

Conference on ‘Malnutrition matters’

Symposium 8: Drugs and nutrition Important drug–nutrient interactions

Pamela Mason

The Rectory, Gwernesney, Usk, Monmouthshire NP15 1HF, UK

Drugs have the potential to interact with nutrients potentially leading to reduced therapeutic efficacy of the drug, nutritional risk or increased adverse effects of the drug. Despite significant interest in such interactions going back to over more than 40 years, the occurrence and clinical significance of many drug–nutrient interactions remains unclear. However, interactions involving drugs with a narrow therapeutic margin such as theophylline and digoxin and those that require careful blood monitoring such as warfarin are likely to be those of clinical significance. Drugs can affect nutrition as a result of changes in appetite and taste as well as having an influence on absorption or metabolism of nutrients. Moreover, foods and supplements can also interact with drugs, of which grapefruit juice and St John’s wort are key examples. Significant numbers of people take both supplements and medication and are potentially at risk from interactions. Professionals, such as pharmacists, dietitians, nurses and doctors, responsible for the care of patients should therefore check whether supplements are being taken, while for researchers this is an area worthy of significant further study, particularly in the context of increasingly complex drug regimens and the plethora of new drugs.

Drugs: Nutrients: Supplements: Interactions: Cytochrome P450 enzymes

A drug–nutrient interaction is considered to be one which results from a physical, chemical, physiological or pathophysiological relationship between a drug and a nutrient present in a food (including an enteral or parenteral feed) or a supplement⁽¹⁾. Drugs and nutrients share several characteristics, including similar sites of absorption in the intestine, the ability to alter physiological processes and the capacity to cause toxicity in high doses⁽²⁾. It is not therefore surprising that drugs can interact with nutrients in several ways. Drugs can potentially influence the bioavailability of nutrients via effects on appetite, absorption, gastrointestinal motility, hepatic metabolism and urinary excretion, while drug absorption and metabolism can sometimes be influenced by nutrients and food supplements⁽³⁾.

Clinical importance

The potential for interactions may appear to be infinite, and it is unclear what proportion of the total has been

identified. More importantly, it is also unclear how many of the identified drug–nutrient interactions are clinically relevant⁽⁴⁾. A drug–nutrient interaction is considered clinically significant if therapeutic response is altered (reduced or enhanced). Such interactions may result in partial or total failure of drug therapy, although the latter is quite rare⁽⁵⁾. Interactions can also cause adverse drug events (e.g. monoamine oxidase inhibitors (MAOI) and certain types of cheese). Such effects can result in the patient discontinuing the drug therapy⁽¹⁾.

Many drug–nutrient interactions, however, are quite harmless, since most drugs are designed to produce blood levels well above those required for therapeutic efficacy. So if a nutrient or food supplement reduces the blood level of a drug, this may not prejudice its clinical effects. Drugs with a narrow therapeutic range (e.g. lithium, phenytoin and theophylline) and those drugs where dosage and blood levels require careful control (e.g. anticoagulants) are those in which drug–nutrient interactions are likely to be the most clinically significant^(3,6).

Patients at risk

The effect of interactions differs from one patient to another with some groups of patients at particular risk⁽²⁾. Infants and children are at particular risk because of the relative inefficiency of the gastrointestinal and hepatic drug metabolising enzymes and poorly developed renal function. Patients on multiple or long-term therapy, who are in an increasing number, are more at risk than patients on short single courses of drugs. Risk of interactions is also increased in patients who are already malnourished because of poor diet and in those with diseases that may lead to nutrient deficiencies (e.g. celiac disease and cystic fibrosis). The risk is also greater in those with increased nutritional requirements (e.g. those with cancer or severe burns).

Mechanistically, there are several types of drug–nutrient interactions. Pharmacokinetic interactions in which a drug can influence the blood concentration of a nutrient or vice versa, often through effects on gastrointestinal absorption, would include the two-way interaction between certain tetracyclines and Ca, and levothyroxine and Ca^(7,8). Pharmacodynamic interactions (e.g. vitamin B₆ and levodopa, folic acid and phenytoin) are those where the interaction influences drug or nutrient action on body systems, often through an effect on enzymes and/or drug receptors⁽³⁾. Another type of interaction is where the drug and nutrient or supplement has similar mechanisms of action (e.g. fish oils and anticoagulants). Many interactions are disadvantageous to the patient, but some are not. Emerging evidence suggests, for example, that antibiotic-associated diarrhoea can be treated with some specific probiotics⁽⁹⁾.

Nutrient intake

Drugs may influence nutrient intake by causing gastrointestinal disturbances or by altering appetite and taste. An enormous number of drugs are associated with nausea and vomiting as a side effect, which can affect desire to eat⁽¹⁰⁾.

Many drugs also lead to changes in appetite^(10,11). Drugs that reduce appetite, such as amantadine, digoxin, fluoxetine, levodopa, lithium, metformin and penicillamine may result in poor nutrition and weight loss, while drugs that increase appetite, such as cyproheptadine, MAOI, tricyclic antidepressants and valproate, may lead to weight gain. When patients complain of poor appetite, a prescribed drug could be one of the causes.

Several drugs may lead to alteration in taste perception (e.g. angiotensin-converting enzyme inhibitors, allopurinol, amiodarone, baclofen, griseofulvin, lithium, metformin, metronidazole, penicillamine and terbinafine)⁽¹²⁾. Taste is mediated by chemosensory nerves that respond to stimulatory chemicals by direct receptor binding, opening ion channels or second messenger channels using nucleotides or phosphorylated inositol^(13,14). Drugs disrupting these cellular processes can cause loss or distortion of taste. Change in taste perception can also be caused by dry mouth. Decreased saliva production alters ion concentrations between saliva and plasma, resulting in decreased taste sensation⁽¹⁵⁾. Antimuscarinic (anticholinergic) drugs (e.g. antihistamines, tricyclic antidepressants, benzatropine,

orphenadrine, oxybutinin, procyclidine, propantheline and trihexyphenidyl hydrochloride) and selegiline cause dry mouth as a side effect⁽¹⁰⁾. Such medication-induced changes can lead to reduced oral intake and weight loss.

Nutrient absorption

Drugs may affect nutrient absorption via several mechanisms. Reduced gastric acid secretion, which can occur as a result of the administration of proton pump inhibitors (e.g. omeprazole) and histamine H₂ antagonists (e.g. cimetidine and ranitidine), can influence the secretion of intrinsic factor and the absorption of vitamin B₁₂⁽¹⁶⁾. In relation to omeprazole, the presence of a polymorphic cytochrome P450 (CYP) enzyme, identified as CYP2C19, significantly affected serum vitamin B₁₂ levels in people on long-term therapy with omeprazole, suggesting that in the future patient genotyping may be useful^(17,18). These drugs, and also antacids, affect gastrointestinal pH, and have been thought to lead to poor absorption of Fe⁽¹⁹⁾. Fe is preferentially absorbed in the ferrous form, but if the pH of the intestine increases, the poorly soluble ferric form is precipitated. However, in a study of 109 patients with Zollinger–Ellison syndrome, continuous treatment with omeprazole for 6 years or continuous treatment with any gastric antisecretory drug for 10 years did not cause decreased body Fe stores or Fe deficiency⁽²⁰⁾. Moreover, an open-label study in 22 patients with Fe deficiency anaemia found that aluminium hydroxide had no significant influence on Fe uptake by the erythrocyte⁽²¹⁾.

Gastric emptying and gastrointestinal motility may be altered by drugs, which in turn can influence nutrient absorption. Gastrointestinal motility can be altered by drugs such as laxatives, metoclopramide, opioids and antimuscarinics. Antimuscarinics and opioids reduce motility, whereas laxatives and metoclopramide increase it⁽²²⁾. A reduction in motility is unlikely to influence nutrient absorption, but an increase in motility may reduce absorption of nutrients⁽¹⁾. Chronic use of stimulant laxatives (e.g. bisacodyl and senna) can lead to depletion of minerals and liquid paraffin causes malabsorption of fat-soluble vitamins⁽¹⁾. Chronic laxative use in older people has also been associated with reduced vitamin B₁₂ status, while use of oat bran as an alternative to laxatives improves vitamin B₁₂ bioavailability⁽²³⁾.

Nutrient absorption may also be influenced by drugs that modulate gastrointestinal mucosal enzymes and transporters. Phenytoin, for example, is hypothesised to interfere with intestinal conjugases that split dietary folates, which are in the polyglutamate form, to the absorbable monoglutamate⁽²⁴⁾. However, findings in relation to this hypothesis have been inconsistent. Studies have shown that phenytoin can reduce the absorption of conjugated folate, whereas there was no decrease in folic acid absorption when phenytoin was administered with the unconjugated form found in multivitamins and fortified foods^(24,25). However, other studies have not confirmed this^(26,27). Phenytoin also acts as a cofactor in the metabolism of folic acid, a less controversial mechanism for the finding that phenytoin reduces folate status⁽²⁸⁾.

Table 1. Grapefruit juice interactions

Bupirone
Ca channel blockers (felodipine, isradipine, lacidipine, lercanidipine, nicardipine, nifedipine, nimodipine, nisoldipine and verapamil)
Carbamazepine
Ciclosporin
Ethinylestradiol
Saquinavir
Sildenafil
Sirolimus and tacrolimus
Simvastatin

Gastrointestinal cytochrome P450 A4 (CYP3A4), present in the epithelial intestinal tissues, plays a role in regulating the oral bioavailability of a large number of drugs and nutrients⁽²²⁾. Functional alteration of CYP3A4, either through induction or through inhibition, can have a profound effect on the amount of nutrients or drug absorbed (i.e. pre-systemic clearance or first-pass metabolism)⁽²⁹⁾.

Grapefruit juice is a classic example of a selective intestinal CYP3A4 inhibitor⁽³⁰⁾. It destroys and deactivates intestinal CYP3A4 enzymes and can increase the bioavailability of some drugs 5-fold^(31–33). Examples of interactions involving grapefruit juice are shown in Table 1. The onset of interaction is immediate with the first glass of grapefruit juice⁽³⁴⁾. The magnitude of enzyme inhibition increases with each glass. Because the interaction involves the destruction of the enzyme, it cannot be corrected by spacing out the doses. On stopping grapefruit juice, increased absorption of the drug is expected to continue for 3–7 d⁽³⁵⁾. There is also evidence that consuming whole grapefruit⁽³⁶⁾, lime juice⁽³⁷⁾ and Seville orange juice⁽³⁸⁾ results in inhibition of the CYP3A4 enzymes and with an impact on the bioavailability of felodipine. In a comparator study with grapefruit juice, citrus-containing soft drinks had no significant impact on ciclosporin metabolism⁽³⁹⁾.

Minerals such as Fe and Zn form insoluble complexes with drugs such as tetracyclines⁽⁴⁰⁾ and 4-quinolones^(41,42). This leads not only to poor absorption of the mineral but also to poor absorption of the drug. Penicillamine, a drug used to chelate excess Cu in the treatment of Wilson's disease, also chelates Fe. Fe has been shown to reduce penicillamine absorption by about two-thirds^(43,44). For optimal absorption of penicillamine, Fe should be given at least 2 h after the penicillamine. This should reduce their admixture in the gut⁽⁴⁴⁾. Absorption of fat-soluble vitamins and folic acid can be reduced by the lipid-lowering drugs, colestyramine and colestipol, which bind bile acids⁽⁴⁵⁾. Bleeding tendency associated with vitamin K malabsorption may increase with these drugs and supplements of fat-soluble vitamins may be prescribed if patients are on long-term treatment⁽⁴⁶⁾.

Nutrient metabolism

Some drugs have so-called antivitamin effects. These include isoniazid, MAOI, methotrexate, phenytoin and

trimethoprim. Isoniazid affects pyridoxine metabolism and may cause peripheral neuropathy, particularly where there are pre-existing risk factors such as diabetes, alcohol dependence, chronic renal failure, malnutrition and HIV infection⁽¹⁰⁾. In these circumstances, pyridoxine (10 mg/d) should be given from the start of treatment⁽¹⁰⁾.

The long-term use of phenytoin and other anticonvulsants can interfere with vitamin D and Ca metabolism and may result in osteomalacia⁽⁴⁷⁾. A small number of reports suggest that people taking phenytoin respond poorly to vitamin D replacement^(48,49) and the dose of vitamin D required to achieve normal plasma 25-hydroxyvitamin D levels in people taking anticonvulsants appears to vary widely⁽⁵⁰⁾. Observational studies also suggest an association between use of anticonvulsant medication, particularly the older drugs such as phenytoin, phenobarbitone, carbamazepine and valproate, and increased metabolism of vitamin D, osteoporosis and fracture^(51,52). All patients taking such medications should have their bone mineral density screened and osteoprotective behaviour such as weight bearing exercise, sunlight exposure, and adequate intakes of Ca and vitamin D should be recommended.

The two-way interaction between phenytoin and folic acid is well known. Phenytoin is a folic acid antagonist, whereas folic acid supplementation can reduce serum phenytoin levels⁽²⁸⁾. Lowering of serum folate by phenytoin has ranged from 27 to 91% and has occurred 6–24 months after starting on phenytoin. Doses of folic acid associated with phenytoin lowering have been in the range of 1–30 mg, rather than the 400 µg dose often taken⁽²⁸⁾. However, women taking phenytoin during pregnancy or when planning a pregnancy are often prescribed a supplement of 5 mg folic acid daily. To date there is limited information on the influence of the newer anti-epileptic medications on folic acid metabolism.

Oral contraceptives have been reported to influence the metabolism of several vitamins, including vitamin A, vitamin C, vitamin B₆ and folic acid⁽⁵³⁾. However, most of these studies are now very old and with the advent of lower dose contraceptives, this interaction may not be significant and there is little justification for women on oral contraceptives taking multivitamins.

Interactions between drugs and dietary supplements

The popularity of over-the-counter food supplements has increased during recent decades, but some supplements may interact with certain drugs. Healthcare professionals, including pharmacists, dietitians, nurses and doctors, should always check whether patients are taking medication and supplements at the same time.

Vitamin B₆ can reduce or abolish the effects of levodopa^(54–56). Dietary intake need not be adjusted as pyridoxine is required for the transformation of levodopa to dopamine, but increased availability of pyridoxine results in excessive transformation of levodopa outside of the brain and the drug fails to reach its target site of action. This interaction is of limited relevance now as levodopa is mostly prescribed in its combination form (e.g. co-beneldopa or co-careldopa). The inclusion of the dopa

decarboxylase inhibitor reduces the wasteful peripheral metabolism of levodopa and much larger amounts are available for entry into the central nervous system. So even in the presence of large amounts of pyridoxine, the peripheral metabolism remains unaffected and the serum levels of levodopa are virtually unaffected⁽⁵⁵⁾.

Supplements containing minerals bind several drugs in the gastrointestinal tract with a consequent reduction in the absorption of both the drug and the mineral. Chelation of levodopa by Fe⁽⁵⁷⁾ can potentially lead to reduced control of Parkinson's disease. Both Fe and Zn form insoluble complexes with several antibiotics, including the tetracyclines⁽⁴⁰⁾ and some of the 4-quinolones (e.g. ciprofloxacin, norfloxacin and ofloxacin)^(41,42), and with penicillamine^(43,44). Because drug absorption is often reduced by more than one mineral, it is wise to separate doses of the drug and mineral preparation by at least 2 h. K supplements should be avoided by patients taking angiotensin-converting enzyme inhibitors (e.g. captopril and enalapril), K-sparing diuretics (e.g. amiloride) and ciclosporin because of a risk of severe hyperkalaemia that can be life threatening⁽¹⁰⁾.

Fish oils contain the long-chain *n*-3 fatty acids, DHA and EPA, which as a result of eicosanoid cascade these particular fatty acids initiate may reduce the coagulability of the blood. This is a potential benefit of fish oil for people at risk of CHD, but many of these patients may also be taking anticoagulants and bleeding tendency may be increased. A case study from Tehran reported that a patient taking warfarin and *n*-3 fatty acids developed a high international normalised ratio, which returned to normal 2 d after the medications were discontinued. The warfarin was then restarted without the *n*-3 fatty acids and the international normalised ratio remained within the normal range⁽⁵⁸⁾. A similar finding came from a US case study⁽⁵⁹⁾, but was not replicated in a patient in an earlier study⁽⁶⁰⁾. Patients taking warfarin and *n*-3 preparations should be carefully monitored.

Some non-nutrient supplements may also interact with drugs. Although a herbal supplement, St John's wort is worthy of particular mention because it is widely used. St John's wort is a potent inducer of CYP450 enzymes, including CYP3A4 that can reduce the bioavailability of various drugs^(61,62). Because the CYP3A4 gene is up-regulated, activity can remain high for weeks after stopping St John's wort. Key drugs with which St John's wort can interact are shown in Table 2. St John's wort is also a potent inducer of P-glycoprotein, an intestinal transporter protein⁽⁶¹⁾. Transporter proteins regulate the rate at which substrates (e.g. drugs or nutrients) are presented to the intestinal metabolising enzymes for regulating the absorption of drugs and nutrients. Some transporter proteins regulate efflux of molecules already absorbed back into the intestinal lumen thereby decreasing bioavailability of some substances. Together, P-glycoprotein and CYP3A4 are the most common regulators affecting oral bioavailability of drugs. For example, ciclosporin absorption is limited by P-glycoprotein efflux and pre-hepatic CYP3A4 enzymes. P-glycoprotein not only regulates how much and how fast ciclosporin is presented to CYP3A4 but also transports some of the absorbed ciclosporin back into the intestinal

Table 2. St John's wort interactions

Anticoagulants (↓ anticoagulant effect)
Antidepressants (↑ serotonergic effect with SSRI)
Antiepileptics
Calcium channel blockers
Cytotoxics (↑ metabolism of irinotecan)
Digoxin (↓ plasma concentration of digoxin)
5HT1 antagonists (↑ serotonergic effect)
Immunosuppressants (↓ plasma concentration of ciclosporin and tacrolimus)
Oral contraceptives (↓ contraceptive effect)
Protease inhibitors (↓ plasma concentration of amprenavir, indinavir, nelfinavir and saquinavir)
Simvastatin (↓ plasma concentration of simvastatin)
Theophylline (↓ plasma concentration of theophylline)

SSRI, selective serotonin re-uptake inhibitor; ↑, increase; ↓, decrease.

lumen⁽⁶³⁾. This provides CYP3A4 with repeated opportunities for metabolism. If this coupled transport metabolism is disrupted (e.g. by grapefruit juice or St John's wort) the absorption of ciclosporin is affected.

Studies evaluating use of supplements and prescribed medicines

Several studies have reported the concomitant use of supplements and prescribed medicines. A US study in 1539 adults found that 44% were taking prescribed medicines and of these 20% were using herbal or high-dose vitamins⁽⁶⁴⁾. A UK study including 164 herbal medicine users found that 59% had taken conventional medicines⁽⁶⁵⁾, while a Canadian study in 195 older patients found that 97% were on prescription medicines and 17% were using natural health products⁽⁶⁶⁾. Studies in cancer patients⁽⁶⁷⁾ and HIV patients⁽⁶⁸⁾ have found that 50–65% use food supplements and/or other complementary medicines. In a US study involving 979 pre-operative patients undergoing anaesthesia, 17.4% reported current use of herbal or dietary supplements⁽⁶⁹⁾.

Studies evaluating the potential for interactions

Further studies have evaluated the potential for interactions among people taking both supplements and medication. A survey among 458 US patients taking prescription medicines found that 197 (43%) were taking supplements, including vitamins, minerals, ginkgo biloba, garlic, saw palmetto and ginseng. Among these patients, 89 (45%) had potential for one or more interactions, of which 6% were potentially serious⁽⁷⁰⁾.

In a further study among cancer patients, supplements were used by 61% (121 patients) with 65 patients (54%) reportedly taking more than one supplement. Risk for interaction was identified in 12% of patients. However, the patient's medical record documented supplement use in only 28% of patients⁽⁷¹⁾.

Supplement use is widespread among cancer patients and longer-term survivors of cancer. In studies combining

different cancer sites, 64–81% of survivors reported using any vitamin or mineral supplements and 26–77% reported using any multivitamins. In contrast, approximately 50% of US adults use dietary supplements and 33% use multi-vitamin/multimineral supplements. Between 14 and 32% of survivors initiate supplement use after diagnosis, and use differs by cancer site. Breast cancer survivors reported the highest use, whereas prostate cancer survivors reported the least. Higher level of education and female sex emerged as factors most consistently associated with supplement use. Up to 68% of physicians are unaware of supplement use among their cancer patients⁽⁷²⁾.

Another study evaluated 1795 patients in six specialty clinics. Among these patients, 39.6% (710) reported use of supplements. In total, 107 interactions with potential clinical significance were identified. Five supplements (garlic, valerian, kava, ginkgo and St John's wort) accounted for 68% of the potentially clinically significant interactions. The four most common classes of prescription medications with a potential for interaction (antithrombotic medications, sedatives, antidepressant agents and anti-diabetic agents) accounted for 94% of the potential clinically significant interactions. No patient was harmed seriously from any interaction⁽⁷³⁾.

In a recent study among 130 older people on the US–Mexico border region, almost half of the older adult participants were at risk for a potential drug–drug interaction, with approximately one-third having a potential interaction between their medications, herbs or nutritional supplements⁽⁷⁴⁾.

Conclusions

Drugs and nutrients share several pharmacokinetic and pharmacodynamic characteristics and can interact according to a variety of mechanisms. Drugs can affect the bioavailability of nutrients, whereas nutrients and supplements can influence the bioavailability of drugs. The theoretic potential for such interactions is almost infinite, but it is unclear how many are clinically relevant. However, the complexity of many drug regimens is sufficient to suggest that patients on such regimens, including older people and those with long-term chronic conditions, may be at greater risk of drug–nutrient interactions than those on single short courses of therapy. Several recent studies have evaluated the numbers of people taking both supplements and medication and found that high proportions, particularly those with conditions such as cancer, take both types of preparation. The risk of serious interactions has been found in 6–12% of patients in these studies. However, clinically relevant data on all potential drug–nutrient interactions have not so far been explored and further research is sorely needed.

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References

- Roe DA (1989) *Diet and Drug Interactions*. New York: Van Nostrand Reinhold.
- Thomas JA (1995) Drug and nutrient interactions. *Nutr Rev* **53**, 271–282.
- Boullata JI & Barber JR (2004) A perspective on drug–nutrient interactions. In *Handbook of Drug–Nutrient Interactions*, pp. 3–27 [JI Boullata and VT Armenti, editors]. New Jersey: Humana Press.
- Santos CA & Boullata JI (2005) An approach to evaluating drug–nutrient interactions. *Pharmacotherapy* **25**, 1789–1800.
- Genser D (2008) Food and drug interaction: consequences for the nutrition/health status. *Ann Nutr Metab* **52**, Suppl. 1, 29–32.
- Leibovitch ER, Deamer RL & Sanderson LA (2004) Food–drug interactions: careful drug selection and patient counseling can reduce the risk in older patients. *Geriatrics* **59**, 19–22, 32–33.
- Mazokopakis EE, Giannakopoulos TG & Starakis IK (2008) Interaction between levothyroxine and calcium carbonate. *Can Fam Physician* **54**, 39.
- Singh N, Singh PN & Hershman JM (2000) Effect of calcium carbonate on the absorption of levothyroxine. *JAMA* **283**, 2822–2825.
- McFarland LV (2006) Meta-analysis of probiotics for the prevention of antibiotic associated diarrhea and the treatment of *Clostridium difficile* disease. *Am J Gastroenterol* **101**, 812–822.
- Martin J (editor) (2010) *British National Formulary*. Number 59. March 2010. London: BMJ Group and RPS Publishing.
- Thomas DR (2009) Anorexia: aetiology, epidemiology and management in older people. *Drugs Aging* **26**, 557–570.
- Doty RL & Bromley SM (2004) Effects of drugs on olfaction and taste. *Otolaryngol Clin North Am* **37**, 1229–1254.
- Roberts CD, Dvoryanchikov G, Roper SD *et al.* (2009) Interaction between the second messengers cAMP and Ca²⁺ in mouse presynaptic taste cells. *J Physiol* **587**, Pt 8, 1657–1668.
- Shimada S, Ueda T, Ishida Y *et al.* (2006) Acid-sensing ion channels in taste buds. *Arch Histol Cytol* **69**, 227–231.
- Matsuo R (2000) Role of saliva in the maintenance of taste sensitivity. *Crit Rev Oral Biol Med* **11**, 216–229.
- Termanini B, Gibril F, Sutliff VE *et al.* (1998) Effect of long-term gastric acid suppressive therapy on serum vitamin B₁₂ levels in patients with Zollinger–Ellison syndrome. *Am J Med* **104**, 422–430.
- Sagar M, Janczewska I, Ljungdahl A *et al.* (1999) Effect of CYP2C19 polymorphism on serum levels of vitamin B₁₂ in patients on long-term omeprazole treatment. *Aliment Pharmacol Ther* **13**, 453–458.
- Ruscin JM, Page RL II & Valuck RJ (2002) Vitamin B₁₂ deficiency associated with histamine(2)-receptor antagonists and a proton-pump inhibitor. *Ann Pharmacother* **36**, 812–816.
- O'Neil-Cutting MA & Crosby WH (1986) The effect of antacids on the absorption of simultaneously ingested iron. *JAMA* **255**, 1468–1470.
- Stewart CA, Termanini B, Sutliff VE *et al.* (1998) Iron absorption in patients with Zollinger–Ellison syndrome treated with long-term gastric acid antisecretory therapy. *Aliment Pharmacol Ther* **12**, 83–98.
- Potgieter MA, Potgieter JH, Venter C *et al.* (2007) Effect of oral aluminium hydroxide on iron absorption from iron(III)-hydroxide polymaltose complex in patients with iron deficiency anemia/a single-centre randomized controlled isotope study. *Arzneimittelforschung* **57**, 392–400.

22. Chan L-N (2006) Drug-nutrient interactions. In *Modern Nutrition in Health and Disease*, pp. 1539–1553 [ME Shils, M Shike, AC Ross, B Caballero and RJ Cousins, editors]. Baltimore and Philadelphia: Lippincott, Williams & Wilkins.
23. Sturtzel B, Dietrich A & Wagner KH (2010) The status of vitamins B₆, B₁₂, folate, and of homocysteine in geriatric home residents receiving laxatives or dietary fiber. *J Nutr Health Aging* **14**, 219–223.
24. Hoffbrand AV & Necheles TF (1968) Mechanism of folate deficiency in patients receiving phenytoin. *Lancet* **2**, 528–530.
25. Rosenberg IH, Godwin HA, Streiff RR *et al.* (1968) Impairment of intestinal deconjugation of dietary folate. A possible explanation of megaloblastic anaemia associated with phenytoin therapy. *Lancet* **2**, 530–532.
26. Houlihan CM, Scott JM, Boyle PH *et al.* (1972) The effect of phenytoin on the absorption of synthetic folic acid polyglutamate. *Gut* **13**, 189–190.
27. Baugh CM & Krumdieck CL (1969) Effects of phenytoin on folic-acid conjugases in man. *Lancet* **2**, 519–521.
28. Berg M (2004) Effects of antiepileptics on nutritional status. In *Handbook of Drug–Nutrient Interactions*, pp. 285–299 [JI Boullata and VT Armenti, editors]. New Jersey: Humana Press.
29. Fujita K (2004) Food–drug interactions via human cytochrome P450 3A (CYP3A). *Drug Metabol Drug Interact* **20**, 195–217.
30. Girenavar B, Jayaprakasha GK & Patil BS (2007) Potent inhibition of human cytochrome P450 3A4, 2D6, and 2C9 isoenzymes by grapefruit juice and its furocoumarins. *J Food Sci* **72**, C417–C421.
31. Dahan A & Altman H (2004) Food–drug interaction: grapefruit juice augments drug bioavailability – mechanism, extent and relevance. *Eur J Clin Nutr* **58**, 1–9.
32. Kiani J & Imam SZ (2007) Medicinal importance of grapefruit juice and its interaction with various drugs. *Nutr J* **6**, 33.
33. Bressler R (2006) Grapefruit juice and drug interactions. Exploring mechanisms of this interaction and potential toxicity for certain drugs. *Geriatrics* **61**, 12–18.
34. Lundahl JU, Regardh CG, Edgar B *et al.* (1998) The interaction effect of grapefruit juice is maximal after the first glass. *Eur J Clin Pharmacol* **54**, 75–81.
35. Lilja JJ, Kivisto KT & Neuvonen PJ (2000) Duration of effect of grapefruit juice on the pharmacokinetics of the CYP3A4 substrate simvastatin. *Clin Pharmacol Ther* **68**, 384–390.
36. Bailey DG, Dresser GK, Kreeft JH *et al.* (2000) Grapefruit–felodipine interaction: effect of unprocessed fruit and probable active ingredients. *Clin Pharmacol Ther* **68**, 468–477.
37. Bailey DG, Dresser GK & Bend JR (2003) Bergamottin, lime juice, and red wine as inhibitors of cytochrome P450 3A4 activity: comparison with grapefruit juice. *Clin Pharmacol Ther* **73**, 529–537.
38. Malhotra S, Bailey DG, Paine MF *et al.* (2001) Seville orange juice–felodipine interaction: comparison with dilute grapefruit juice and involvement of furocoumarins. *Clin Pharmacol Ther* **69**, 14–23.
39. Schwarz UI, Johnston PE, Bailey DG *et al.* (2006) Impact of citrus soft drinks relative to grapefruit juice on ciclosporin disposition. *Br J Clin Pharmacol* **62**, 485–491.
40. Andersson KE, Bratt L, Dencker H *et al.* (1976) Inhibition of tetracycline absorption by zinc. *Eur J Clin Pharmacol* **9**, 131–134.
41. Lehto P, Kivisto KT & Neuvonen PJ (1994) The effect of ferrous sulphate on the absorption of norfloxacin, ciprofloxacin and ofloxacin. *Br J Clin Pharmacol* **37**, 82–85.
42. Polk RE, Healy DP, Sahai J *et al.* (1989) Effect of ferrous sulfate and multivitamins with zinc on absorption of ciprofloxacin in normal volunteers. *Antimicrob Agents Chemother* **33**, 1841–1844.
43. Osman MA, Patel RB, Schuna A *et al.* (1983) Reduction in oral penicillamine absorption by food, antacid, and ferrous sulfate. *Clin Pharmacol Ther* **33**, 465–470.
44. Lyle WH (1976) Penicillamine and iron. *Lancet* **2**, 420.
45. Leonard JP, Desager JP, Beckers C *et al.* (1979) *In vitro* binding of various biological substances by two hypocholesterolaemic resins. Cholestyramine and colestipol. *Arzneimittelforschung* **29**, 979–981.
46. Knodel LC & Talbert RL (1987) Adverse effects of hypolipidaemic drugs. *Med Toxicol* **2**, 10–32.
47. Kulak CA, Borba VZ, Bilezikian JP *et al.* (2004) Bone mineral density and serum levels of 25 OH vitamin D in chronic users of antiepileptic drugs. *Arq Neuropsiquiatr* **62**, 940–948.
48. Asherov J, Weinberger A & Pinkhas J (1977) Lack of response to vitamin D therapy in a patient with hypoparathyroidism under anticonvulsant drugs. *Helv Paediatr Acta* **32**, 369–373.
49. Rubinger D, Korn-Lubetzki I, Feldman S *et al.* (1980) Delayed response to an alpha-hydroxycholecalciferol therapy in a case of hypoparathyroidism during anticonvulsant therapy. *Isr J Med Sci* **16**, 772–774.
50. Collins N, Maher J, Cole M *et al.* (1991) A prospective study to evaluate the dose of vitamin D required to correct low 25-hydroxyvitamin D levels, calcium, and alkaline phosphatase in patients at risk of developing antiepileptic drug-induced osteomalacia. *Q J Med* **78**, 113–122.
51. Lee RH, Lyles KW & Colon-Emeric C (2010) A review of the effect of anticonvulsant medications on bone mineral density and fracture risk. *Am J Geriatr Pharmacother* **8**, 34–46.
52. Nakken KO & Tauboll E (2010) Bone loss associated with use of antiepileptic drugs. *Expert Opin Drug Saf* **2010**, 4.
53. Prasad AS, Oberleas D, Moghissi KS *et al.* (1975) Effect of oral contraceptive agents on nutrients: II. Vitamins. *Am J Clin Nutr* **28**, 385–391.
54. Hsu TH, Bianchine JR, Preziosi TJ *et al.* (1973) Effect of pyridoxine on levodopa metabolism in normal and parkinsonian subjects. *Proc Soc Exp Biol Med* **143**, 578–581.
55. Mars H (1973) Metabolic interactions of pyridoxine, levodopa, and carbidopa in Parkinson's disease. *Trans Am Neurol Assoc* **98**, 241–245.
56. Duvoisin RC, Yahr MD & Cote LD (1969) Pyridoxine reversal of L-dopa effects in Parkinsonism. *Trans Am Neurol Assoc* **94**, 81–84.
57. Greene RJ, Hall AD & Hider RC (1990) The interaction of orally administered iron with levodopa and methyldopa therapy. *J Pharm Pharmacol* **42**, 502–504.
58. Jalili M & Dehpour AR (2007) Extremely prolonged INR associated with warfarin in combination with both trazodone and omega-3 fatty acids. *Arch Med Res* **38**, 901–904.
59. Buckley MS, Goff AD & Knapp WE (2004) Fish oil interaction with warfarin. *Ann Pharmacother* **38**, 50–52.
60. Bender NK, Kraynak MA, Chiquette E *et al.* (1998) Effects of marine fish oils on the anticoagulation status of patients receiving chronic warfarin therapy. *J Thromb Thrombolysis* **5**, 257–261.
61. Izzo AA & Ernst E (2009) Interactions between herbal medicines and prescribed drugs: an updated systematic review. *Drugs* **69**, 1777–1798.
62. Kober M, Pohl K & Efferth T (2008) Molecular mechanisms underlying St. John's wort drug interactions. *Curr Drug Metab* **9**, 1027–1037.

63. Staatz CE, Goodman LK & Tett SE (2010) Effect of CYP3A and ABCB1 single nucleotide polymorphisms on the pharmacokinetics and pharmacodynamics of calcineurin inhibitors: part I. *Clin Pharmacokinet* **49**, 141–175.
64. Eisenberg DM, Davis RB, Ettner SL *et al.* (1998) Trends in alternative medicine use in the United States, 1990–1997: results of a follow-up national survey. *JAMA* **280**, 1569–1575.
65. Gulian C, Barnes J & Francis S (2002) Types and preferred sources of information concerning herbal medicinal products: face to face interviews with users of herbal medicinal products. *Int J Pharm Pract* **10**, Suppl., R33.
66. Dergal JM, Gold JL, Laxer DA *et al.* (2002) Potential interactions between herbal medicines and conventional drug therapies used by older adults attending a memory clinic. *Drugs Aging* **19**, 879–886.
67. Patterson R, Neuhouser M, Hedderson M *et al.* (2002) Types of alternative medicine used by patients with breast, colon, or prostate cancer: predictors, motives, and costs. *J Altern Complement Med* **8**, 477–485.
68. Risa KJ, Nepon L, Justis JC *et al.* (2002) Alternative therapy use in HIV-infected patients receiving highly active anti-retroviral therapy. *Int J STD AIDS* **13**, 706–713.
69. Meyer T, Baisden CE, Roberson CR *et al.* (2002) Survey of preoperative patients use of herbal products and other selected dietary supplements. *Hosp Pharm* **37**, 1301–1306.
70. Peng CC, Glassman PA, Trilli LE *et al.* (2004) Incidence and severity of potential drug–dietary supplement interactions in primary care patients: an exploratory study of 2 outpatient practices. *Arch Intern Med* **164**, 630–636.
71. Lee AH, Ingraham SE, Kopp M *et al.* (2006) The incidence of potential interactions between dietary supplements and prescription medications in cancer patients at a Veterans Administration Hospital. *Am J Clin Oncol* **29**, 178–182.
72. Velicer CM & Ulrich CM (2008) Vitamin and mineral supplement use among US adults after cancer diagnosis: a systematic review. *J Clin Oncol* **26**, 665–673.
73. Sood A, Sood R, Brinker FJ *et al.* (2008) Potential for interactions between dietary supplements and prescription medications. *Am J Med* **121**, 207–211.
74. Loya AM, Gonzalez-Stuart A & Rivera JO (2009) Prevalence of polypharmacy, polyherbacy, nutritional supplement use and potential product interactions among older adults living on the United States–Mexico border: a descriptive, questionnaire-based study. *Drugs Aging* **26**, 423–436.