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Seed persistence and germination strategies of *Carajasia cangae* (Rubiaceae): an endemic forb from the Amazon ironstone outcrop

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Abstract

Seed persistence, desiccation tolerance, and dormancy play a crucial role in plant population and community dynamics. However, these life-history traits remain largely understudied in perennial herbaceous species, particularly in tropical ecosystems. We evaluated the seed storage behaviour, potential longevity, soil seed bank, seed dormancy alleviation in the field and the effects of after-ripening temperature and time on seed dormancy alleviation in Carajasia cangae - an endangered perennial forb endemic to the ironstone outcrops of the Eastern Amazon. We performed germination experiments to examine the effect of storage conditions (-20, 5 and 28°C, as well as field storage) and time on seed viability, mean germination time and percentage. Our results suggested that C. cangae seeds form a transient soil seed bank and show orthodox storage behaviour. The seeds' longevity was favoured in all controlled storage conditions in relation to soil-stored seeds (field). However, the marked loss of seed viability in less than 1 year, regardless of storage condition, indicates a low potential for long-term germplasm conservation through seed banking. Seed dormancy was fully alleviated after 3 months of field storage during the dry season. Moreover, seeds stored for 6 months at 28°C had their dormancy partially alleviated, indicating that environmental conditions found throughout the dry season in the species habitat are required to alleviate its seed dormancy. A transient seed bank type is favoured by predictable seasonal variations in climate in the region, species iteroparity and seed dormancy alleviation during the dry season, which delays germination until the onset of the next rainy season.

Introduction

Seed persistence in the soil and germination timing have significant ecological and evolutionary importance in plant population and community dynamics as they strongly affect population fitness components, such as survival and fecundity (Saatkamp et al., 2014; Long et al., 2015; Yang et al., 2021). Soil seed banks are reservoirs of viable seeds in or on the soil, and their ecological roles depend on seed persistence in the soil, germination patterns over time, the species' life history (e.g. lifespan and reproductive strategy), and environmental conditions (Thompson and Grime, 1979;Saatkamp et al., 2014). Persistent seed banks, in which a fraction of the seed population remains viable in the soil for more than 1 year after dispersal, are viewed as a bet-hedging strategy that spreads the risk of reproductive failure and distributes genetic diversity over time, thereby increasing long-term fitness, population persistence, and biodiversity maintenance (Saatkamp et al., 2014; Long et al., 2015; Gioria et al., 2020; Yang et al., 2021). Alternatively, transient seed banks are those in which seeds persist in the soil up to 1 year after dispersal. In species with transient seed banks, dormancy alleviation and/ or germination cues that signal favourable conditions for seedling establishment ensure seeds germinate under optimal conditions that favour seedling growth and survival (Saatkamp et al., 2014; Yang et al., 2021).

Seed storage behaviour is intrinsically linked to seed persistence in the soil, and both are crucial to understand regeneration ecology and to make informed decisions in both *in situ* management of species and *ex situ* conservation (Wyse and Dickie, 2018; Tweddle et al., 2003). Seeds are classified into three storage categories according to their response to desiccation and storage temperature: orthodox seeds, which tolerate desiccation and display increased seed longevity with negative storage temperatures; intermediate seeds, which also tolerate desiccation but negative storage temperatures reduce seed longevity; and recalcitrant seeds, which do not tolerate desiccation (Hong and Ellis, 1996). Recalcitrant species generally disperse seeds with the onset of the rainy season that either germinate rapidly after dispersal or form short-lived transient seed banks (Daws et al., 2005; Wyse and Dickie, 2018). Instead, desiccation-tolerant species can form persistent or transient soil seed banks according to the species' regeneration niche, evolutionary history, life-history traits (such as seed mass and shape, life



form, reproductive frequency and germination strategy), and the environmental conditions of where the species occur (such as environmental predictability, abiotic stresses and soil type) (Fenner and Thompson, 2005; Wyse and Dickie, 2017; Gioria et al., 2020; Yang et al., 2021; Mašková et al., 2022; Chen et al., 2024).

Seed persistence in the soil is strongly driven by environmental predictability. Environments with unpredictable climate (i.e. high interannual variation), such as deserts, select species with persistent soil seed banks as it spreads seed germination in time, increasing the chances of at least some seeds germinating in favourable years (Saatkamp et al., 2014; Gioria et al., 2020). By contrast, when climates change in a temporally predictable form, such as in seasonal ecosystems, transient rather than persistent soil seed banks tend to form, with germination timing determined by cues for dormancy loss or by germination requirements of nondormant seeds matching the favourable season for growth and survival (Thompson and Grime, 1979; Saatkamp et al., 2014; Escobar et al., 2021). However, persistent banks might be adaptive in seasonal ecosystems when the regeneration opportunities are unpredictable (e.g. gap formation) or as a strategy to cope with high spatial heterogeneity in environmental conditions, sibling competition, and density-dependent mortality (Hyatt and Evans, 1998; Saatkamp et al., 2014; Long et al., 2015; Gioria et al., 2020).

Seed persistence in the soil, desiccation tolerance, and seed dormancy are expected to be negatively correlated with other lifehistory traits, such as iteroparity, long life span, and large seeds that also buffer risks of local extinction caused by reproductive failure (Metcalf et al., 2003; Long et al., 2015). Global-scale analysis shows higher proportions of orthodox seed behaviour, persistent soil bank, and seed dormancy in herbaceous rather than woody species, in annual rather than perennial herbs, and for light rather than heavy seeds (Baskin and Baskin, 2014; Rubio de Casas et al., 2015; Wyse and Dickie, 2017; Gioria et al., 2020; Chen et al., 2024). However, seed dormancy seems to be a poor predictor of seed persistence in the soil and storage behaviour, except for physically dormant seeds, which tend to form persistent seed banks (Gioria et al., 2020; Chen et al., 2024). Nevertheless, these global-scale analyses have a notable bias as seed desiccation responses and soil seed banking strategies in perennial herbaceous species remain deeply neglected, especially in tropical ecosystems (Wyse and Dickie, 2017; Chen et al., 2024).

Here, we evaluated the seed storage behaviour and potential longevity (i.e. ex situ longevity), soil seed bank strategy, seed dormancy alleviation in the field, and the effect of after-ripening temperature and time on seed dormancy alleviation of Carajasia cangae (R.M.Salas, E.L.Cabral & Dessein), an endangered rockdwelling perennial forb of the Amazon ironstone outcrops (Salas et al., 2015; Amorim and Bicalho, 2022). C. cangae is an edaphic specialist and a narrow endemic species of the ironstone outcrops (locally known as canga) that occurs throughout the southern mountain range of the Carajás National Forest (FLONA de Carajás), Eastern Amazon region (Giulietti et al., 2019). Ironstone outcrops are a seasonally dry, grassy-shrubby vegetation mosaic with nutrient-impoverished and shallow soils and a low water holding capacity (Schaefer et al., 2016; Silveira et al., 2016; Miola et al., 2021). Moreover, plant species associated with cangas face high temperatures and high ultraviolet exposure (Jacobi et al., 2007; Viana et al., 2016). This vegetation mosaic shows a huge variation in environmental conditions on a scale of metres, which is mainly determined by soil depth and

landscape roughness (Gastauer et al., 2021). The *canga* vegetation can vary from areas with extremely shallow soils (~5 cm), where the substrate forms a rocky carapace with small soil cavities and herbaceous species are dominant (locally known as *canga couraçada* and referred to as rocky grasslands henceforth), to the *capão de mata*, where soils are deeper than 30 cm and can support small-statured forests (Mota et al., 2015).

We hypothesize that *C. cangae* displays an orthodox seed storage behaviour, as it is a herbaceous, small-seeded species (<1 mm) that release seeds during the rainy-to-dry season transition (Salas et al., 2015; Santos et al., 2023), a period of great abiotic stress. Moreover, mountain-top grasslands and shrublands are dominated by species with orthodox storage behaviour (Wyse and Dickie, 2017). We expect a transient seed bank strategy due to its perennial and iteroparity life-history strategy and reliable yet strongly seasonal climate, with seed dormancy alleviation cues synchronizing germination with the rainy season. However, the restricted distribution of C. cangae to rocky grasslands together with the fine-grain mosaic vegetation could select a persistent seed bank, as the recolonization of a previously occupied safe site is easier to occur from the seed bank in a local patch than in a distant one because of the high environmental heterogeneity on a microscale (Gutterman, 1994; Satterthwaite, 2009; Rubio de Casas et al., 2015). Furthermore, its small seed size and herbaceous life form are usually related to persistent soil banks (Gioria et al., 2020; Chen et al., 2024). This study provides important insights into in situ management and ex situ conservation of C. cangae, showing the species germination strategy and how seed dormancy status changes in the soil seed bank. We also show how seed dormancy of C. cangae seeds can be alleviated and potential longevity increased, thus assisting conservation actions. Moreover, our study contributes to filling the sizable knowledge gap of soil seed persistence in tropical ecosystems.

Methods

Study area and species

The Carajás National Forest, which is located in the Eastern Amazon (Pará, Brazil), is a protected area created in 1998 where vast forests surround mountain-tops that are rich in iron ore, where the ironstone outcrop (canga) ecosystems occur. The canga ecosystems of FLONA of Carajás are distributed throughout the northern mountain range (Serra Norte) and the southern mountain range (Serra Sul; Fig. 1A). The climate is hot and humid (Köppen Aw), with a marked dry season from May to October and a rainy season from November to April, during which most of the annual precipitation occurs (1,800–2,300 mm; Viana et al., 2016). Mean temperatures fluctuate between 20 and 23°C at night and between 23 and 35°C during the day (Fig. 2), but maximum temperatures on the soil surface can reach up to 60°C in rocky grasslands (T. Zupo et al., unpublished data).

C. cangae is a small-sized rock-dwelling perennial forb species (<11 cm in height, Salas et al., 2015). However, many individuals exhibit an annual behaviour, completing their life cycle (recruitment, growth, reproduction and death) within one growing season, but reaching only very small sizes (3–5 cm in height; T. Zupo, *personal communication*). This species has a narrow geographic and ecological distribution, limited to rocky grassland formations of the largest *canga* plateau (S11, approximately 48.1 km² of area, Souza-Filho et al., 2019) in the southern mountain range of the Carajás National Forest (–50.424934, –6.341843; Fig. 1;

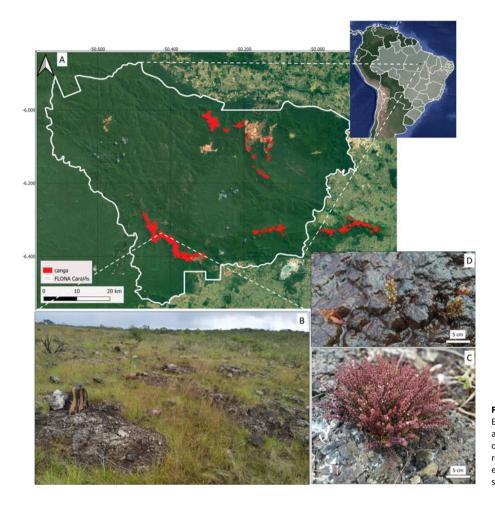


Figure 1. (A) Location of the Carajás National Forest, Eastern Amazon (north of Brazil), where the northern and southern mountain ranges of ironstone outcrops occur. (B) Habitat of *C. cangae*, where it grows over rocks of the rocky grassland formations in the southern mountain range of Carajás. (C) A large and (D) small mature individuals of *C. cangae* during flowering.

Costa et al., 2023). Seeds are 0.7-0.75 mm long, with a fresh seed mass of 0.0729 ± 0.0028 mg (mean \pm SD) (n=16 groups of 100 seeds, weighed using an analytical balance – Mettler Toledo, MS204S; Columbus, OH, USA), and they have no apparent dispersal structures. Seeds are dispersed between the end of the

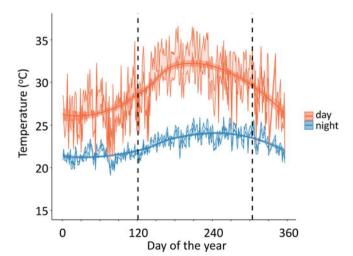


Figure 2. Temperature (mean ± se) during the day (6 AM to 6 PM) and night (6 PM to 6 AM). Dashed vertical lines represent the beginning and end of the dry season. Temperature values were extracted from two Hobo MX2301A sensors placed 30 cm above the soil in the area where *C. cangae* occurs. Sensors registered air temperature every hour for 1 year (December 2022 to December 2023).

rainy season and the onset of the dry season (April to June) and exhibit physiological dormancy (Salas et al., 2015; Zanetti et al., 2020; Santos et al., 2023; Boanares et al., 2024).

Seed harvest

Mature fruits were collected in May 2022 from at least 30 individuals. Permits to sample fruits were granted by SISBIO (Sistema de Autorização e Informação em Biodiversidade, permission number 76784-1). Fruits were put into paper bags and taken to the laboratory, where they were put together and sorted to avoid empty seeds. Fresh seeds were stored in room conditions (around 25°C and 50% relative humidity) for 14 days prior to the germination and *ex situ* storage experiments and for 30 days prior to the *in situ* storage experiments (see details below).

Seed storage and germination experiments

We assessed seed longevity and the alleviation of seed dormancy, both ex-situ and in-situ, by comparing the germination and viability of fresh and dry seeds with those stored under controlled and field conditions over time. We performed two germination experiments, one to determine ex situ and another to determine in situ seed longevity. To determine ex situ seed longevity, we used a factorial design with three storage temperatures (-20, 5 and 28° C) and three storage times (92, 187, and 227 days); seeds were also stored for 365 days at -20° C. We were only able to store seeds for 365 days at -20° C due to seed availability.

To determine *in situ* seed longevity, we used three storage periods (108, 223 and 339 days) and one storage condition (field condition). The storage periods correspond to the end of the dry season, the first half of the rainy season and the beginning of the next dry season.

Fresh and dried seeds were used as the controls. The same controls were used for both experiments given the same seed lot was used for both *in situ* and *ex situ* germination experiments (see details below). For the fresh seed control, we used recently harvested seeds kept for 14 days at 23°C and 50% RH before the germination experiments. For the dried seed control, fresh seeds were placed in a desiccator containing silica gel (10–12 RH%) at room temperature (23°C) for 50 days. We used the seed equilibrium relative humidity (eRH%) as a measure of the seed moisture status of both fresh and dried seeds. The eRH was determined at 23°C by a hygrometer (HOBO UX 100-023) within a sealed glass chamber. For such, we placed a sample of seeds into the hygrometer chamber and allowed 60–90 min to reach equilibrium with the relative air humidity. The seed sample occupied at least 20% of the chamber's total volume (Probert et al., 2003).

Seeds stored ex situ were firstly dried for 50 days at 10-12 RH% and 23°C. They were then placed in tightly capped 2 mL polypropylene cryovials, wrapped in aluminium foil and stored at -20, 5 and 28°C; each cryovials contained 120 seeds. At each storage period (92, 187, 227 and 365 days), a cryovial per temperature was removed, and seeds were tested for germination and viability as described below. To determine in situ longevity, we placed fresh seeds (kept for 30 days at 50% RH and 23°C) in nylon mesh bags of approximately 15 × 5 cm and 1 mm mesh size (30 seeds per bag), which were then placed on the rocky surface of the area where a population of C. cangae naturally occurs. To prevent the bags from dispersing, we tied them to the rocks with galvanized iron wire (Fig. 1B). Three bags (one per storage period) were placed in one of six locations (replicates) distributed along the area. At each storage period (108, 223 and 339 days), a bag was removed from each location (total of six bags/storage period) and taken to the laboratory. Seeds were then retrieved and tested for germination and viability, as described below.

The germination tests were carried out in Petri dishes containing filter paper saturated with distilled water and incubated at alternating temperatures of 24/32°C with a 12/12 h photoperiod (\sim 50 μ mol s⁻¹ m⁻² photosynthetically active radiation). These temperatures were selected as they correspond to the approximate mean day and night temperatures (Fig. 2). For the controls and ex situ storage, six replicates (Petri dish) of 20 seeds each (n = 120)were used, while, for the in-situ storage, the number of seeds per Petri dish varied among storage periods due to the availability of seeds retrieved from the mesh bags (4-29 seeds, Supplementary Table S1). The experiments were monitored once a week for 90 days, and radicle protrusion was used as the germination criterion. At the end of the experiment, the viability of non-germinated seeds was analyzed by means of the tetrazolium test using 1% solution at 37 ± 2 °C for 72 h incubation. We extended the incubation period to 72 h and increased the temperature above the recommended levels for crop species (up to 48 h and approximately 30°C, BRASIL, 2009) to facilitate the penetration of the tetrazolium salt into the embryo. Following incubation, seeds were longitudinally sectioned using a stereoscope (ZEISS Discovery.V12) at 27.2× magnification. Seeds were classified as viable if all embryonic structures were stained bright red (Supplementary Figure S1).

Data analysis

Seed storage behaviour and seed longevity

Seed storage behaviour was determined based on Hong and Ellis (1996). First, we determined the seed desiccation tolerance by comparing the viability of fresh and dried seeds through a general linear model (GLM) using a binomial distribution. Then, we used the GLM to examine the effect of storage conditions and time on seed viability. To test if seed longevity differs between storage conditions, we performed a GLM with a factorial design (4×1) with four storage conditions (-20, 5 and 28°C, and field storage), the control and storage time as a continuous variable; fresh seeds were considered as having zero storage time. In the model above, we used storage time as a continuous variable to show the general tendency of viability change over time according to storage conditions. To detect changes in seed viability in relation to fresh seeds and between storage periods, we fitted two other GLMs with storage time as a discrete variable, one model for ex situ storage conditions and another for in situ storage conditions (field). That is, we compared the viability of fresh seeds with stored seeds in controlled conditions (-20, 5 and 28°C) in each storage time (92, 187 and 227 days), and similarly, we compared the viability of fresh seeds with seeds stored in the field for three periods (108, 223 and 339 days). In all three GLMs, seed viability was the response variable, and storage condition and time were the predictors. We fitted the full models that included all main effects and their interactions and performed a stepwise backward selection to choose the best model according to the Akaike information criterion (AIC) value; GLMs were fitted using a binomial distribution (seeds could be viable or non-viable) and logit-link function. Additionally, we tested the viability of seeds stored at -20°C for 12 months to evaluate their potential for germplasm conservation through conventional seed banking.

Seed dormancy alleviation

We evaluated seed dormancy alleviation in the field and the effect of after-ripening temperature and time on seed dormancy alleviation by fitting GLMs that compared seed germination time and the percentage of stored seeds in the field and controlled conditions across time. Thus, we assessed the effect of storage condition and time in seed germination by performing three GLMs for each germination parameter, seed germination percentage, and mean germination time, totalling six GLMs. First, we tested if germination parameters differed between storage conditions using a factorial design (4×1) with four storage conditions (-20, 5 and28°C, as well as field), the control and storage time as a continuous variable. Then, we compared the germination parameter of fresh seeds with stored seeds in the field in each storage time (108, 223 and 339 days). Finally, we compared the germination parameter of fresh seeds with stored seeds in controlled conditions in each storage time (0, 92, 187 and 227 days). Similar to the seed longevity models, we used storage time as a continuous variable to assess the general trend of germination parameters changing across time according to storage conditions. In contrast, we used storage time as a discrete variable to compare changes in germination between storage periods in each storage condition. The fitted GLMs used the mean germination time or percentage as a response variable and storage temperature and time as predictors. For the mean germination time GLMs, we used a Gamma distribution with an inverse link function, while, for seed germination percentage, we used a binomial distribution with a logit link function. We fitted the full models that included all main effects

and their interactions and performed a stepwise backward selection to choose the best model according to the AIC value. All analyses were carried out in the R software (R Core Development Team, 2024).

Results

Seed storage behaviour and seed longevity

Fresh seeds of *C. cangae* showed a moderate moisture status (45.5 eRH%) and high viability (79.5 \pm 0.12%), which was not reduced (GLM, z = 0.432, P > 0.6) after seed desiccation to low moisture (14.8 eRH%). Seed viability decreased with storage time, but the rates of decline varied depending on the storage condition (Fig. 3A, Supplementary Table S2). The rate of viability loss was higher for seeds stored in the field than for those stored at 28 and -20° C, but it did not differ from seeds stored at 5°C (Fig. 3A). The probability that seeds remain viable is less than

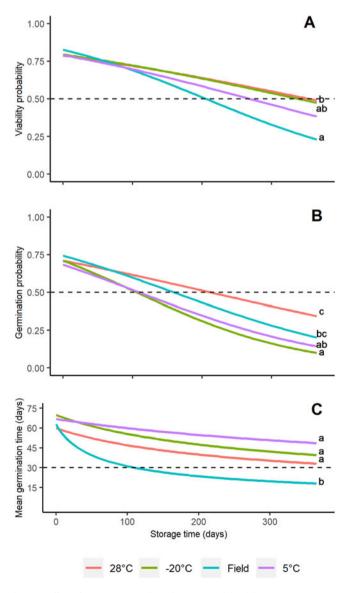


Figure 3. Effect of storage time and conditions on viability (A), germination proportion (B) and germination time (C) of *C. cangae* seeds stored in controlled conditions and in the field. Different letters indicate statistical differences. Storage time was used as a continuous variable.

50% after 213 days stored in the field, 264 days stored at 5°C, 333 days at -20°C and 350 stored at 28°C (Fig. 3A). Seed viability seems to stabilize after 277 days at -20°C, as there was no difference in seed viability between seeds stored for 277 and 365 days (Table 1).

Seed dormancy alleviation

Seed germination depended on the interaction between storage time and conditions (Supplementary Tables S3 and S4). Germination percentage decreased with time, potentially due to a decline in seed viability. However, seeds stored at -20 and 5° C lost germinability faster than those stored at 28°C. There was no difference in germination percentage between seeds stored at 28°C and seeds stored in the field (Fig. 3B). Although seed germination tended to decrease with storage time, it significantly increased in relation to fresh seeds when stored in the field or at -20°C for 108 and 187 days, respectively (Tables 1 and 2). Moreover, seeds stored in the field and at 28°C for 108 and 187 days, respectively, germinated significantly faster (lower mean germination time) than fresh seeds (Tables 1 and 2; Supplementary Figure S2). Indeed, seeds stored in the field showed faster germination than all other storage conditions (Fig. 3C).

Discussion

Our results showed that C. cangae displays an orthodox seed storage behaviour as seeds tolerate desiccation and storage at sub-zero temperatures. Seed longevity depends on storage conditions and time, where seeds stored in the field retain a viability of at least 50% for about 7 months, while seeds stored in controlled conditions (28, 5 and -20°C) retain a viability of at least 50% for about 9 months. The marked loss of seed viability in less than 1 year, regardless of storage condition, indicates a transient soil seed bank and low potential for germplasm conservation through conventional seed banking. Seed dormancy was fully alleviated after 3 months of field storage during the dry season. Moreover, seeds stored for 6 months at high temperatures (28°C) had their dormancy partially alleviated as seed germination remained high and mean germination time was deeply reduced, while low storage temperatures did not affect seed dormancy, indicating that environmental conditions found throughout the dry season in the C. cangae habitat are required to alleviate seed dormancy in this species.

Orthodox seed storage behaviour is expected in C. cangae as their seeds are released during the harsh conditions of the dry season, i.e., high temperatures and soil desiccation. Therefore, seed desiccation tolerance allows seed survival until the next rainy season. Moreover, orthodox storage behaviour is strongly correlated with herbaceous life forms, small seeds, and mountain-top grassland and shrubland ecosystems (Wyse and Dickie, 2017). Despite the orthodox storage behaviour, C. cangae seeds exhibited short longevity in ex situ storage. This short potential longevity might be related to the presence of endosperm in mature C. cangae seeds (D. Escobar, personal observation), as endospermic seeds persist for shorter periods in conventional seed banks compared to those without endosperm (Probert et al., 2009; Merritt et al., 2014; Tausch et al., 2019). Potential longevity is also associated with climate, increasing from cool and moist climates to warm and seasonally dry climates (Probert et al., 2009; Mondoni et al., 2011; Merritt et al., 2014). However, the expected relationship between potential high seed longevity and climate was not

Table 1. Effect of storage time and temperature on the viability and germination of C. canque seeds stored in controlled conditions

Storage time (days)	Storage condition	Viability proportion	Germination proportion	Mean germination time (days)
0	Fresh seeds	0.80 ± 0.12 b	0.56 ± 0.19 c	62.47 ± 10. 15 abc
92	-20°C	0.73 ± 0.08 b	0.47 ± 0.08 c	65.08 ± 8.7 ab
	5°C	0.73 ± 0.15 b	0.58 ± 0.19 c	67.88 ± 6.91 a
	28°C	0.78 ± 0.08 b	0.73 ± 0.13 cd	45.79 ± 5.11 bcde
187	-20°C	0.89 ± 0.07 b	0.81 ± 0.07 d	48.62 ± 5.58 abcd
	5°C	0.81 ± 0.09 b	0.63 ± 0.15 cd	52.36 ± 7.42 abcd
	28°C	0.79 ± 0.09 b	0.74 ± 0.11 cd	32.73 ± 5.63 e
277	-20°C	0.43 ± 0.16 a	0.03 ± 0.04 a	NA
	5°C	0.34 ± 0.15 a	0.04 ± 0.05 a	51.17 ± 19.90 cde
	28°C	0.46 ± 0.18 a	0.25 ± 0.08 b	42.77 ± 5.71 de
365	-20°C	0.45 ± 0.07 a	0 ± 0	NA

Note: Storage time as a discrete variable.

Different letters indicate statistical differences (P < 0.05) in viability and germination parameters between storage conditions and time. Values within a column followed by different bold letters indicate statistically significant differences (P < 0.05) in viability and germination parameters between stored and fresh seeds.

supported by our results, as our studied species, which is from strongly seasonally dry environments, showed short potential longevity. The short potential longevity of *C. cangae* seed may be a consequence of their evolutionary history, as endosperm presence seems conserved at the family level (Tausch et al., 2019, but see Felix et al., 2021). Nevertheless, studies accessing both embryoseed ratio evolution and potential seed longevity in the tropics are scarce and, thus, are needed to better understand seed trait correlations and plant conservation methods.

Considering that C. cangae disperses its seed from April to June and the seed persistence in the soil was approximately 7 months, the seed could retain the ability to germinate until the onset of the next rainy season (November-January). Thus, C. cangae seeds do not persist long enough to form a persistent soil seed bank but rather form a transient bank in which viable seeds persist in the soil for less than 1 year (Saatkamp et al., 2014). Our results support that soil seed bank strategies are strongly driven by climate predictability; even in a small-seeded herbaceous species, the transient soil seed bank has been selected. Moreover, C. cangae seed dormancy was fully and partially alleviated by field storage during the dry season and by ex-situ dry storage at 28°C, respectively, indicating that hot and dry climatic conditions are required to alleviate seed dormancy. Besides, seed dormancy alleviation by short duration of dry storage suggests that *C. cangae* seeds show non-deep physiological dormancy (Baskin and Baskin, 2014). Thus, dormancy enhances seed persistence in the soil by delaying germination until the onset of the next rainy season, ultimately synchronizing germination with the favourable season for seedling establishment (Donohue, 2005b).

Synchronizing seed germination with the onset of the rainy season maximizes seed fitness, given early germination increases the duration of the growing period (before the onset of the dry season), ultimately resulting in higher seedling survival (Verdù and Traveset, 2005). However, even in C. cangae seeds that were released from dormancy after dry storage in the field, germination did not occur rapidly (germination started after 10 days and mean germination was attained after 20 days; Supplementary Figure S2), suggesting that slow germination is likely due to high-water requirements for germination. Slow germination is seen as an adaptation to reduce the risk of drought-induced seedling mortality in seasonal environments, such as the occurrence of unpredictable dry spells during the onset of the rainy season (Daws et al., 2002, 2008; Donohue, 2005a; Verdù and Traveset, 2005; Kos and Poschlod, 2008; Salazar et al., 2012). Further studies examining the germination response of C. cangae seeds to hydric potential are needed to determine the role of water requirements for germination as a risk-reduction strategy in the species.

The strategies that plants use to avoid reproductive failure and ensure population persistence are the outcome of adaptive interactions among risk-reduction traits and the environmental conditions where species occur (Buoro and Carlson, 2014; Escobar et al., 2021; Chen et al., 2024). The predictability of climatic conditions in *canga* vegetation can favour protection against temporal habitat variability through transient soil seed banks, with seed

Table 2. Effect of storage time on seed viability and germination of C. cangae seeds stored in the field

Storage time (days)	Storage condition	Viability proportion	Germination proportion	Mean germination time (days)
0	Fresh seeds	0.80 ± 0.12 a	0.56 ± 0.19 b	62.47 ± 10.15 a
108	Field	0.85 ± 0.08 a	0.82 ± 0.08 c	19.55 ± 1.14 b
223	Field	0.39 ± 0.17 b	0.30 ± 0.20 a	21.36 ± 18.34 b
339	Field	0.21 ± 0.17 b	0.13 ± 0.13 a	27.33 ± 12.34 ab

Different letters indicate statistical differences (P < 0.05) in viability germination parameters between storage conditions and time. Values within a column followed by different bold letters indicate statistically significant differences (P < 0.05) in viability and germination parameters between stored and fresh seeds.

dormancy controlling the time of year in which seeds may respond to germination stimuli and slow germination time and maybe high-water requirements – avoiding seedling mortality due to dry-spells (Fenner and Thompson, 2005; Yang et al., 2021). Moreover, perenniality and iteroparity are also risk reduction traits that C. cangae exhibits to deal with the highly spatially heterogeneous environment where it occurs, allowing it to occupy safe sites via annual seed recruitment and adult survival (Saatkamp et al., 2014). Iteroparity and plant lifespan are negatively correlated with seed persistence in the soil, as high reproduction frequency buffers better against risks of local extinction caused by reproductive failure than persistent soil seed banks (Metcalf et al., 2003; Chen et al., 2024). Therefore, predictability in climatic conditions, long lifespan and iteroparity could favour transient soil seed banks in C. cangae, regardless of the low seed mass, herbaceous life form, harsh environmental conditions and high spatial variability of the canga vegetation on a micro-scale that increases the likelihood of seeds being dispersed to non-safe sites (Jacobi et al., 2007; Dayrell et al., 2021).

However, given C. cangae populations recover via freshly dispersed seeds rather than persistent soil seed banks and many individuals (around 80%) of C. cangae exhibit a short lifespan (annual behaviour, T. Zupo, personal communication, October 2024), population persistence will rely on the annual seed production and subsequent seedling establishment. Therefore, this species may suffer severe impacts due to population reduction, as it will not have a seed bank to re-establish the population, and the even short-term persistence of C. cangae depends on individuals in the area being able to grow and fruit. Hence, in situ management actions must be oriented to avoid habitat loss or degradation across the species occurrence area. Conventional seed banks do not seem to be suitable for long-term ex situ conservation as seed viability decreased from 79.5 ± 11.6 to $44.9 \pm 7.1\%$ after 1 year at -20°C and 15 RH%, but long-term seed storage studies and/or controlled aging tests are needed to understand potential longevity dynamics. We suggest examining cryopreservation as an alternative to conventional seed banking due to C. cangae desiccation-tolerant embryo (Center for Plant Conservation, 2019). We also showed that seed dormancy was fully alleviated by the environmental conditions during the dry season and partially alleviated after-ripening storage at a high constant temperature. Nonetheless, further studies testing gradients of alternating after-ripening temperatures can indicate the most effective conditions for ex situ seed dormancy alleviation.

Conclusions

C. cangae displays an orthodox seed storage behaviour; however, seed viability promptly decreases in ex situ storage, indicating the low potential for germplasm conservation through conventional seed banking. C. cangae copes with temporal habitat variability by exhibiting seed dormancy and slow germination, which synchronizes seed germination with favourable conditions for seedling establishment. Moreover, perenniality and iteroparity life-history strategies may buffer against risks of local extinction caused by reproductive failure, selecting transient rather than persistent soil seed banks. Therefore, our results suggest that in predictable climates, multiple reproduction events along the plant's lifespan (iteroparity) can favour a transient seed bank strategy even in herbaceous, small-seeded species with narrow ecological niches occurring in a highly spatially variable environment. This study also provides important insights to optimize the ex situ conservation of C. cangae.

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References

- Amorim E, Bicalho M (2022). Carajasia cangae (Rubiaceae). Rio de Janeiro -RJ, Brazil: Lista Vermelha da Flora Brasileira: Centro Nacional de Conservação da Flora/ Instituto de Pesquisas Jardim Botânico do Rio de Janeiro.
- Baskin CC and Baskin JM (2014) Seeds: Ecology, Biogeography and Evolution of Dormancy and Germination, 2nd Edn. San Diego: Academic Press.
- Boanares D, Cardoso AF, Escobar DFE, Costa KJA, Bitencourt JA, Costa PHO, Ramos S, Gastauer M and Caldeira CF (2024) The impact of rhizospheric and endophytic bacteria on the germination of *Carajasia cangae*: a threatened Rubiaceae of the Amazon Cangas. *Microorganisms* 12, 1843. https://doi.org/10.3390/microorganisms12091843
- BRASIL (2009) Ministério da Agricultura e Reforma Agrária. Secretaria Nacional de Defesa Agropecuária. Regras para análise de sementes. Brasília: Mapa/ACS, 399 p.
- **Buoro M and Carlson SM** (2014) Life-history syndromes: integrating dispersal through space and time. *Ecology Letters* 17, 756–767.
- Center for Plant Conservation (2019) CPC Best Plant Conservation Practices to Support Species Survival in the Wild. Escondido, CA: Center for Plant Conservation.
- Chen S-C, Hu X-W, Baskin CC and Baskin JM (2024) A long-term experiment reveals no trade-off between seed persistence and seedling emergence. New Phytologist 241, 623–631. https://doi.org/10.1111/nph.19270
- Costa LEN, Pinho FC, Silva KR, Carvalho CS, Tavares V, Assis R and Watanabe MTC (2023) Estudos de distribuição e abundância de espécies endêmicas de Carajás: Daphnopsis filipedunculata, Parapiqueria cavalcantei e Carajasia cangae. Relatório Técnico Interno. Belém: ITV.
- Daws MI, Burslem DFRP, Crabtree LM, Kirkman P, Mullins CE and Dalling JW (2002) Differences in seed germination responses may promote coexistence of four sympatric Piper species. Functional Ecology 16, 258–267.
- Daws MI, Garwood NC and Pritchard HW (2005) Traits of recalcitrant seeds in a semi-deciduous tropical forest in Panama: some ecological implications. Functional Ecology 19, 874–885.
- Daws MI, Crabtree LM, Dalling JW, Mullins CE and Burslem DF (2008) Germination responses to water potential in neotropical pioneers suggest large-seeded species take more risks. Annals of Botany 102, 945–951.
- Dayrell RLC, Fuzessy LF, Azevedo LO, Lambers H, Veneklaas EJ and Silveira FAO (2021) How does spatial micro-environmental heterogeneity influence seedling recruitment in ironstone outcrops? *Journal of Vegetation Science* 32, e13010. https://doi.org/10.1111/jvs.13010
- Donohue K (2005a) Niche construction through phenological plasticity: life history dynamics and ecological consequences. New Phytologist 166, 83–92.
- **Donohue K** (2005b) Seeds and seasons: interpreting germination timing in the field. Seed Science Research 15, 175–187. https://doi.org/10.1079/SSR2005208
- Escobar DFE, Rubio de Casas R and Morellato LPC (2021) Many roads to success: different combinations of life-history traits provide accurate germination timing in seasonally dry environments. *Oikos* 130, 1865–1879. https://doi.org/10.1111/oik.08522
- Felix FC, de Oliveira FN, de Freitas NWN, de Freitas RMO, Jilani IBH and Torres SB (2021) Seed characterization and germination morphophysiology of *Simira gardneriana* (Rubiaceae). *Journal of Seed Science* 43, e202143025. https://doi.org/10.1590/2317-1545v43250771
- Fenner M and Thompson K (2005) The Ecology of Seeds. Cambridge: Cambridge University Press.
- Gastauer M, Mitre SK, Carvalho CS, Trevelin LC, Sarmento PSM, Neto JAAM, Caldeira CF, Ramos SJ and Rodolfo J (2021) Landscape heterogeneity and habitat amount drive plant diversity in Amazonian canga

ecosystems. Landscape Ecology **36**, 393–406. https://doi.org/10.1007/s10980-020-01151-0

- Gioria M, Pyšek P, Baskin CC and Carta A (2020) Phylogenetic relatedness mediates persistence and density of soil seed banks. *Journal of Ecology* 108, 2121–2131. https://doi.org/10.1111/1365-2745.13437
- Giulietti AM, Giannini TC, Mota NFO, Watanabe MTC, Viana PL, Pastore M, Silva UCS, Siqueira MF, Pirani JR, Lima HC, Pereira JBS, Brito RM, Harley RM, Siqueira JO and Zappi DC (2019) Edaphic endemism in the Amazon: vascular plants of the canga of Carajás, Brazil. *Botanical Review* 85, 357–383. https://doi.org/10.1007/s12229-019-09214-x
- Gutterman Y (1994) Strategies of seed dispersal and germination in plants inhabiting deserts. *Botanical Review* **60**, 373–425.
- Hong TD and Ellis RH (1996) A Protocol to Determine Seed Storage Behavior.
 Technical Bulletin no. 1. Rome, Italy: International Plant Genetic Resources Institute.
- **Hyatt LA and Evans AS** (1998) Is decreased germination fraction associated with risk of sibling competition? *Oikos* **83**, 29–35.
- Jacobi CM, do Carmo FF, Vincent RC and Stehmann JR (2007) Plant communities on ironstone outcrops: a diverse and endangered Brazilian ecosystem. *Biodiversity and Conservation* 16, 2185–2200. https://doi.org/10.1007/s10531-007-9156-8
- Kos M and Poschlod P (2008) Correlates of inter-specific variation in germination response to water stress in a semi-arid savannah. Basic and Applied Ecology 9, 645–652.
- Long RL, Gorecki MJ, Renton M, Scott JK, Colville L, Goggin DE, Commander LE, Westcott DA, Cherry H and Finch-Savage WE (2015) The ecophysiology of seed persistence: a mechanistic view of the journey to germination or demise. Biological Reviews of the Cambridge Philosophical Society 90, 31–59.
- Mašková T, Phartyal SS, Abedi M, Bartelheimer M and Poschlod P (2022) Soil moisture level and substrate type determine long-term seed lifespan in a soil seed bank. *Plant Soil* 477, 475–485. https://doi.org/10.1007/s11104-022-05449-7
- Merritt DJ, Martyn AJ, Ainsley P, Young RE, Seed LU, Thorpe M, Hay FR, Commander LE, Shackelford N, Offord CA, Dixon KW and Probert RJ (2014) A continental-scale study of seed lifespan in experimental storage examining seed, plant, and environmental traits associated with longevity. *Biodiversity and Conservation* 23, 1081–1104.
- Metcalf JC, Rose KE and Rees M (2003) Evolutionary demography of monocarpic perennials. *Trends in Ecology & Evolution* 18, 471–480.
- Miola DTB, Ramos VDV and Silveira FAO (2021) A brief history of research in campo rupestre: identifying research priorities and revisiting geographical distribution of an ancient, widespread Neotropical biome. *Biological Journal of the Linnean Society* 133, 464–480.
- Mondoni A, Probert RJ, Rossi G, Vegini E and Hay FR (2011) Seeds of alpine plants are short lived: implications for long-term conservation. *Annals of Botany* 107, 171–179.
- Mota NFO, Silva LVC, Martins FD and Viana PL (2015) Vegetação sobre Sistemas Ferruginosos da Serra dos Carajás. In Carmo FF and Kamino LHY (orgs.) (eds) Geossistemas Ferruginosos no Brasil. Belo Horizonte: Instituto Prístino, pp. 289–315.
- Probert RJ, Manger KR and Adams J (2003) Non-destructive measurement of seed moisture. In Smith RD, Dickie JB, Linington SH, Pritchard HW, and Probert RJ (eds), Seed Conservation: Turning Science into Practice. Kew, UK: Royal Botanic Gardens, pp. 367–387.
- Probert RJ, Daws MI and Hay FR (2009) Ecological correlates of ex situ seed longevity: a comparative study on 195 species. *Annals of Botany* 104, 57–69.
- R Core Development Team (2024) R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing. Available at https://www.R-project.org.
- Rubio De Casas R, Donohue K, Venable DL and Cheptou P-O (2015) Gene-flow through space and time: dispersal, dormancy and adaptation to changing environments. Evolutionary Ecology 29, 813–831.
- Saatkamp A, Poschlod P and Venable DL (2014) The functional role of soil seed banks in natural communities. In Gallagher RS (ed.) Seeds: the

- Ecology of Regeneration in Plant Communities. Wallingford, UK: CABI, pp. 236–294.
- Salas RM, Viana PL, Cabral EL, Dessein S and Janssens S (2015) Carajasia (Rubiaceae), a new and endangered genus from Carajás mountain range, Pará, Brazil. *Phytotaxa* 206, 14. https://doi.org/10.11646/phytotaxa.206.1.4
- Salazar A, Goldstein G, Franco AC and Miralles-Wilhelm F (2012) Differential seedling establishment of woody plants along a tree density gradient in neotropical savannas. *Journal of Ecology* 100, 1411–1421.
- Santos F, Cavalcante A, Cardoso A, Caldeira Júnior C, Carvalho Neto C, Escobar D, Silveira F, Tyski L, Zanetti M and Morais R (eds.) (2023) Guia de coleta de sementes e protocolos de germinação: espécies de interesse para conservação das cangas de Carajás. Nova Lima, Brazil: Bioma Meio Ambiente, Vale.
- Satterthwaite WH (2009) Competition for space can drive the evolution of dormancy in a temporally invariant environment. *Plant Ecology* 208, 167–185.
- Schaefer CE, Corrêa GR, Candido HG, Arruda DM, Nunes JA, Araujo RW, Rodrigues PMS, Filho EIF, Pereira AFS, Brandão PC and Neri AV (2016)

 The physical environment of rupestrian grasslands (*Campos rupestres*) in Brazil: geological, geomorphological and pedological characteristics, and interplays. In Fernandes GW (ed.) *Ecology and Conservation of Mountaintop Grasslands in Brazil*. Heidelberg: Springer.
- Silveira FAO, Negreiros D, Barbosa NPU, Buisson E, Carmo FF, Carstensen DW, Conceição AA, Cornelissen TG, Echternacht L, Fernandes GW, Garcia QS, Guerra TJ, Jacobi CM, Lemos-Filho JP, Stradic SL, Morellato LPC, Neves FS, Oliveira RS, Schaefer CE, Viana PL and Lambers H (2016) Ecology and evolution of plant diversity in the endangered campo rupestre: a neglected conservation priority. *Plant and Soil* 403, 129–152.
- Souza-Filho PWM, Giannini TC, Jaffé R, Giulietti AM, Santos DC, Nascimento WR, Guimarães JTF, Costa MF, Imperatriz- Fonseca VL and Siqueira JO (2019) Mapping and quantification of ferruginous outcrop savannas in the Brazilian Amazon: a challenge for biodiversity conservation. *PLoS ONE* 14, e0211095. doi:10.1371/journal.pone.0211095
- Tausch S, Leipold M, Reisch C and Poschlod P (2019) Dormancy and endosperm presence influence the ex situ conservation potential in central European calcareous grassland plants. AoB PLANTS 11, plz035. https://doi.org/10.1093/aobpla/plz035
- **Thompson K and Grime JP** (1979) Seasonal variation in the seed banks of herbaceous species in ten contrasting habitats. *Journal of Ecology* **67**, 893–921.
- Tweddle JC, Dickie JB, Baskin CC and Baskin JM (2003) Ecological aspects of seed desiccation sensitivity. *Journal of Ecology* **91**, 294–304.
- Verdù M and Traveset A (2005) Early emergence enhances plant fitness: a phylogenetically controlled meta-analysis. *Ecology* 86, 1385–1394.
- Viana PL, Mota NFDO, Gil ADSB, Salino A, Zappi DC, Harley RM, Ilkiu-Borges AL, Secco RDS, Almeida TE, Watanabe MTC, dos Santos JUM, Trovó M, Maurity C and Giulietti AM (2016) Flora of the cangas of the Serra dos Carajás, Pará, Brazil: history, study area and methodology. Rodriguésia 67, 1107–1124.
- Wyse SV and Dickie JB (2017) Predicting the global incidence of seed desiccation sensitivity. *Journal of Ecology* 105, 1082–1093.
- Wyse SV and Dickie JB (2018) Taxonomic affinity, habitat and seed mass strongly predict seed desiccation response: a boosted regression trees analysis based on 17539 species. *Annals of Botany* 121, 71–83. https://doi.org/10.1093/aob/mcx128
- Yang X, Baskin CC, Baskin JM, Pakeman RJ, Huang Z, Gao R and Cornelissen JHC (2021) Global patterns of potential future plant diversity hidden in soil seed banks. *Nature Communications* 12, 7023. https://doi.org/10.1038/s41467-021-27379-1
- Zanetti M, Dayrell RLC, Wardil MV, Damasceno A, Fernandes T, Castilho A, Santos FMG and Silveira FAO (2020) Seed functional traits provide support for ecological restoration and ex situ conservation in the threatened Amazon ironstone outcrop flora. Frontiers in Plant Science 11, 599496. https://doi.org/10.3389/fpls.2020.599496