided by preservation via entombment within volcanic ash in a marine setting (Siveter et al. 2020; El Albani et al. 2024). Like other types of exceptional preservation, this mechanism is exceedingly rare in the rock record and requires precisely the right conditions, namely an ash-dominated sediment-gravity flow within the marine environment, in order to occur. These conditions, however, are not restricted in time and could occur in favorable settings today. The entire suite of volcanic-associated pathways for exceptional preservation (Fig. 1B) thus represent actualistic means of preservation, as in other fossil Lagerstätten.

Bitter Springs-type preservation, the entombment of microbes in silica in carbonate tidal flat sediments, is an anactulistic mode of fossilization. The evaporative concentration of silica in such settings is thought to have been curtailed by the onset of silica biomineralization by metazoans, which may have permanently limited silica concentrations in the oceans (Butterfield 2003). Early diagenetic silica cementation in normal marine settings (i.e., non-evaporitic) of the Ediacaran, however, appears to require additional explanation.

It is perhaps no coincidence that the two other modes of preservation considered anactulistic are also the two most widespread and abundant pathways for exceptional preservation in the Ediacaran and Cambrian, respectively. It has long been recognized that exceptional fossil deposits are unusually abundant in this interval of time (Seilacher et al. 1985), a feature that is not an artifact of sedimentary rock volume (Allison and Briggs 1993; Muscente et al. 2017; Segesenman and Peters 2022). It is furthermore clear that, although numerous modes of exceptional preservation occur within these two periods (Xiao and Knoll 2000; Maas et al. 2006; Cai et al. 2012; Xiao et al. 2021); statistically, the signal of exceptional preservation in the Ediacaran is dominated by Ediacara-type preservation and that of the Cambrian is dominated by Burgess Shale-type preservation (Gaines 2014; Muscente et al. 2017).

Unlike the conditions that promote exceptional preservation in other types of fossil Lagerstätten, conditions favoring these two styles of preservation were widespread globally in the Ediacaran and Cambrian. During the Ediacaran, Ediacara-type preservation occurred in sandy, normal marine environments ranging from shallow- to deep-water settings where burial conditions were optimized, and in the Cambrian to Early Ordovician, Burgess Shale-type preservation occurred regularly in outer shelf and shelf break settings at the edge of the oxycline (Gaines 2014). Both of these broad environmental settings are common in the later Phanerozoic rock record, but nevertheless, Ediacara-type and Burgess Shale-type preservation both represent “extinct” modes of fossilization in comparable marine settings. The widespread pattern of their distribution in space suggests that the anactulistic circumstances that promoted these means of fossilization were not restricted to unusual, niche environments in the late Neoproterozoic and early Phanerozoic oceans. We posit that the widespread nature of anactulistic fossil preservation in the late Neoproterozoic and early Paleozoic reflect aspects of the global marine conditions in which animals first evolved. In the following sections, we explore conditions of the Ediacaran–early Paleozoic world that played a direct role in exceptional fossilization and may have been important to the earliest metazoan ecosystems during the early evolutionary history of animals.

## The Ediacaran–Cambrian World

### Bioturbation

The widespread organic mats of the Ediacaran were central to the lifestyles of much of the Ediacara Biota (Droser et al. 2022). The increase in trace fossil diversity is well documented through this interval (e.g., Jensen et al. 2006; Mángano and Buatois 2017; Darroch et al. 2021; Cribb et al. 2023) but the development of the mixed layer in sediments and its impacts on ocean and sediment geochemistry and on the physical nature of sediments is more nuanced and drawn out. The advent of bioturbation and subsequent development of the mixed ground has been “blamed” for a wide range of physical and biological changes that occur over the Ediacaran and Cambrian Periods, including the decline of widespread matgrounds (Allison and Briggs 1993; Orr et al. 2003; Darroch et al. 2021; Cribb et al. 2023).

The advent of active mobility is a key innovation in the evolution of animals, with the oldest widely accepted trace fossils associated with the fossils of the Ediacaran White Sea assemblage (Jensen et al. 2006; Evans et al. 2018). *Helminthoidichnites*, made by the bilaterian, *Ikaria* (Evans et al. 2020) occurs at the base of the Ediacara Member of the Rawnsley Quartzite in South Australia. *Ikaria* was a mat grazer, leaving a furrowed trace fossil within mats on the top of the mat surface. *Ikaria* further followed mats under very thin discontinuous sands, leaving furrowed trace fossils preserved in negative relief on the base of thin sandstone beds. While other taxa, such as *Dickinsonia*, *Yorgia*, and *Kimberella* grazed the mat surface, there is no evidence of vertical bioturbation during the reign of the White Sea assemblage. Trace fossil diversity increases into and through the Nama assemblage, as exemplified in the stratigraphic sections of Namibia (Darroch et al. 2021). By the end of the Ediacaran, penetrative burrows in shallow-marine settings were common (e.g., Jensen and Runnegar 2005; Darroch et al. 2021) but did not produce a true mixed layer and did not lead to significant pore-water oxygenation in sediments (Cribb et al. 2023). This trend continued into the early Paleozoic with only a shallow mixed layer until well into the Ordovician (Tarhan et al. 2015; Cribb et al. 2023). The development of extensive grazing and,

ultimately, sediment mixing, along with decreasing silica concentrations in the oceans, led to the closure of the taphonomic window for Ediacara-type preservation (Tarhan et al. 2016).

While the mixed layer was not well developed, the onset of the Phanerozoic is signaled by an increase in the depth and extent of bioturbation (Droser and Bottjer 1989; Mángano and Buatois 2017). This trend of increasing bioturbation has been invoked as a potential cause of the closure of the early Paleozoic Burgess Shale-type “taphonomic window” (Allison and Briggs 1993; Orr et al. 2003); however, outer shelf habitats where Burgess Shale-type preservation occurs did not experience significant sediment mixing or bioirrigation during the Cambrian and Early Ordovician (Tarhan et al. 2016; Cribb et al. 2023). Furthermore, it has been documented that BST preservation, unlike Ediacara-type preservation, requires oxygen-deficient conditions (Gaines and Droser 2010; Gaines et al. 2012b), which exclude bioturbators and most benthos from such settings in modern environments. Indeed, the depth and extent of bioturbation are directly regulated by dissolved oxygen concentrations of bottom waters (Savrdá et al. 1984; Savrdá and Bottjer 1986, 1991). BST fossils therefore occur overwhelmingly in well-laminated, unbioturbated shale intervals inferred to have been deposited under anoxic conditions (Gaines and Droser 2010). Burgess Shale-type settings, however, occur near at the junction of anoxic and oxygenated bottom waters. Because the oxycline fluctuates over time, many BST deposits are characterized by interbedding of unbioturbated shale intervals, which bear exceptional fossils, with weakly bioturbated intervals that do not bear exceptional fossils (Gaines and Droser 2005, 2010; Garson et al. 2012). Thus, the impact of a post-Cambrian increase in the depth and extent of bioturbation may have curtailed the prevalence of this pathway for soft-bodied preservation in many outer shelf settings by eliminating the possibility for preservation in finely interbedded intervals. However, because large portions of many BST deposits were accumulated under persistently anoxic conditions, including those of the Burgess Shale, Chengjiang, and several of the most prominent BST deposits, an increase in the depth and extent of bioturbation could not have affected preservation in settings comparable to these later in time. Therefore a post-Cambrian increase in the depth and extent of bioturbation could not have been responsible for the loss of the Burgess Shale-type preservation pathway from marine settings (Gaines et al. 2012b).

### Oxygen

Research over the last 50 years has profoundly reshaped our view of global redox evolution across the Ediacaran and early Paleozoic. Historically, it was assumed that molecular oxygen rose to near-modern concentrations during the Neoproterozoic/Cambrian transition, coincident with the rise of complex life (e.g., Berner et al. 2003). Although work in recent decades unquestionably points to a major oxygenation event during the late Neoproterozoic, this work has also revealed that global atmospheric oxygen ( $pO_2$ ) remained at surprisingly low levels (<20% of the present atmospheric concentration) across this interval (Och and Shields-Zhou 2012; Lyons et al. 2021). Critically, recent work has also revealed that  $pO_2$  was unstable across the Ediacaran and Cambrian and experienced repeated pulses and crashes throughout this interval (Gill et al. 2011, 2021; Li et al. 2018; Zhang et al. 2019; Krause et al. 2022). Low global redox conditions and redox instability profoundly shaped the marine environments of early metazoan evolution (Bowyer et al. 2017; Zhang et al. 2019).

While the redox picture for early Phanerozoic oceans is still coming into focus, it is clear that they were characterized by widespread oxygen deficiency and instability (Saltzman et al. 2011; Li et al. 2018; Dahl et al. 2019; Gill et al. 2021; Pruss and Gill 2024). Redox instability in particular has also been proposed as a driver of evolutionary innovation for the earliest Phanerozoic animals (He et al. 2019). As low-lying cratons were progressively flooded during the Cambrian, the connections of rapidly expanding shallow-marine settings in continental interiors to the open ocean also became restricted, amplifying local redox effects (Peters 2007). Furthermore, the productive, nutrient-rich settings that sustained early animal communities also imposed an oxygen demand in the water column and at the seafloor, resulting in habitat fragmentation and barriers to migration of benthos (Hammarlund et al. 2017; Guilbaud et al. 2018).

As outlined earlier, oxygen also played an essential role in providing favorable settings for exceptional preservation, particularly in the early Phanerozoic. Although the redox conditions required for Ediacara-type preservation—if any—are not yet clear (Tarhan et al. 2016), widespread benthic anoxia in the early Phanerozoic was an essential prerequisite for Burgess Shale-type preservation, which was favored in outer shelf settings at the junction of anoxic and oxic water masses (Butterfield 1995; Conway Morris 1986; Gaines and Droser 2010), and helps to explain, in part, the statistical prevalence of exceptional preservation in the early Phanerozoic (Gaines 2014; Muscente et al. 2017).

### Tectonics

The tectonic context of the late Neoproterozoic to early Phanerozoic world is characterized by a reorganization of global tectonics in the aftermath of the protracted rifting of the long-lived supercontinent Rodinia (Li et al. 2008). Indeed, much of the Ediacaran sedimentary rock record reflects deposition in rift basins along continental margins (Segessenman and Peters 2022). As widely recognized, most continental cratons were surrounded by passive margins and became clustered around the equator (e.g., Hoffmann and Schrag 2002). A major transition in global tectonics occurred during the Ediacaran, when several cratons collided in the Pan-African orogeny to form the supercontinent Gondwana, while other cratons remained tectonically quiescent, including Baltica, Laurentia, and Siberia (Rino et al. 2008).

While the Ediacara Biota flourished in shallow-marine rift-basin and rifted-margin settings, the Cambrian is characterized by widespread flooding of low-lying cratonic interiors and the rapid expansion of shallow-marine habitats (Peters and Gaines 2012; Peters and Husson 2017). This rapid expansion of shelf area has been linked to the Cambrian explosion of animal diversity (Dalziel 2014). During middle early Cambrian time (stage 3), the Pan-African orogen began to undergo gravitational collapse, leading to another large-scale plate tectonic reconfiguration around some margins of the supercontinent, where subduction was initiated or enhanced, resulting in uplift and loss of continental margin shelf area (Myrow et al. 2024).

Environments in which Burgess Shale-type deposits of early Paleozoic age formed were established following flooding and the development of outer shelf conditions on the continents, where the chemocline intersected the seafloor. The flooding of broad, rapidly subsiding continental margins was key to the widespread development of such conditions across passive margins.

### Climate

The base of the Ediacaran period is marked by the abrupt termination of “Snowball Earth” conditions and the transition to a greenhouse world (Hoffman and Schrag 2002). Greenhouse conditions subsided by the mid-Ediacaran Gaskiers glaciation, which was rapidly followed by the first fossil records of the Ediacara Biota, on the Avalon Peninsula (Pu et al. 2016). Additional geologic evidence from the cold-water carbonate mineral glendonite points to mid-Ediacaran cooling (Wang et al. 2020).

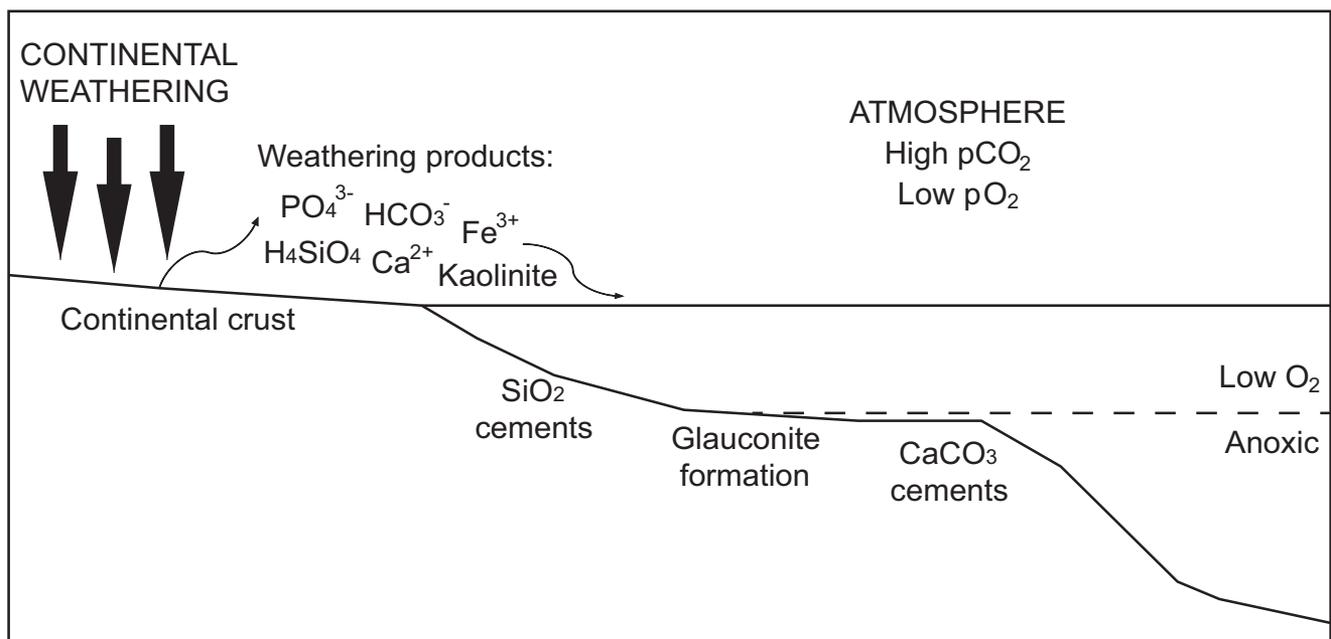
Geologic evidence and carbon cycle models indicate a return to hothouse conditions by the early Cambrian, with  $p\text{CO}_2$  likely in the range of 16–32X PAL (preindustrial atmospheric level) (Berner 2006; Wong Hearing et al. 2018, 2021). Greenhouse conditions that were maintained across the Cambrian were sustained by volcanic outgassing (McKenzie et al. 2014). Detrital zircon abundance, assessed through compilation, suggests a Phanerozoic high in arc volcanism during the Cambrian (McKenzie et al. 2016) that matches well with geologic observations of elevated  $p\text{CO}_2$  and greenhouse climate (Wong Hearing et al. 2021).

Records of continental weathering confirm that the Cambrian greenhouse was accompanied by extensive continental weathering, which in the absence of land plants was driven by elevated  $p\text{CO}_2$  (Driese et al. 2007; Medaris et al. 2018; Colwyn et al. 2019; Wong Hearing et al. 2021). Lithium isotope evidence from shales suggests that the early Cambrian in particular was a time of intense weathering and clay formation (Wei et al. 2024). High surface temperatures may also have led to sluggish oceanic circulation and overturn, particularly in epicratonic seas, where many episodes of the Cambrian are associated with the development of anoxic conditions (Peters 2007; Gill et al. 2011; Zhuravlev et al. 2023).

### Widespread Exceptional Preservation and the Ediacaran–Cambrian World

Both Ediacara-type and Burgess Shale-type preservation are a consequence of the physical, biological, and chemical world during the rise of animal life. Both styles of preservation have been linked to early cementation of sediments. In the Ediacaran, early lithification by seawater-derived silica cements captured soft-bodied biotas as molds in sandstone (Tarhan et al. 2016), whereas in Burgess Shale-type preservation, seawater-derived calcite cements formed crusts over the seafloor following obrution events, restricting exchange of pore-waters with oxidant-bearing seawater, thereby curtailing microbial activity and resulting in the conservation of organic remains of soft-bodied fossils (Gaines et al. 2012c). In addition, there is evidence that an abundance of the clay mineral kaolinite in Burgess Shale-type settings played a role in helping to stabilize organic remains against decomposition (Wilson and Butterfield 2014; Anderson et al. 2018).

The features that promoted these types of exceptional preservation, the prevalence of dissolved silica and carbonate and the abundance of the clay mineral kaolinite in the oceans, all share a common origin (Fig. 5). Kaolinite is produced by intense chemical weathering of continental crust, which releases silicic acid, calcium, and bicarbonate to the oceans. Earth system conditions in the late Neoproterozoic and Cambrian appear to have been primed to promote the delivery of all three of these weathering products to the oceans. Chemical weathering is limited by exposure of basement rock. The Ediacaran–Cambrian interval is characterized by extensive exposure of igneous and metamorphic basement rock that is unique in at least the last 900 Myr of Earth history (Peters and Gaines 2012). All available evidence indicates an intense greenhouse climate followed the Snowball Earth epoch of the Cryogenian and,



**Figure 5.** Cartoon diagram illustrating factors important to widespread exceptional fossil preservation in the Ediacaran and Cambrian Periods. Intense chemical weathering of igneous and metamorphic basement rock under a high  $p\text{CO}_2$  atmosphere resulted in an elevated flux of weathering products via rivers to the oceans. Key weathering products include the nutrients  $\text{Fe}^{3+}$  and  $\text{PO}_4^{3-}$ , silicic acid ( $\text{H}_4\text{SiO}_4$ ), calcium ( $\text{Ca}^{2+}$ ), and bicarbonate ( $\text{HCO}_3^-$ ), from which silica and calcium carbonate cements precipitated, and the clay mineral kaolinite. Low atmospheric  $p\text{O}_2$  and greenhouse climates resulted in widespread oxygen-deficient conditions on marine shelves.

following the Gaskiers glaciation in the mid-Ediacaran, was maintained across the Cambrian Period (Wong Hearing et al. 2018, 2021; Wei et al. 2024). Atmospheric losses of CO<sub>2</sub> to continental weathering were offset by extensive volcanic outgassing around the margins of Gondwana associated with the pan-African orogeny (Tasistro-Hart and Macdonald 2023). Macrostratigraphic facies data and geochemical proxies also support intensive chemical weathering during this time (Peters and Gaines 2012).

In addition to promoting the early lithification of sediments in the Ediacaran and Cambrian, an ocean rich in products of chemical weathering of the continental crust, in both dissolved and clastic form, may have held additional significance for early animals and their ecosystems. An influx of dissolved minerals to the oceans has been linked to the advent of biomineralization (Peters and Gaines 2012; Wood 2018). Chemical weathering also liberates iron and phosphorous, two of the three major nutrients limiting to primary productivity, from silicate minerals. During weathering, iron and phosphorous are retained in soils, often adhering to the charged surfaces of clay minerals, including kaolinite. The delivery of kaolinite to the oceans in significant quantities was thus accompanied by the delivery of nutrients, stimulating the primary productivity that sustained early metazoan ecosystems (Hammarlund et al. 2017). The abundance of glauconite in Cambrian marine sediments also resulted directly from the delivery of kaolinite and iron (Peters and Gaines 2012). Low-oxygen conditions of the late Neoproterozoic–Cambrian, which played a significant role in the distribution of exceptional preservation across this interval (Muscente et al. 2017), also would have impacted early metazoan ecosystems (Hammarlund et al. 2017; Evans et al. 2018, 2022).

It is meaningful that, after their decline in open-marine settings, both type Ediacara-like preservation as molds and casts in sandstones and type Burgess Shale-like carbonaceous compression fossils occur in other types of environments that favor early lithification. Mold and cast preservation on tidal flats (Hagadorn et al. 2002; Mayoral et al. 2004; Sappenfield et al. 2017) may have resulted from early cementation of shoreline sands under evaporative conditions. Carbonaceous preservation of soft-bodied animals in the post-Cambrian occurs most prominently in alkaline stratified lake systems (Fig. 1A) prone to inorganic cementation. Both examples are informative, as they point to the unique nature of ocean chemistry during the late Neoproterozoic to early Phanerozoic. Study of fossil Lagerstätten has helped to reveal that animals evolved and first proliferated in oceanic environments prone to early cementation, high productivity, shelf oxygen deficiency, and habitat fragmentation.

**Acknowledgments.** We are grateful to T. Clements, J. Schiffbauer, an anonymous reviewer, and editor M. Patzkowsky, who provided thoughtful and substantive comments that strengthened the article considerably. We thank W. Weyland for drafting artwork in Figure 1 and R. Bicknell, D. Briggs, P. Cohen, D. Fu, J. Hagadorn, J. Utrup, and D. Fu for photographs used in Figures 2 and 3. R.R.G. acknowledges support from Pomona College and from NSF EAR-1554897.

**Competing Interests.** The authors declare no competing interests.

## Literature Cited

Allison, J., S. Bernard, C. Le Guillou, D. Daval, F. Skouri-Panet, M. Kuga, and F. Robert. 2017. Organic molecular heterogeneities can withstand diagenesis. *Scientific Reports* 7:1508.

Allison, P. A. 1988a. The role of anoxia in the decay and mineralization of proteinaceous macro-fossils. *Paleobiology* 14:139–154.

Allison, P. A. 1988b. Konservat-Lagerstätten: cause and classification. *Paleobiology* 14:331–344.

Allison, P. A., and D. E. G. Briggs. 1993. Exceptional fossil record: distribution of soft-tissue preservation through the Phanerozoic. *Geology* 21:527–530.

Anderson, R. P., N. J. Tosca, R. R. Gaines, N. Mongiardino Koch, and D. E. G. Briggs. 2018. A mineralogical signature for Burgess Shale-type fossilization. *Geology* 46:347–350.

Anderson, E. P., J. D. Schiffbauer, S. M. Jacquet, J. C. Lamsdell, J. Kluesendorf, and D. G. Mikulic. 2021. Stranger than a scorpion: a reassessment of *Parioscorpio venator*, a problematic arthropod from the Llandoveryan Waukesha Lagerstätte. *Palaeontology* 64:429–474. <https://doi-org.cclidm.oclc.org/10.1111/pala.12534>

Aria, C., T.-Y. S. Park, R. R. Gaines, and J. Vannier. 2023. Interpreting fossilized nervous tissues. *BioEssays* 45:2200167.

Ayris, P., and P. Delmelle. 2012. Volcanic and atmospheric controls on ash iron solubility: a review. *Physics and Chemistry of the Earth, parts A/B/C* 45:103–112.

Baird, G. C., S. D. Sroka, C. W. Shabica, and G. J. Kuecher. 1986. Taphonomy of Middle Pennsylvanian Mazon Creek area fossil localities, northeast Illinois: significance of exceptional fossil preservation in syngenetic concretions. *Palaios* 1:271–285.

Behrensmeyer, A. K., S. M. Kidwell, and R. A. Gastaldo. 2000. Taphonomy and paleobiology. *Paleobiology* 26:103–147.

Berner, R. A. 2006. GEOCARBSULF: a combined model for Phanerozoic atmospheric O<sub>2</sub> and CO<sub>2</sub>. *Geochimica et Cosmochimica Acta* 70:5653–5664.

Berner, R. A., D. J. Beerling, R. Dudley, J. M. Robinson, and R. A. Wildman. 2003. Phanerozoic atmospheric oxygen. *Annual Review of Earth and Planetary Sciences* 31:105–134.

Bicknell, R. D., J. Ortega-Hernández, G. D. Edgecombe, R. R. Gaines, and J. R. Paterson. 2021. Central nervous system of a 310-my-old horseshoe crab: expanding the taphonomic window for nervous system preservation. *Geology* 49:1381–1385.

Bowyer, F., R. A. Wood, and S. W. Poulton. 2017. Controls on the evolution of Ediacaran metazoan ecosystems: a redox perspective. *Geobiology* 15:516–551.

Brett, C. E., and G. C. Baird. 1986. Comparative taphonomy: a key to paleoenvironmental interpretation based on fossil preservation. *Palaios* 1:207–227.

Briggs, D. E. 1995. Experimental taphonomy. *Palaios* 10:539–550.

Briggs, D. E. 2003. The role of decay and mineralization in the preservation of soft-bodied fossils. *Annual Review of Earth and Planetary Sciences* 31:275–301.

Briggs, D. E. 2023. The taphonomy of Konservat-Lagerstätten—now and next. *Fossilagerstätten and Taphonomy* 23:8.

Briggs, D. E., and A. J. Kear. 1993a. Decay and preservation of polychaetes: taphonomic thresholds in soft-bodied organisms. *Paleobiology* 19:107–135.

Briggs, D. E., and A. J. Kear. 1993b. Fossilization of soft tissue in the laboratory. *Science* 259:1439–1442.

Briggs, D. E., and S. McMahon. 2016. The role of experiments in investigating the taphonomy of exceptional preservation. *Palaeontology* 59:1–11.

Briggs, D. E., and P. R. Wilby. 1996. The role of the calcium carbonate-calcium phosphate switch in the mineralization of soft-bodied fossils. *Journal of the Geological Society* 153:665–668.

Briggs, D. E. G., S. H. Bottrell, and R. Raiswell. 1991. Pyritization of soft-bodied fossils: Beecher's Trilobite Bed, Upper Ordovician, New York State. *Geology* 19:1221–1224.

Butterfield, N. J. 1995. Secular distribution of Burgess-Shale-type preservation. *Lethaia* 28:1–13.

Butterfield, N. J. 2002. *Leandroilia* guts and the interpretation of three-dimensional structures in Burgess Shale-type fossils. *Paleobiology* 28:155–171.

Butterfield, N. J. 2003. Exceptional fossil preservation and the Cambrian explosion. *Integrative and Comparative Biology* 43:166–177.

Cai, Y., J. D. Schiffbauer, H. Hua, and S. Xiao. 2012. Preservation modes in the Ediacaran Gaojiashan Lagerstätte: pyritization, aluminosilicification, and carbonaceous compression. *Palaeogeography, Palaeoclimatology, Palaeoecology* 326:109–117.

Chiappe, L. M. 2009. Downsized dinosaurs: the evolutionary transition to modern birds. *Evolution: Education and Outreach* 2:248–256.

Clements, T., and S. Gabbott. 2022. Exceptional preservation of fossil soft tissues. Pp.1–10 in *Encyclopedia of Life Sciences*. <https://doi.org/10.1002/9780470015902.a0029468>.

- Clements, T., A. Dolocan, P. Martin, M. A. Purnell, J. Vinther, and S. E. Gabbott. 2016. The eyes of *Tullimonstrum* reveal a vertebrate affinity. *Nature* 532:500–503.
- Clements, T., M. Purnell, and S. Gabbott. 2019. The Mazon Creek Lagerstätte: a diverse late Paleozoic ecosystem entombed within siderite concretions. *Journal of the Geological Society* 176:1–11.
- Clements, T., M. A. Purnell, and S. Gabbott. 2022. Experimental analysis of organ decay and pH gradients within a carcass and the implications for phosphatization of soft tissues. *Palaeontology* 65:e12617.
- Colwyn, D. A., N. D. Sheldon, J. B. Maynard, R.R. Gaines, A. Hofman, X. Wang, B. Gueguen, D. Asael, C.T. Rienhard, and N.J. Planavsky. 2019. A paleosol record of the evolution of Cr redox cycling and evidence for an increase in atmospheric oxygen during the Neoproterozoic. *Geobiology* 17: 579–593. <https://doi-org.ccl.idm.oclc.org/10.1111/gbi.12360>
- Conway Morris, S. C. 1986. The community structure of the Middle Cambrian phyllopod bed (Burgess Shale). *Palaeontology* 29:423–467.
- Cotroneo, S., J. D. Schiffbauer, V. E. McCoy, U. G. Wortmann, S. A. F. Darroch, Y. Peng, and M. Laflamme. 2016. A new model of the formation of Pennsylvanian iron carbonate concretions hosting exceptional soft-bodied fossils in Mazon Creek, Illinois. *Geobiology* 14:543–555.
- Cribb, A. T., S. J. van de Velde, W. M. Berelson, D. J. Bottjer, and F. A. Corsetti. 2023. Ediacaran–Cambrian bioturbation did not extensively oxygenate sediments in shallow marine ecosystems. *Geobiology* 21:435–453.
- Dahl, T. W., E. U. Hammarlund, A. D. Anbar, D. P. Bond, B. C. Gill, G. W. Gordon, A. H. Knoll, A. T. Nielsen, N. H. Schovsbo, and D. E. Canfield. 2010. Devonian rise in atmospheric oxygen correlated to the radiations of terrestrial plants and large predatory fish. *Proceedings of the National Academy of Sciences USA* 107:17911–17915.
- Dahl, T. W., M.-L. Siggaard-Andersen, N. H. Schovsbo, D. O. Persson, S. Husted, I. W. Hougård, A. J. Dickson, K. Kjær, and A. T. Nielsen. 2019. Brief oxygenation events in locally anoxic oceans during the Cambrian solves the animal breathing paradox. *Scientific Reports* 9:11669.
- Dalziel, I. W. 2014. Cambrian transgression and radiation linked to an Iapetus-Pacific oceanic connection? *Geology* 42:979–982.
- Darroch, S. A., A. T. Cribb, L. A. Buatois, G. J. Germs, C. G. Kenchington, E. F. Smith, H. Mocke, G. R. O’Neil, J. D. Schiffbauer, and K. M. Maloney. 2021. The trace fossil record of the Nama Group, Namibia: exploring the terminal Ediacaran roots of the Cambrian explosion. *Earth-Science Reviews* 212:103435.
- Darroch, S. A. F., M. Laflamme, J. D. Schiffbauer, and D. E. G. Briggs. 2012. Experimental formation of a microbial death mask. *Palaios* 27:293–303.
- Decombeix, A.-L., C. J. Harper, C. Prestianni, T. Durieux, M. Ramel, and M. Krings. 2023. Fossil evidence of tylosis formation in Late Devonian plants. *Nature Plants* 9:695–698.
- Demaris, P. J. 2000. Formation and distribution of coal balls in the Herrin Coal (Pennsylvanian), Franklin County, Illinois Basin, USA. *Journal of the Geological Society* 157:221–228.
- Driese, S. G., L. G. Medaris Jr., M. Ren, A. C. Runkel, and R. P. Langford. 2007. Differentiating Pedogenesis from Diagenesis in Early Terrestrial Paleoweathering Surfaces Formed on Granitic Composition Parent Materials *The Journal of Geology* 115(4): 387–406 <https://doi-org.ccl.idm.oclc.org/10.1086/518048>
- Droser, M. L., and D. J. Bottjer. 1989. Ordovician increase in extent and depth of bioturbation: implications for understanding early Paleozoic ecospace utilization. *Geology* 17:850–852.
- Droser, M. L., and J. G. Gehling. 2015. The advent of animals: the view from the Ediacaran. *Proceedings of the National Academy of Sciences USA* 112: 4865–4870.
- Droser, M. L., S. D. Evans, L. G. Tarhan, R. L. Surprenant, I. V. Hughes, E. B. Hughes, and J. G. Gehling. 2022. What happens between depositional events, stays between depositional events: the significance of organic mat surfaces in the capture of Ediacara communities and the sedimentary rocks that preserve them. *Frontiers in Earth Science* 10:826353.
- Duggen, S., N. Olgun, P. Croot, L. Hoffmann, H. Dietze, P. Delmelle, and C. Teschner. 2010. The role of airborne volcanic ash for the surface ocean biogeochemical iron-cycle: a review. *Biogeosciences* 7:827–844.
- El Albani, A., A. Mazurier, G. D. Edgecombe, A. Azizi, C. Elbakhouch, H. O. Berks, E. H. Bouougri, et al. 2024. Rapid volcanic ash entombment resolves the anatomy of Cambrian trilobites. *Science* 384:1429–1435.
- Erwin, D. H., M. Laflamme, S. M. Tweedt, E. A. Sperling, D. Pisani, and K. J. Peterson. 2011. The Cambrian conundrum: early divergence and later ecological success in the early history of animals. *Science* 334:1091–1097.
- Evans, S. D., C. W. Diamond, M. L. Droser, and T. W. Lyons. 2018. Dynamic oxygen and coupled biological and ecological innovation during the second wave of the Ediacara Biota. *Emerging Topics in Life Sciences* 2:223–233.
- Evans, S. D., I. V. Hughes, J. G. Gehling, and M. L. Droser. 2020. Discovery of the oldest bilaterian from the Ediacaran of South Australia. *Proceedings of the National Academy of Sciences USA* 117:7845–7850.
- Evans, S. D., C. Tu, A. Rizzo, R. L. Surprenant, P. C. Boan, H. McCandless, N. Marshall, S. Xiao, and M. L. Droser. 2022. Environmental drivers of the first major animal extinction across the Ediacaran White Sea-Nama transition. *Proceedings of the National Academy of Sciences USA* 119:e2207475119.
- Farrell, Ú. C. 2014. Pyritization of soft tissues in the fossil record: an overview. Pp. 35–58 in M. Laflamme, J. D. Schiffbauer, and S. A. F. Darroch, eds. *Reading and writing of the fossil record: preservational pathways to exceptional fossilization*, Vol. 20. The Paleontological Society, Knoxville, Tenn.
- Farrell, Ú. C., D. E. Briggs, and R. R. Gaines. 2011. Paleoecology of the olenid trilobite *Triarthrus*: new evidence from Beecher’s Trilobite Bed and other sites of pyritization. *Palaios* 26:730–742.
- Farrell, Ú. C., D. E. G. Briggs, E. U. Hammarlund, E. A. Sperling, and R. R. Gaines. 2013. Paleoredox and pyritization of soft-bodied fossils in the Ordovician Frankfort Shale of New York. *American Journal of Science* 313:452–489.
- Fedonkin, M. A. 2007. *The rise of animals: evolution and diversification of the kingdom Animalia*. Johns Hopkins University Press, Baltimore, Md.
- Fu, D., G. Tong, T. Dai, W. Liu, Y. Yang, Y. Zhang, L. Cui, L. Li, H. Yun, and Y. Wu. 2019. The Qingjiang biota—a Burgess Shale-type fossil Lagerstätte from the early Cambrian of South China. *Science* 363:1338–1342.
- Gaines, R., and M. L. Droser. 2005. New approaches to understanding the mechanics of Burgess Shale-type deposits: from the micron scale to the global picture. *Sedimentary Record* 3:4–8.
- Gaines, R. R. 2014. Burgess Shale-type preservation and its distribution in space and time. Pp. 123–146 in M. Laflamme, J. D. Schiffbauer, and S. A. F. Darroch, eds. *Reading and writing of the fossil record: preservational pathways to exceptional fossilization*, Vol. 20. The Paleontological Society, Knoxville, Tenn.
- Gaines, R. R., and M. L. Droser. 2010. The paleoredox setting of Burgess Shale-type deposits. *Palaeogeography, Palaeoclimatology, Palaeoecology* 297:649–661.
- Gaines, R. R., D. E. Briggs, and Z. Yuanlong. 2008. Cambrian Burgess Shale-type deposits share a common mode of fossilization. *Geology* 36:755–758.
- Gaines, R. R., D. E. G. Briggs, P. J. Orr, and P. Van Roy. 2012a. Preservation of giant anomalocaridids in silica-chlorite concretions from the Early Ordovician of Morocco. *Palaios* 27:317–325.
- Gaines, R. R., M. L. Droser, P. J. Orr, D. Garson, E. Hammarlund, C. Qi, and D. E. Canfield. 2012b. Burgess shale-type biotas were not entirely burrowed away. *Geology* 40:283–286.
- Gaines, R. R., E. U. Hammarlund, X. Hou, C. Qi, S. E. Gabbott, Y. Zhao, J. Peng, and D. E. Canfield. 2012c. Mechanism for Burgess Shale-type preservation. *Proceedings of the National Academy of Sciences* 109:5180–5184.
- Garson, D. E., R. R. Gaines, M. L. Droser, W. D. Liddell, and A. Sappenfield. 2012. Dynamic paleoredox and exceptional preservation in the Cambrian Spence Shale of Utah. *Lethaia* 45:164–177.
- Gehling, J. G. 1999. Microbial mats in terminal Proterozoic siliciclastics; Ediacaran death masks. *Palaios* 14:40–57.
- Gill, B. C., T. W. Lyons, S. A. Young, L. R. Kump, A. H. Knoll, and M. R. Saltzman. 2011. Geochemical evidence for widespread euxinia in the Later Cambrian ocean. *Nature* 469:80–83.
- Gill, B. C., T. W. Dahl, E. U. Hammarlund, M. A. LeRoy, G. W. Gordon, D. E. Canfield, A. D. Anbar, and T. W. Lyons. 2021. Redox dynamics of later Cambrian oceans. *Palaeogeography, Palaeoclimatology, Palaeoecology* 581: 110623.
- Guilbaud, R., B. J. Slater, S. W. Poulton, T. H. P. Harvey, J. J. Brocks, B. J. Nettersheim, and N. J. Butterfield. 2018. Oxygen minimum zones in the early Cambrian ocean. *Geochemical Perspectives Letters* 6:33–38.
- Hagadorn, J. W., R. H. Dott Jr., and D. Damrow. 2002. Stranded on a Late Cambrian shoreline: medusae from central Wisconsin. *Geology* 30:147–150.
- Hammarlund, E. U., R. R. Gaines, M. G. Prokopenko, C. Qi, X.-G. Hou, and D. E. Canfield. 2017. Early Cambrian oxygen minimum zone-like conditions at Chengjiang. *Earth and Planetary Science Letters* 475:160–168.

- He, T., M. Zhu, B. J. Mills, P. M. Wynn, A. Y. Zhuravlev, R. Tostevin, P. A. Pogge von Strandmann, A. Yang, S. W. Poulton, and G. A. Shields. 2019. Possible links between extreme oxygen perturbations and the Cambrian radiation of animals. *Nature Geoscience* 12:468–474.
- Hoffman, P. F., and D. P. Schrag. 2002. The snowball Earth hypothesis: testing the limits of global change. *Terra Nova* 14:129–155.
- Iniesto, M., C. Laguna, M. Florin, M. C. Guerrero, A. Chicote, A. D. Buscalioni, and A. I. Lopez-Archilla. 2015. The impact of microbial mats and their microenvironmental conditions in early decay of fish. *Palaio* 30:792–801.
- Jensen, S., and B. N. Runnegar. 2005. A complex trace fossil from the Spitskop Member (terminal Ediacaran–? Lower Cambrian) of southern Namibia. *Geological Magazine* 142:561–569.
- Jensen, S., M. L. Droser, and J. G. Gehling. 2006. A critical look at the Ediacaran trace fossil record. *Neoproterozoic Geobiology and Paleobiology*:115–157.
- Krause, A. J., B. J. W. Mills, A. S. Merdith, T. M. Lenton, and S. W. Poulton. 2022. Extreme variability in atmospheric oxygen levels in the late Precambrian. *Science Advances* 8:eabm8191.
- Li, C., M. Cheng, M. Zhu, and T. W. Lyons. 2018. Heterogeneous and dynamic marine shelf oxygenation and coupled early animal evolution. *Emerging Topics in Life Sciences* 2:279–288.
- Li, Z.-X., Sv. Bogdanova, A. S. Collins, A. Davidson, B. De Waele, R. E. Ernst, I. C. W. Fitzsimons, R. A. Fuck, D. P. Gladkochub, and J. Jacobs. 2008. Assembly, configuration, and break-up history of Rodinia: a synthesis. *Pre-cambrian Research* 160:179–210.
- Liu, A. G., S. McMahon, J. J. Matthews, J. W. Still, and A. T. Brasier. 2019. Petrological evidence supports the death mask model for the preservation of Ediacaran soft-bodied organisms in South Australia. *Geology* 47: 215–218.
- Lyons, T. W., C. W. Diamond, N. J. Planavsky, C. T. Reinhard, and C. Li. 2021. Oxygenation, life, and the planetary system during Earth's middle history: an overview. *Astrobiology* 21:906–923.
- Maas, A., A. Braun, X.-P. Dong, P. C. Donoghue, K. J. Müller, E. Olempska, J. E. Repetski, D. J. Siveter, M. Stein, and D. Waloszek. 2006. The “Orsten”—more than a Cambrian Konservat-Lagerstätte yielding exceptional preservation. *Palaeworld* 15:266–282.
- Mángano, M. G., and L. A. Buatois. 2017. The Cambrian revolutions: trace-fossil record, timing, links and geobiological impact. *Earth-Science Reviews* 173:96–108.
- Mayoral, E., E. Liñán, J. A. G. Vintaned, F. Muñoz, and R. Gozalo. 2004. Stranded jellyfish in the lowermost Cambrian (Corduban) of Spain. *Spanish Journal of Palaeontology* 19:191–198.
- McCoy, V. E. 2014. Concretions as agents of soft-tissue preservation: a review. *Paleontological Society Papers* 20:147–162.
- McCoy, V. E., R. T. Young, and D. E. Briggs. 2015a. Factors controlling exceptional preservation in concretions. *Palaio* 30:272–280.
- McCoy, V. E., R. T. Young, and D. E. Briggs. 2015b. Sediment permeability and the preservation of soft-tissues in concretions: an experimental study. *Palaio* 30:608–612.
- McCoy, V. E., E. E. Saupe, J. C. Lamsdell, L. G. Tarhan, S. McMahon, S. Lidgard, P. Mayer, C. D. Whalen, C. Soriano, and L. Finney. 2016. The “Tully monster” is a vertebrate. *Nature* 532:496–499.
- McKenzie, N. R., N. C. Hughes, B. C. Gill, and P. M. Myrow. 2014. Plate tectonic influences on Neoproterozoic–early Paleozoic climate and animal evolution. *Geology* 42:127–130.
- McKenzie, N. R., B. K. Horton, S. E. Loomis, D. F. Stockli, N. J. Planavsky, and C.-T. A. Lee. 2016. Continental arc volcanism as the principal driver of icehouse-greenhouse variability. *Science* 352:444–447.
- McMahon, S., R. P. Anderson, E. E. Saupe, and D. E. G. Briggs. 2016. Experimental evidence that clay inhibits bacterial decomposers: implications for preservation of organic fossils. *Geology* 44:867–870.
- McNamara, M. E. 2013. The taphonomy of colour in fossil insects and feathers. *Palaontology* 56:557–575.
- McNamara, M. E., P. J. Orr, S. L. Kearns, L. Alcalá, P. Anadón, and E. Peñalver Molla. 2009. Soft-tissue preservation in Miocene frogs from Libros, Spain: insights into the genesis of decay microenvironments. *Palaio* 24:104–117.
- McNamara, M. E., P. J. Orr, L. Alcalá, P. Anadón, and E. Peñalver. 2012. What controls the taphonomy of exceptionally preserved taxa—environment or biology? A case study using frogs from the Miocene Libros Konservat-Lagerstätte (Teruel, Spain). *Palaio* 27:63–77.
- McNamara, M. E., D. E. Briggs, P. J. Orr, N. S. Gupta, E. R. Locatelli, L. Qiu, H. Yang, Z. Wang, H. Noh, and H. Cao. 2013. The fossil record of insect color illuminated by maturation experiments. *Geology* 41:487–490.
- McNamara, M. E., P. J. Orr, S. L. Kearns, L. Alcalá, P. Anadón, and E. Peñalver. 2016. Reconstructing carotenoid-based and structural coloration in fossil skin. *Current Biology* 26:1075–1082.
- McNamara, M. E., F. Zhang, S. L. Kearns, P. J. Orr, A. Toulouse, T. Foley, D. W. E. Hone, C. S. Rogers, M. J. Benton, D. Johnson, X. Xu, and Z. Zhou. 2018. Fossilized skin reveals coevolution with feathers and metabolism in feathered dinosaurs and early birds. *Nature Communications* 9:2072.
- Medaris Jr., L. G., S. G. Driese, G. E. Stinchcomb, J. H. Fournelle, S. Lee, H. Xu, L. DiPietro, P. Gopon, and E. K. Stewart. 2018. Anatomy of a Sub-Cambrian Paleosol in Wisconsin: Mass Fluxes of Chemical Weathering and Climatic Conditions in North America during Formation of the Cambrian Great Unconformity. *The Journal of Geology* 126(3): 261–283, <https://doi.org/ccl.idm.oclc.org/10.1086/697037>
- Murdock, D. J., S. E. Gabbott, G. Mayer, and M. A. Purnell. 2014. Decay of velvet worms (Onychophora), and bias in the fossil record of lobopodians. *BMC Evolutionary Biology* 14:222.
- Muscente, A. D., A. D. Hawkins, and S. Xiao. 2015. Fossil preservation through phosphatization and silicification in the Ediacaran Doushantuo Formation (South China): a comparative synthesis. *Palaogeography, Palaeoclimatology, Palaeoecology* 434:46–62.
- Muscente, A. D., J. D. Schiffbauer, J. Broce, M. Laflamme, K. O'Donnell, T. H. Boag, M. Meyer, A. D. Hawkins, J. W. Huntley, and M. McNamara. 2017. Exceptionally preserved fossil assemblages through geologic time and space. *Gondwana Research* 48:164–188.
- Muscente, A. D., R. C. Martindale, J. D. Schiffbauer, A. L. Creighton, and B. A. Bogan. 2019. Taphonomy of the Lower Jurassic Konservat-Lagerstätte at Ya Ha Tinda (Alberta, Canada) and its significance for exceptional fossil preservation during oceanic anoxic events. *Palaio* 34:515–541.
- Muscente, A. D., O. Vinnes, S. Sinha, J. D. Schiffbauer, E. E. Maxwell, G. Schweigert, and R. C. Martindale. 2023. What role does anoxia play in exceptional fossil preservation? Lessons from the taphonomy of the Posidonia Shale (Germany). *Earth-Science Reviews*:104323.
- Myrow, P. M., J. W. Goodge, G. A. Brock, M. J. Betts, T.-Y. S. Park, N. C. Hughes, and R. R. Gaines. 2024. Tectonic trigger to the first major extinction of the Phanerozoic: the early Cambrian Sinsk event. *Science Advances* 10:ead13452.
- Nanglu, K., J.-B. Caron, and R. R. Gaines. 2020. The Burgess Shale paleocommunity with new insights from Marble Canyon, British Columbia. *Paleobiology* 46:58–81.
- Och, L. M., and G. A. Shields-Zhou. 2012. The Neoproterozoic oxygenation event: environmental perturbations and biogeochemical cycling. *Earth-Science Reviews* 110:26–57.
- Orr, P. J. 2014. Late Proterozoic–early Phanerozoic “taphonomic windows”: the environmental and temporal distribution of recurrent modes of exceptional preservation. *Paleontological Society Papers* 20:289–313.
- Orr, P. J., D. E. Briggs, D. J. Siveter, and D. J. Siveter. 2000. Three-dimensional preservation of a non-biomineralized arthropod in concretions in Silurian volcanoclastic rocks from Herefordshire, England. *Journal of the Geological Society* 157:173–186.
- Orr, P. J., M. J. Benton, and D. E. Briggs. 2003. Post-Cambrian closure of the deep-water slope-basin taphonomic window. *Geology* 31:769–772.
- Paytan, A., and K. McLaughlin. 2007. The oceanic phosphorus cycle. *Chemical Reviews* 107:563–576.
- Perrier, V., and S. Charbonnier. 2014. The Montceau-les-Mines Lagerstätte (Late Carboniferous, France). *Comptes Rendus Palevol* 13:353–367.
- Peters, S. E. 2007. The problem with the Paleozoic. *Paleobiology* 33:165–181.
- Peters, S. E., and R. R. Gaines. 2012. Formation of the “Great Unconformity” as a trigger for the Cambrian explosion. *Nature* 484:363–366.
- Peters, S. E., and J. M. Husson. 2017. Sediment cycling on continental and oceanic crust. *Geology* 45:323–326.
- Pruss, S. B., and B. C. Gill. 2024. Life on the edge: the Cambrian marine realm and oxygenation. *Annual Review of Earth and Planetary Sciences* 52.
- Pu, J. P., S. A. Bowring, J. Ramezani, P. Myrow, T. D. Raub, E. Landing, A. Mills, E. Hodgkin, and F. A. Macdonald. 2016. Dodging snowballs: geochronology of the Gaskiers glaciation and the first appearance of the Ediacaran biota. *Geology* 44:955–958.

- Pulsipher, M. A., E. P. Anderson, L. S. Wright, J. Kluessendorf, D. G. Mikulic, and J. D. Schiffbauer. 2022. Description of *Acheronauta* gen. nov., a possible mandibulate from the Silurian Waukesha Lagerstätte, Wisconsin, USA. *Journal of Systematic Palaeontology*, **20**(1), 1–24. <https://doi-org.ccl.idm.oclc.org/10.1080/14772019.2022.2109216>
- Purnell, M. A., P. J. Donoghue, S. E. Gabbott, M. E. McNamara, D. J. Murdock, and R. S. Sansom. 2018. Experimental analysis of soft-tissue fossilization: opening the black box. *Palaeontology* **61**:317–323.
- Raiswell, R., R. Newton, S. H. Bottrell, P. M. Coburn, D. E. G. Briggs, D. P. G. Bond, and S. W. Poulton. 2008. Turbidite depositional influences on the diagenesis of Beecher's Trilobite Bed and the Hunsrück Slate; sites of soft tissue pyritization. *American Journal of Science* **308**:105–129.
- Rino, S., Y. Kon, W. Sato, S. Maruyama, M. Santosh, and D. Zhao. 2008. The Grenvillian and Pan-African orogens: world's largest orogenies through geologic time, and their implications on the origin of superplume. *Snowball Earth to Cambrian Explosion* **14**:51–72.
- Saleh, F., C. Qi, L. A. Buatois, M. G. Mángano, M. Paz, R. Vaucher, Q. Zheng, X.-G. Hou, S. E. Gabbott, and X. Ma. 2022. The Chengjiang Biota inhabited a deltaic environment. *Nature Communications* **13**:1569.
- Saleh, F., T. Clements, V. Perrier, A. C. Daley, and J. B. Antcliffe. 2023. Variations in preservation of exceptional fossils within concretions. *Swiss Journal of Palaeontology* **142**:20.
- Saltzman, M. R., S. A. Young, L. R. Kump, B. C. Gill, T. W. Lyons, and B. Runnegar. 2011. Pulse of atmospheric oxygen during the late Cambrian. *Proceedings of the National Academy of Sciences USA* **108**:3876–3881.
- Saltzman, M. R., C. T. Edwards, J. M. Adrain, and S. R. Westrop. 2015. Persistent oceanic anoxia and elevated extinction rates separate the Cambrian and Ordovician radiations. *Geology* **43**:807–810.
- Sansom, R. S. 2014. Experimental decay of soft tissues. *Paleontological Society Papers* **20**:259–274.
- Shappenfield, A., L. G. Tarhan, and M. L. Droser. 2017. Earth's oldest jellyfish strandings: a unique taphonomic window or just another day at the beach? *Geological Magazine* **154**:859–874.
- Savrda, C. E., and D. J. Bottjer. 1986. Trace-fossil model for reconstruction of paleo-oxygenation in bottom waters. *Geology* **14**:3–6.
- Savrda, C. E., and D. J. Bottjer. 1991. Oxygen-related biofacies in marine strata: an overview and update. *Geological Society of London Special Publication* **58**: 201–219.
- Savrda, C. E., D. J. Bottjer, and D. S. Gorsline. 1984. Development of a comprehensive oxygen-deficient marine biofacies model: evidence from Santa Monica, San Pedro, and Santa Barbara Basins, California Continental Borderland. *AAPG Bulletin* **68**:1179–1192.
- Schiffbauer, J. D., S. Xiao, Y. Cai, A. F. Wallace, H. Hua, J. Hunter, H. Xu, Y. Peng, and A. J. Kaufman. 2014. A unifying model for Neoproterozoic–Palaeozoic exceptional fossil preservation through pyritization and carbonate compression. *Nature Communications* **5**:5754.
- Scott, A. C., and G. Rex. 1985. The formation and significance of Carboniferous coal balls. *Philosophical Transactions of the Royal Society B* **311**: 123–137.
- Segessenman, D. C., and S. E. Peters. 2022. Macrostratigraphy of the Ediacaran system in North America. In *Laurentia: turning points in the evolution of a continent*. *Geological Society of America Memoir* **220**:399–424.
- Seilacher, A. 1970. Begriff und Bedeutung der Fossil-Lagerstätten [Concept and importance of fossil deposits]. *Neues Jahrbuch für Geologie Und Paläontologie, Monatshefte* **1**:34–39.
- Seilacher, A. 1999. Biomat-related lifestyles in the Precambrian. *Palaios* **14**: 86–93.
- Seilacher, A., W.-E. Reif, and F. Westphal. 1985. Sedimentological, ecological and temporal patterns of fossil Lagerstätten. *Philosophical Transactions of the Royal Society B* **311**:5–24.
- Siveter, D. J., G. Tanaka, U. C. Farrell, M. J. Martin, D. J. Siveter, and D. E. Briggs. 2014. Exceptionally preserved 450-million-year-old Ordovician ostracods with brood care. *Current Biology* **24**:801–806.
- Siveter, D. J., D. E. Briggs, D. J. Siveter, and M. D. Sutton. 2020. The Herefordshire Lagerstätte: fleshing out Silurian marine life. *Journal of the Geological Society* **177**:1–13.
- Slagter, S., W. Hao, N. J. Planavsky, K. O. Konhauser, and L. G. Tarhan. 2022. Biofilms as agents of Ediacara-style fossilization. *Scientific Reports* **12**: 8631.
- Slater, T. S., S. Ito, K. Wakamatsu, F. Zhang, P. Sjövall, M. Jarenmark, J. Lindgren, and M. E. McNamara. 2023. Taphonomic experiments reveal authentic molecular signals for fossil melanins and verify preservation of phaeomelanin in fossils. *Nature Communications* **14**:5651.
- Strullu-Derrien, C., P. Kenrick, and A. H. Knoll. 2019. The Rhynie chert. *Current Biology* **29**:R1218–R1223.
- Tarhan, L. G., M. L. Droser, N. J. Planavsky, and D. T. Johnston. 2015. Protracted development of bioturbation through the early Palaeozoic Era. *Nature Geoscience* **8**:865–869.
- Tarhan, L. G., A. V. Hood, M. L. Droser, J. G. Gehling, and D. E. Briggs. 2016. Exceptional preservation of soft-bodied Ediacara Biota promoted by silica-rich oceans. *Geology* **44**:951–954.
- Tarhan, L. G., A. V. Hood, M. L. Droser, J. G. Gehling, D. E. Briggs, R. R. Gaines, L. J. Robbins, and N. J. Planavsky. 2019. Petrological evidence supports the death mask model for the preservation of Ediacaran soft-bodied organisms in South Australia: comment. *Geology* **47**:e473.
- Tasistro-Hart, A. R., and F. A. Macdonald. 2023. Phanerozoic flooding of North America and the Great Unconformity. *Proceedings of the National Academy of Sciences USA* **120**:e2309084120.
- Trinajstić, K., D. E. Briggs, and J. A. Long. 2022. The Gogo Formation Lagerstätte: a view of Australia's first great barrier reef. *Journal of the Geological Society* **179**:jgs2021–105.
- Varejão, F. G., L. V. Warren, M. G. Simões, F. T. Fürsich, S. A. Matos, and M. L. Assine. 2019. Exceptional preservation of soft tissues by microbial entombment: insights into the taphonomy of the Crato Konservat-Lagerstätte. *Palaios* **34**:331–348.
- Vuillemin, A., R. Wirth, H. Kemnitz, A. M. Schleicher, A. Friese, K. W. Bauer, R. Simister, et al. 2019. Formation of diagenetic siderite in modern ferruginous sediments. *Geology* **47**:540–544.
- Wang, Z., C. Chen, J. Wang, E. Suess, X. Chen, X. Ma, G. Wang, and S. Xiao. 2020. Wide but not ubiquitous distribution of glendonite in the Doushantuo Formation, South China: implications for Ediacaran climate. *Precambrian Research* **338**:105586.
- Wei, G.-Y., M. Zhao, E. A. Sperling, R. R. Gaines, B. Kalderon-Asael, J. Shen, C. Li, et al. 2024. Lithium isotopic constraints on the evolution of continental clay mineral factory and marine oxygenation in the earliest Paleozoic Era. *Science Advances* **10**:eadk2152.
- Wendruff, A. J., L. E. Babcock, J. Kluessendorf, and D. G. Mikulic. 2020. Paleobiology and taphonomy of exceptionally preserved organisms from the Waukesha Biota (Silurian), Wisconsin, USA. *Palaeogeography, Palaeoclimatology, Palaeoecology* **546**:109631.
- Wilby, P. R., D. E. Briggs, P. Bernier, and C. Gaillard. 1996. Role of microbial mats in the fossilization of soft tissues. *Geology* **24**:787–790.
- Wilson, L. A., and N. J. Butterfield. 2014. Sediment effects on the preservation of Burgess Shale-type compression fossils. *Palaios* **29**:145–154.
- Wong Hearing, T., T. H. Harvey, M. Williams, M. J. Leng, A. L. Lamb, P. R. Wilby, S. E. Gabbott, A. Pohl, and Y. Donnadieu. 2018. An early Cambrian greenhouse climate. *Science Advances* **4**:ear5690.
- Wong Hearing, T. W., A. Pohl, M. Williams, Y. Donnadieu, T. H. Harvey, C. R. Scotese, P. Sepulchre, A. Franc, and T. R. Vandenbroucke. 2021. Quantitative comparison of geological data and model simulations constrains early Cambrian geography and climate. *Nature Communications* **12**:3868.
- Wood, R. 2018. Exploring the drivers of early biomineralization. *Emerging Topics in Life Sciences* **2**:201–212.
- Xiao, S., and A. H. Knoll. 2000. Phosphatized animal embryos from the Neoproterozoic Doushantuo formation at Weng'an, Guizhou, South China. *Journal of Paleontology* **74**:767–788.
- Xiao, S., M. Droser, J. G. Gehling, I. V. Hughes, B. Wan, Z. Chen, and X. Yuan. 2013. Affirming life aquatic for the Ediacara biota in China and Australia. *Geology* **41**:1095–1098.
- Xiao, S., Z. Chen, K. Pang, C. Zhou, and X. Yuan. 2021. The Shibantan Lagerstätte: insights into the Proterozoic–Phanerozoic transition. *Journal of the Geological Society* **178**:jgs2020–135.
- Zhang, F., S. Xiao, S. J. Romaniello, D. Hardisty, C. Li, V. Melezhik, B. Pokrovsky, et al. 2019. Global marine redox changes drove the rise and fall of the Ediacara biota. *Geobiology* **17**:594–610.
- Zhuravlev, A. Y., R. A. Wood, and F. T. Bowyer. 2023. Cambrian radiation speciation events driven by sea level and redoxcline changes on the Siberian Craton. *Science Advances* **9**:eadh2558.