

A review of welfare indicators for sea turtles undergoing rehabilitation, with emphasis on environmental enrichment

R Diggins*, R Burrie, E Ariel, J Ridley, J Olsen, S Schultz, A Pettett-Willmet, G Hemming and J Lloyd

College of Public Health, Medical and Veterinary Sciences, James Cook University, 1 Solander Drive, Douglas, QLD 4811, Australia

* Contact for correspondence: Rebeccalouise.diggins@my.jcu.edu.au

Abstract

For animals undergoing rehabilitation it is vital to monitor welfare in a way that is feasible, practical, and limits stress to the animal. The industry gold standard is to assess welfare under the Five Domains model, including nutrition, environment, physical health, and behaviour as the first four physical domains and mental domain as the fifth. Feasibility and effectiveness of these domains for assessing welfare of sea turtles undergoing rehabilitation were reviewed and it was determined that the mental state can be best assessed through behavioural changes. A scoping review of the literature was conducted using Scopus and Web of Science to investigate use of environmental enrichment devices (EEDs) as a measure of welfare in sea turtles. Behavioural assessments using EEDs were found to be well-documented; however, most EED studies pertained largely to livestock or zoo animals. Furthermore, studies rarely concentrated on reptiles, and specifically sea turtles. Results also showed that certain welfare assessment methods may be less appropriate for short-term captivity experienced during rehabilitation. Additionally, the hospital environment limits the ability to address some of the domains (ie biosecurity, feasibility, safety of turtle, etc, might be compromised). This review shows that only three of the nine environmental enrichment strategies described in the literature suit the specific requirements of sea turtles in rehabilitation: feeding, tactile, and structural. It is documented that turtles display behaviours that would benefit from EEDs and, therefore, more specific studies are needed to ensure the best welfare outcomes for sea turtles undergoing rehabilitation.

Keywords: animal welfare, behaviour, captivity, enrichment devices, marine turtle, testudine

Introduction

Welfare for animals under human care is an evolving concept and one that is implemented by individual organisations (Flint *et al* 2017), resulting in varied welfare outcomes for the animals. Accredited institutions of the World Association of Zoos and Aquaria (WAZA) or the Zoo and Aquarium Association (ZAA) Australasia, for example, are bound by regulated welfare standards. For animals undergoing rehabilitation, however, welfare standards are set by specific national or state legislation, which is not always so clear or well-regulated (Englefield *et al* 2019) and often aimed at terrestrial animals and too general to be of direct relevance to sea turtles.

There are multiple ways to consider welfare. Dawkins (2008) proposed that animal welfare be determined and defined by two questions: (i) are the animals healthy? and (ii) do the animals have what they want? Ideally, the desire is for animals to experience 'good' welfare. Identifiable in the Five Freedoms of animal welfare (Farm Animal Welfare Council [FAWC] 1993), and recognised by Barnett and Hemsworth (2009), are three primary facets of welfare: basic health and functioning, psychological or affective states, and natural

living. The current industry standard for welfare assessment is the Five Domains model (Mellor 2017), which assesses animals holistically based on four physical domains (nutrition, environment, physical health, behaviour) and a fifth, mental domain. Originally this model was developed as an assessment of welfare compromise for animals held in research, teaching and testing environments (Mellor & Reid 1994). Subsequently, it has been updated to include additional categories of animals under human care, such as domestic, livestock and zoo, and to incorporate and emphasise positive states of welfare (Mellor & Beausoleil 2015).

There is no single, fully inclusive method in the determination of welfare specifically for sea turtles; however, a species-specific welfare assessment based on the Five Domains model could benefit them. A similar assessment was developed by Clegg (2015) for captive cetaceans. A species-specific assessment metric for sea turtles would have to consider individual requirements of species due to the variation between the seven species in diet and behaviours observed naturally in the wild. Whitham and Wielebnowski (2009) developed a three-step process for the maintenance of welfare for the individual animal. These involve: (i) the development of a welfare score-

sheet (based on extensive knowledge of normal parameters for the particular species); (ii) the validation of the score-sheet through a six-month behavioural and physiological assessment; finally resulting in (iii) a welfare score-sheet personalised to each species.

Such an assessment tool would be useful in a rehabilitation setting for sea turtles to ensure positive welfare, therefore promoting speedy recovery.

The rehabilitation setting is a specific environment that would require the assessment to have different considerations than if it were for sea turtles housed in zoos or aquaria without intention of release to the wild. Common causes of hospitalisation for sea turtles include boat strike, ingestion or entanglement in fishing gear or marine debris, limb damage or loss, fibropapillomatosis or other disease, and floating syndrome (Flint *et al* 2017). Each cause of hospitalisation requires consideration when housing and treating the turtles during rehabilitation. The average time spent by sea turtles in rehabilitation centres has decreased over the last couple of decades but can range from one day to more than a year, with the average time to release after rehabilitation being approximately four months (Flint *et al* 2017). Furthermore, since the aim of a rehabilitated turtle is to release it back into the wild, it is important to limit turtle-human interactions, which might be more common in an aquarium setting. Therefore, for an assessment of turtles undergoing rehabilitation, it is most important to determine the desirable state a turtle must reach before it can be released and how quickly this can be measured (Deem & Harris 2017).

Following cyclone Yasi in January 2011, in Australia's Far North Queensland, the region experienced a significant increase in sick, injured, and stranded sea turtles (Meager & Limpus 2012). Several turtle rehabilitation centres opened in response to this increase, and the College of Public Health, Medical and Veterinary Sciences, James Cook University (JCU) was transiently part of this response. Close observation of these wild animals spurred research into environmental enrichment (EE) for sea turtles in rehabilitation (Lloyd *et al* 2012), many of which have to spend months in plain plastic tanks whilst undergoing treatment. Newberry (1995) defined EE as an "improvement in the biological functioning of captive animals resulting from modifications to their environment." Hoy *et al* (2010) later organised enrichment strategies under eight classifications: feeding, tactile, structural, auditory, olfactory (ie exposing the animal to the smell of its prey), visual, social, and human-animal interaction. Maple and Perdue (2013) suggested that 'cognitive' also be included in this list. Ideally, one environmental enrichment device (EED) will be able to satisfy multiple different enrichment styles.

With an anticipated increase in hospitalised turtles following future cyclones and anthropogenically induced environmental damage, a thorough review to assess measures of welfare is critical, particularly in regard to how EE can increase speed of recovery and optimise chance of survival upon release back into the wild. This review covers suitable welfare assessment methods and how they can be adapted for turtles in rehabilitation, examples of past EE studies, and

a discussion on the design of appropriate EEDs for sea turtles in rehabilitation. Detailed explanations of auditory and olfactory EEDs are not included in this review, as there is little information on the uses of these in sea turtles.

Materials and methods

A scoping review was conducted to explore the literature pertaining to use of EEDs in turtles as a measure of welfare. Two databases were used for the search: Scopus and Web of Science. Ovid Medline was tested but yielded no relevant results so was excluded. Search terms were (environment*) AND (enrich* OR welfare OR entertain*) AND turtle* OR cheloni* OR testudine* OR reptile* OR loggerhead* OR leatherback* OR hawksbill* OR Ridley OR terrapin*) AND (rehab* OR hospital* OR clinic* OR recover* OR captiv* OR recuperat*). Searches included the full date range of each database (Scopus: 1970–present); Web of Science: 1965–present) for articles related to environmental enrichment and welfare of non-pet testudines. The reference lists of the most relevant papers were used to look for additional papers that had been missed in the database search.

From the literature search, excluding duplicates, 87 articles were identified. Titles and abstracts were reviewed against the selection criteria, which narrowed the results to 15 articles. Any literature not directly pertaining to turtles interacting with environmental enrichment was excluded. All types of environmental enrichment were included and both marine and freshwater turtle studies were included; however, tortoises were excluded. Assessment of full texts reduced the total to 11 articles (see Figure 1), of which only one was specifically relating to environmental enrichment for rehabilitation of hospitalised sea turtles. Due to the lack of specific literature, this paper reviews wider literature in the context of the Five Domains as they relate to sea turtles.

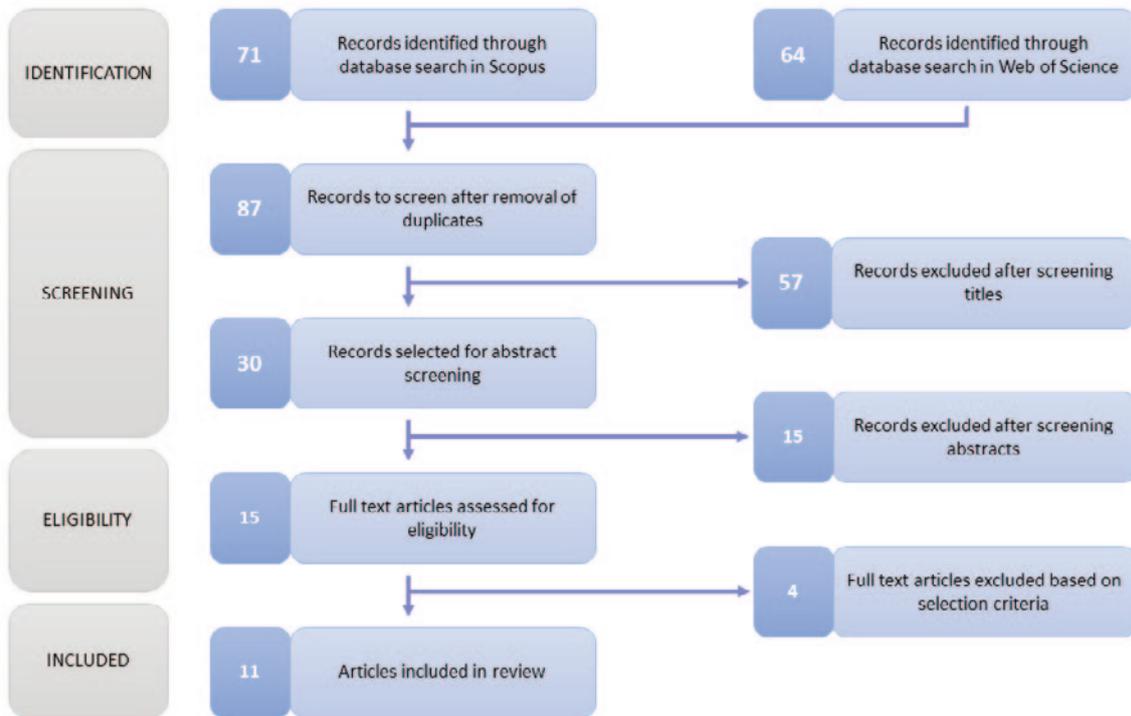
Assessing sea turtle welfare in a rehabilitation setting

Physical health evaluation

Assessing physical health in sea turtles is met with many challenges, mostly due to the absence of reliable physical and biochemical reference values (March *et al* 2018). However, there are several general parameters that are relevant across all animal species and these can be considered in a modified version for sea turtles undergoing rehabilitation.

Presence of disease and injury in a captive setting are normally considered indicators of poor welfare (Barber & Mellen 2013); however, in the rehabilitation setting, this assessment of welfare may be less useful as turtles enter the establishment already diseased/injured. Therefore, it is more logical to assess recovery rate and absence of husbandry mutilations. These can be routinely evaluated by sea turtle carers and veterinarians in rehabilitation centres based on visual inspection, behaviour and activity levels. An unpublished example of a green turtle physical exam score card (Figure 2) is provided from an Australian

Figure 1



PRISMA flow diagram of scoping review search. Papers were excluded if they did not directly discuss enrichment of freshwater or sea turtles. Papers were included even if they were not in the context of rehabilitation. Only one paper directly discussed implications of environmental enrichment of turtles in a rehabilitation setting. Review papers were excluded.

Figure 2

| Animal ID: | | | | | Location | | | | | | | |
|--------------------------|---------------------------|---|--|---|---|------|------|------|------|------|------|------|
| Comment: | | | | | Date | Date | Date | Date | Date | Date | Date | Date |
| Demeanour | Bright, alert, responsive | 0 | Quiet, alert, responsive | 1 | Non-responsive | 2 | | | | | | |
| Swim ability | Strong upright | 0 | Weak upright | 1 | Strong/Weak circling | 2 | | | | | | |
| Skin Appearance | Healthy | 0 | Minor lesions | 1 | Generalised sloughing | 2 | | | | | | |
| Skin Epiobiotic load | X<10% | 0 | 10<x<50% | 1 | 50<x<100% | 2 | | | | | | |
| Fibropapillomatosis | Nil | 0 | Less than 5 lesions | 1 | Greater than 5 lesions | 2 | | | | | | |
| Carapace Epiobiotic load | X<10% | 0 | 10<x<50% | 1 | 50<x<100% | 2 | | | | | | |
| Carapace integrity | Firm | 0 | Soft at margins | 1 | Generalised weakness | 2 | | | | | | |
| Plastron | Convex | 0 | 0 <Concave<3 cm | 1 | 3 cm <Concave | 2 | | | | | | |
| Plastron integrity | Clean | 0 | Moderate damage | 1 | Marked damage | 2 | | | | | | |
| Muscle tone | Strong | 0 | Poor | 1 | Absent | 2 | | | | | | |
| Buoyancy | Neutral | 0 | Abnormal buoyancy with ability to dive | 1 | Abnormal buoyancy without the ability to dive | 2 | | | | | | |
| Neurological exam | Jaw tone present | 0 | Jaw tone reduced | 1 | Jaw tone absent | 2 | | | | | | |
| | Palpebral present | 0 | Palpebral reduced | 1 | Palpebral absent | 2 | | | | | | |
| | Menace present | 0 | Menace reduced | 1 | Menace absent | 2 | | | | | | |
| Total | | | | | | | | | | | | |

Green turtle (*Chelonia mydas*) physical exam score card. Developed in consultation with participants in a workshop at the Turtle Health and Rehabilitation Symposium 2017, Townsville, Australia, facilitated by Dr Duane March and implemented at Dolphin Marine Magic, Coffs Harbour, Australia.

rehabilitation centre (courtesy of Dr Duane March). The level of epibionts and external parasites on admission can be visually assessed and easily treated with a freshwater bath on entry. Internal parasite infections are assumed and treated as a standard rule; however, these parasites may be resistant to treatment and therefore cause ongoing problems during rehabilitation.

Reproductive fitness may not be a reliable indicator of good welfare as captive animals have been known to reproduce well despite poor environments, and the opposite is also true (Wickins-Drazilova 2006). Specifically, for sea turtles undergoing rehabilitation, it is a poor indication of welfare as it would not be feasible to replicate the environmental conditions appropriate for successful reproduction in sea turtles. Furthermore, many of the individuals undergoing rehabilitation are sexually immature.

Stress has been linked to negative welfare (Broom & Johnson 1993) and therefore assessment of stress could be an indicator of welfare in sea turtles undergoing rehabilitation. Activation of the hypothalamic-pituitary-adrenal axis, and the subsequent release of glucocorticoids are commonly used to determine levels of stress (Stabenau & Vietti 2013; Hunt *et al* 2016). Glucocorticoid measurements may provide an indication of acute or chronic stress, depending on the chosen method of collection (blood, saliva and faecal/urine for acute stress, and samples of integumentary structures for chronic stress); however, there are numerous issues to this evaluation technique (Jessop *et al* 2004). Primarily, stress associated with reptile-capture and blood and saliva collection can interfere with results (Silvestre 2014). Additionally, glucocorticoids may be released in response to arousal, and not aversive stimuli (Latham 2010). Furthermore, there are incongruences as to the correlation of glucocorticoid levels to stress levels in sea turtle literature (Gregory *et al* 1996; Jessop *et al* 2002a,b). Finally, there seems to be a delay in green turtles' (*Chelonia mydas*) adrenocortical responses to stress (Jessop 2001). There may also be potential for adrenal fatigue in animals that are chronically debilitated (March *et al* 2018). Ironically, many of these parameters are obtained via invasive collection techniques, which may cause undue stress and actually decrease the welfare of the animal (Mason & Veasey 2010).

A number of blood parameters normally used to assess health in mammals were found to be of limited prognostic value for green turtles undergoing rehabilitation in Australia (March *et al* 2018). Although some of the parameters would provide a general indication of health, such as heterophil count and haematocrit level, none were correlated to recovery. This could be because of the particular suite of diseases encountered locally. The heterophil to lymphocyte ratio and blood glucose levels have been used to assess stress response (Davis *et al* 2008; Krams *et al* 2012), but it is clear that more research is needed to provide reliable prognostic biomarkers for each species of marine turtle in rehabilitation.

With all of these inconsistencies in mind, as well as the expense, specialised skillset, and human-turtle contact

required, measurement of glucocorticoid levels and other blood parameters are not ideal adjunctive methods of health assessment for determining welfare status of sea turtles. Of course, they are necessary for determining the health and rehabilitation status of the turtles.

Nutritional evaluation

Sea turtles entering rehabilitation centres are frequently emaciated and therefore weight gain is a priority. Some literature has shown that adult green turtles appear to do very well on high protein diets in captivity (Wood & Wood 1981; Amorcho & Reina 2008). High weight gain is achievable on such diets, which can be either animal matter (Caldwell 1962) or commercially prepared high protein, readily digestible pellets (Wood & Wood 1981). However, it is important to consider the optimal diet for sea turtles undergoing rehabilitation. There is a natural variation in the diets of wild sea turtles of different species and life stages (Limpus & Limpus 2000; Arthur *et al* 2009). Therefore, diet needs to be tailored to the specific nutritional requirements of the individual to reflect their natural preferences. A number of rehabilitation centres have been known to feed turtles a high protein diet to encourage rapid weight gain, irrespective of species (E Ariel, personal communication 2021). For a predominantly herbivorous species, such as the green turtle, this does not reflect a natural diet and may lead to uraemia and hypercholesterolaemia (March *et al* 2018).

Weight gain by itself is not necessarily an indicator of welfare; however, it can be used in conjunction with body condition scoring (Limpus *et al* 2012) to show progress for rehabilitation of emaciated sea turtles. Body condition reflects not only the availability of appropriate and nutritious food items in the captive setting, but also appetite and physiological ability to convert food to build muscle and support activity. This method can be applied for sea turtles, where the body condition index (BCI) is recorded regularly, and release is dependent on having achieved a BCI consistent with wild populations (Bjorndal 1980). A more accurate method of scoring body condition would be bio-impedance analysis as that would differentiate between weight gain caused by fluid, fat or muscle (Kophamel submitted). However, this requires specialised equipment and training, as well as additional human-turtle interactions. Melvin *et al* (2021) have also found evidence that malnutrition is a key factor in mortality of sea turtles undergoing rehabilitation and suggest monitoring metabolomic profiles for earlier diagnosis and treatment of metabolic failure.

Whilst poor body condition/weight loss is often precipitated by stress, it is also influenced by diet, activity levels (Mason & Mendl 1993), and disease. Cachexia is a common finding in sea turtles presenting to rehabilitation clinics (March *et al* 2021). Ideally, in a rehabilitation setting, each turtle's diet would be formulated to cater for maintenance, whilst taking activity levels and disease status into consideration. Overall, measuring weight in conjunction with body scoring is a useful method to assess welfare. It is minimally invasive and can be obtained on a weekly basis by rehabilitation staff and carers.

Environmental evaluation

The environmental domain for a captive turtle can be evaluated in two stages: (i) the initial set-up of the tank; and (ii) the ongoing maintenance of tank conditions. Considerations when designing an enclosure for sea turtles should include substrate, structure/shape, size, depth, material and colour (Stamper *et al* 2017; New South Wales Government 2020). Substrate, structure and material for a sea turtle tank should consider that turtles are likely to ingest anything small enough (Hoopes *et al* 2017). Particularly in a rehabilitation setting, it would be disadvantageous to put turtles in an environment where they may do more harm to themselves through ingestion or scraping against rough surfaces. Juvenile green turtles showed a preference toward the colour blue under experimental settings; therefore, implementation of blue tanks may improve their comfort (Hall *et al* 2018). Tanks should be deep enough to provide refuge, but weak turtles are at risk of drowning, and so fitness of the turtle needs to be considered (Stamper *et al* 2017). These features of the environment are likely to remain constant throughout the entire rehabilitation period and so anticipated length of time in captivity (as well as species) should be considered at set-up. This is particularly relevant to enclosure size as turtles must have sufficient space to manoeuvre and engage in positive natural behaviours (Stamper *et al* 2017).

Environmental conditions that can be regularly and simply monitored to ensure comfort for sea turtles include temperature, light, UV, salinity and other water quality parameters (Stamper *et al* 2017). Sea turtles have a range of tolerability for each of these parameters; if they are not well-monitored and maintained, it is possible that sea turtles already in a weakened state, such as those undergoing rehabilitation, might become further compromised by sub-optimal environmental conditions. For example, as ectotherms, reduced temperatures will reduce the efficiency of the digestive and immune system, which would be detrimental for underweight sick turtles (Hoopes *et al* 2017). These are all environmental conditions that are always essential to the physical well-being of sea turtles; however, variety in non-essential environmental stimuli has been shown to positively affect welfare of other animals (Burghardt 2013) and should, therefore, be considered for use with sea turtles. EEDs can be introduced to do this and the change in behaviour of the turtles can be used to assess the impact on welfare.

Behavioural evaluation

It has commonly been perceived that stereotypic behaviour is indicative of either past and/or present poor welfare (Mason 1991; Mason & Latham 2004; Garner 2005; Mason *et al* 2007). Indeed, the presence or absence of stereotypic behaviour remains one of the best validated measures of animal welfare (Maple & Perdue 2013). Mason *et al* (2007) proposed that stereotypic behaviour, as a broad term, should refer to “repetitive behaviour induced by frustration, repeated attempts to cope and/or central nervous system (brain)

dysfunction.” In the rehabilitation setting, changes in behaviour could be due to brain damage caused by parasites such as spirorchiid flukes (Glazebrook *et al* 1989) or coccidia (Gordon *et al* 1993) or, alternatively, it could be environmentally induced as a result of boredom or reduced welfare. This is particularly likely if the turtles are kept in sterile, empty hospital tanks, devoid of environmental enrichment.

Abnormal behaviours indicating stress in turtles include grafting of jaw (rasping of ramphotheca), pseudo-vocalisation (squeaks or whines), pattern swimming, poor posture when resting at the bottom of the tank (flopped and lifeless rather than propped up on front flippers), and boundary exploration (related to exploratory and escape activity) (Tynes 2010; Arena *et al* 2014). Leatherback turtles (*Dermochelys coriacea*) are particularly difficult to keep in captivity due to their inability to register boundaries. They are continuous swimmers and can cause additional damage to themselves if allowed to swim into the sides of a rehabilitation tank (Jones *et al* 2000; Levy *et al* 2005). Turtles recently hospitalised, or handled in and out of the water, may display behavioural floating for a period. This could be as a response to stress or a preference to be at the surface due to weakened physical condition (Manire *et al* 2017). Buoyancy disorder due to gas accumulation within the coelomic cavity will be discussed later. Associated with the presence of or contact with humans, other stress-related behaviours include cloacal evacuations upon handling, projection of penis or hemi-pene, voluntary regurgitation of food, and human-directed aggression. Often these signs are related to fear and are common in overly restrictive and inappropriate environments (Warwick *et al* 2013).

Stereotypic behaviour tends to be associated with negative welfare in healthy animals (ie in zoos/aquaria), but in the case of sick turtles, it can actually illustrate improved health via increased energy levels. However, if they are to be kept longer for full rehabilitation, stereotypic behaviours should be discouraged. EEDs are a useful tool, commonly used in captive settings to discourage stereotypic behaviours and encourage positive behaviours (Mason *et al* 2007). Consequentially, observing animals for the presence or absence of negative behaviours could be used as a proficient welfare evaluation measure, and potentially as a means of determining the effectiveness of EEDs, particularly in turtles that have spent several months in rehabilitation. Additionally, comparing captive animal behaviour with wild animal behaviour (Burghardt *et al* 1996; Smith & Litchfield 2010; Phillips *et al* 2011) is another measure of welfare. The more a captive-held animal engages in behaviour exhibited in the wild, the better its welfare is deemed. Similarly, the effectiveness of EE can be deduced by comparing the proportion of time an animal is engaged in a type of behaviour before and after introduction of an EED (Therrien *et al* 2007; Lloyd *et al* 2012).

Mental evaluation

The physical domains (health, nutrition, environment and behaviour) all contribute to the mental state of the turtles (Mellor 2017). The affective state of an animal can be assessed via study of its behaviour (Bracke & Hopster 2006). Stress fever and tachycardia, both physiological responses associated with emotion in other vertebrates, have been observed in iguanas (*Iguana iguana*) (Cabanac 1999) and wood turtles (*Clemmys insculpta*) (Cabanac & Biernieri 2000). Cabanac (1999) also discovered that rather than venture into a cold environment to obtain food, iguanas preferred to remain in a warm environment, suggesting that their motivation was influenced by sensory pleasure. Therefore, it appears that basic affective states exist in reptiles, turtles included. In the assessment of affective states, there is a potential issue of over-anthropomorphism and evaluator bias.

Using EEDs to monitor welfare

Modification of the environment to provide more opportunities and promote positive behaviours can be used to infer the affective state of the turtles and assess their welfare. EEDs should be designed to increase positive affective state of turtles but must also be suitable for the rehabilitation setting. EEDs are all designed to enhance environmental opportunity and choice, but depending on the specific device, could also promote positive behavioural expression, increase fitness and aid nutrition. Thus, contributing to a positive affective state for the turtles and improved welfare. It is on this premise that EEDs may be able to contribute to a speedier recovery and shorter rehabilitation time of hospitalised turtles.

The psychological and physical benefits of EEDs are well documented in captive mammals (Newberry 1995; Mellen & MacPhee 2001; Young 2013), but less so in the case of marine and terrestrial reptiles (de Azevedo *et al* 2007; Maple & Perdue 2013; Eagan 2019). Reptiles have previously been considered too sedentary to interact with, and thus benefit from, EE (Bennett 1982; Burghardt 2013). Turtles housed at JCU proved this to be a misconception by actively interacting with EEDs (Lloyd *et al* 2012). Furthermore, a literature review by Lambert *et al* (2019) found multiple studies that showed sentience in reptiles, including multiple turtle species. We therefore found it timely to conduct a thorough review of past reptile-specific EED studies as well as to draw from existing knowledge of wild sea turtle ecology to explore the potential for EEDs in assisting with rehabilitation of hospitalised turtles.

EEDs for turtles undergoing rehabilitation

At this point, it is necessary to make a distinction between EE for hospitalised turtles and those that are permanently captive (such as in public aquaria). For all captive turtles, it is desirable for their captive conditions to be as similar to their wild conditions as practically possible (Newberry 1995). Hospital settings, however, are often not conducive to this as they must remain sterile to reduce likelihood of infection, for example. As such,

EEDs should aim to stimulate natural behaviours safely without jeopardising the necessary sanitation standards of a hospital setting or the safety of the turtle. Therefore, EEDs should encourage ‘preferred’ naturalistic living. The term ‘preferred’ is used to omit negative aspects of naturalistic living, such as famine and predation (Hutchins 2006). Predatory avoidance behaviours correlated with stress could reduce longevity of animals in long-term captivity, which would be associated with negative welfare. However, anti-predator responses are necessary for temporarily captive turtles to ensure a good chance of survival on release. Turtles intended for release after rehabilitation, therefore, need to maintain a level of fearfulness, which could be promoted through subjection to occasional and temporary unpleasant stimuli (Guy *et al* 2013). With respect to this, it is difficult to prepare sea turtles for natural life in an artificial environment, especially in a rehabilitation setting where emphasis is on improving health and fitness. An ideal welfare evaluation plan for sea turtles in the rehabilitation setting would adhere to the following considerations:

- Be **safe** for the turtle;
- Be **feasible** in the rehabilitation setting;
- Be **cost-effective**;
- Be **easily implemented** by carers without the requirement for specialised skills or training;
- Be **minimally invasive to induce little or no stress** on the turtles, which is especially important as these turtles are diseased and/or injured and added stress is likely to exacerbate immunosuppression, subsequently lengthening recovery time;
- Accurately measure **stress in conjunction with behavioural assessment**;
- Require **minimal human-turtle contact**; and
- Require a **short-term** evaluation of welfare variables to provide a reliable indication of welfare.

Feeding enrichment

Turtles in the wild appear to feed in bouts — early to mid-morning and mid to late afternoon (Ogden *et al* 1983) — and therefore reproducing this pattern in the captive setting to maintain the natural rhythm may be beneficial for release. Food-oriented devices appear to be a very effective form of EE (Maple & Perdue 2013). As a reflection of their natural foraging behaviour, hunting of live jellyfish, ctenophores, and squid would be a valuable EED for turtles in captivity or those undergoing rehabilitation. However, the ethical dilemma associated with live feeding, biosecurity, and the availability of such prey may exclude this EED. The lettuce feeders on the tank floor reported by Therrien *et al* (2007) may prove an interesting activity for turtles as this mimics grazing behaviour (Van de Merve *et al* 2009; Hart & Fujisaki 2010) and serves a dual purpose, as a hiding place.

Injuries and ailments of each individual turtle need to be considered when designing the EED. ‘Floating syndrome’,

which affects the turtle's buoyancy, can be caused by air trapped in the lungs, coelomic cavity, or intestine of the turtle. The air upsets diving proficiency, which prevents the turtle from reaching the tank floor, resulting in major feeding constraints (Norton 2005). However, occasional bottom feeding for floating turtles would encourage them to try to dive down when they have enough energy. A possible alternative could consist of a frozen ice-block, containing squid and vegetable matter, such as cos lettuce and nori, to encourage foraging and provide the turtles with a focused interactive activity for an extended period of time. Entanglement is another common cause of turtle hospitalisation. Entanglement may result in amputation of a flipper, causing restricted movement, which also needs consideration when designing EEDs. In general, natural foraging on the tank floor should be encouraged as well as a disassociation between humans and food.

Tactile enrichment

Hoy *et al* (2010) described tactile EE as “the provision of objects that are physically stimulating to the animal.” To reflect their natural environment, turtles may benefit from the inclusion of muddy or sandy floor bottoms, perhaps contained within a tray to maintain ease of cleaning and water drainage; however, this is unlikely to be feasible in a sterile rehabilitation setting. Employment of stones too large to ingest, however, could provide excellent enrichment, for green turtles in particular, as they are attracted to rocky rubble to perform self-cleaning behaviours (Heithaus *et al* 2002; E Ariel & J Lloyd, personal observation 2012). Whilst captive turtles have been observed to swim under brooms in order to groom themselves (Brill *et al* 1995; Lloyd *et al* 2012), turtles have also been known to eat the broom bristles. Consequentially, this EED comes with risks and, if utilised, should only be provided under supervision. Provision of a ‘waterfall’, as well as toys such as hoops and balls, would provide valuable tactile enrichment (Burghardt 2005).

Structural enrichment

In promoting naturalistic living, turtles should have access to shallow water for resting (Brill *et al* 1995). This can be achieved in the form of a platform suspended from the wall of the tank or positioned in the centre of the tank. Alternatively, water levels could be lowered for floating turtles, to enable them to reach the tank floor and right themselves with their flippers. Turtles should also have deeper parts in their tanks, ideally with 3D structures that could mimic caves (Brill *et al* 1995). A pipe on the tank floor, large enough for hiding their head, allows turtles to hide and/or exclude external stimuli during resting periods (Therrien *et al* 2007; Lloyd *et al* 2012). Hatchlings and young post-hatchlings can be buoyant and so EEDs on the tank floor may not be appropriate. Therefore, mounting pipes to the side of the tank or in shallow water for young or floating turtles would provide a suitable refuge.

Social and visual enrichment

Sea turtles in restricted environments should be housed individually due to their typically solitary tendencies (Heithaus *et al* 2002) and documented aggression in overcrowded facilities (Arena *et al* 2014) and during mating (Schofield *et al* 2007). However, cohabitation with other species, such as a green turtle and brown tang (*Acanthurus nigrofuscus*) or yellow tang (*Zebrasoma flavescens*) (Balazs *et al* 1994) could potentially act as a form of social EE. Inter-species cohabitation would also provide visual enrichment (something to look at), whilst additionally satisfying the natural behaviour of the green turtle to be clean. However, Zamzow (1998) showed that whilst this cohabitation may be beneficial for control of ectoparasites, reef fish may serve as vectors in the spread of fibropapillomatosis or create an opportunity for infection if the turtle is wounded during cleaning. This would also require additional husbandry for the fish, which would be costly to the rehabilitation facility in terms of time and money.

Cognitive and human-animal enrichment

Maple and Perdue (2013; p 108) described cognitive enrichment as: “challenging and stimulating an organism's memory, decision-making, judgment, perception, attention, problem-solving, executive functioning, learning and species-specific abilities.” A training routine using associative learning (Lopez *et al* 2001; Wilkinson *et al* 2007, 2009) would provide this type of enrichment and has been proven possible in marine turtles (Mellgren & Mann 1998; Bartol *et al* 2003). However, since rehabilitation turtles only remain in facilities temporarily, training may not be a worthwhile form of EE due to the potential time investment required for it to be successful. Additionally, although human-turtle interactions may be encouraged in aquaria to increase familiarity and reduce stress (Claxton 2011), they should be limited in temporary captive settings. Turtles may have extensive long-term memory (Bartol *et al* 2003; Davis 2009; Davis & Burghardt 2012); therefore, human-turtle interactions could cause potential overdependence and ‘trust’ towards humans. Lack of caution towards humans would be disadvantageous to the turtles after release as it could lead to injury (Addison & Nelson 2000).

Past examples of EE in captive turtles

A case study from a Spanish rehabilitation centre, based on the work of Therrien *et al* (2007), showed that EE aided in the successful rehabilitation and release of a sea turtle that was previously considered unfit for release due to a flipper amputation (Monreal-Pawlowsky *et al* 2017). Recognising the limitations of implementing EE in a rehabilitation environment, enrichment was based on feeding, tactile and structural stimuli. Enrichment primarily involved eating live food and aimed to prepare the turtle to avoid unnatural objects in the water, such as buoys. Despite being in captivity for ten years, including a two-year rehabilitation

period, two-months of EE was sufficient to prepare the turtle for release into the wild. This successful release was confirmed by ten-month transmission from a satellite tag that showed the loggerhead turtle (*Caretta caretta*) crossed an expansive body of water. It is unknown how quickly a turtle might be released with a timelier introduction to EE as no specific studies for this were found in the literature. However, it is important to note that EE in this case study was administered over a short time-period, easy to implement, cost-effective and required minimal human interaction as a webcam was used for monitoring.

Research was undertaken on the effects of EE on four captive display sea turtles (three loggerhead turtles and one blind green turtle) in Florida (Therrien *et al* 2007). The behaviour of the turtles was assessed both with and without enrichment present. The EEDs were designed to stimulate their tactile sense, increase exploratory swimming, and satisfy their need to forage. The study showed that there was a significant increase in amount of time engaged in naturalistic behaviours with the use of EEDs. The devices for the blind turtle were modified to suit its special needs and successfully decreased the stereotypical behaviour and increased the exploratory behaviour of the animal. In an enrichment study of captive-raised, collectively housed green turtles intended for release, Kanghae *et al* (2021) found that enrichment devices decreased negative behaviour. Specifically, the turtles exposed to enrichment had fewer bite wounds than turtles without enrichment and without other health parameters affected. EE appears to be just as effective for marine reptiles as it is for mammalian species, and should be encouraged for captive sea turtles, including disabled ones, and particularly when housed collectively.

A preliminary study on hospitalised sea turtles, conducted by Lloyd *et al* (2012) arrived at similar conclusions. Lloyd *et al* (2012) demonstrated that there was an overall decrease in pattern swimming and resting behaviours observed amongst the turtles in the presence of EE. Additionally, it was found that each turtle responded to different EEDs in their own specific ways, highlighting the apparent variances in natural behaviours and preferences between individuals. It is also important to consider the possibility that turtles will habituate to an EED if given unrestricted access to it. Consequentially, EEDs should be rotated and their use potentially supervised (Lloyd *et al* 2012). Furthermore, the placement of structural elements of the captive environment should be altered two to three times a year to maintain their novelty factor (Hawkings & Willemsen 2004).

Relatively few studies on EE in sea turtles are published. For this reason, we have included studies on freshwater turtles. Case *et al* (2005) assessed the preference as well as the physiological and behavioural effects of enriched versus barren environments on 38 box turtles (*Terrapene carolina*). Preference for the habitat-enriched environment was apparent. Following the preference tests, turtles were housed for a one-month period in one of the two environments. Behaviourally, turtles with habitat enrichment spent less time engaged in negative behaviours, and physiologically they had

significantly lower heterophil to lymphocyte (H/L) ratios than turtles in the barren environment. This illustrates that turtles prefer EE, that enrichment improves their welfare, and importantly, that this improvement can be observed in their behaviour. Similarly, Tetzlaff *et al* (2018, 2019a,b) found that even captive-born *T. carolina* intrinsically preferred enriched habitats, and that enriched environments, along with time for growth in captivity, might aid survival post-release.

Food-centred enrichment for freshwater turtles has also been studied. Bryant and Kother (2015) used puzzle-based feeding enrichment devices to successfully increase time spent feeding and promote foraging behaviour of Fly River turtles (*Carettochelys insculpta*) on display at ZSL London Zoo, UK. Bannister *et al* (2021) introduced scented and unscented enrichment devices pre-feeding to reduce negative behaviour in a group of freshwater (*Pseudemys* sp and *Trachemys scripta* ssp) display turtles at Tynemouth Aquarium, UK. Presence of enrichment devices pre-feeding successfully reduced escape behaviour and turtles showed greater interest in scented devices than unscented, indicating that olfactory enrichment is appropriate for captive turtles.

Burghardt (2005) observed ‘play’ behaviour in a captive Nile soft-shell turtle (*Trionyx triunguis*) that was introduced to five EEDs: two basketballs of different colours, a hoop, a rubber fill hose, and live fish for feeding. Burghardt (2005; p 82) defined play as “repeated, incompletely functional behaviour differing from more serious versions structurally, contextually, or ontogenetically, and initiated voluntarily when the animal is in a relaxed or low stress setting.” These EEDs were introduced in an effort to reduce boredom-induced self-mutilation (Burghardt *et al* 1996). It was observed that this soft-shelled turtle played with the EEDs for 21% of observed time. This play is longer than juvenile captive mammals, including primates, which play between 1 and 10% of the time (Fagen 1981). Burghardt (2005) also mentioned object play behaviour in another two Nile soft-shelled turtles at Toronto Zoo, as relayed by reptile curator Robert Johnson. Indeed, there are other examples of play in turtles, including object play in a loggerhead turtle (Burghardt 2005), locomotor play in a wood turtle and social play in Emydidae turtles (Burghardt 2005). Therefore, EEDs designed to encourage play should be considered for hospitalised turtles in order to increase welfare and reduce rehabilitation time.

Animal welfare implications

Maintaining positive welfare of animals under human care is of utmost importance. When considering appropriate methods to assess welfare status and promote positive welfare some distinctions need to be made specifically for sea turtles undergoing rehabilitation. Species- and lifestage-specific considerations need to be made but also limitations due to the hospital environment should be considered. The Five Domains model of welfare can be applied to assess welfare of sea turtles, and reviewed for appropriateness, effectiveness and feasibility for application in the rehabilitation setting. Physical health evaluation methods are highly specialised, invasive and expensive and not easily imple-

mented by rehabilitation staff. Nutritional evaluation should always be carefully considered with rehabilitation turtles and more research is needed to assess effects of poor diet on the physical health of sea turtles in captivity. The environmental implications on welfare of turtles undergoing rehabilitation can be difficult to manage due to the need for the environment to be sterile and easily cleaned, which makes this domain difficult to assess. The behavioural domain is easily assessed by rehabilitation staff and can be used to infer mental state of the sea turtles. For this reason, behavioural assessment of turtles and mental affective states whilst undergoing rehabilitation should be routinely undertaken to promote positive welfare.

The limited literature shows that sea turtles respond to EEDs and can benefit from enrichment to improve their welfare whilst in captivity. They have been observed to have basic affective states, engage in play behaviours, and to respond positively to the introduction of EEDs. Through the use of EEDs (including devices to encourage foraging, complex multi-dimensional environments, and hides), designed according to the requirements of the rehabilitation centre and the needs of the individual turtle, it is possible to cover the three main facets of welfare, and thereby assist in the recovery and preparation of rehabilitated turtles for release back into the wild. The authors hope that this literature review will contribute to the recognition of the advantages and significance of EE in hospitalised sea turtles, and to encourage turtle rehabilitators to effectuate and employ EEDs. Future research projects may also assess the impact of various EEDs to determine the most beneficial of these on the welfare of hospitalised and other captive sea turtles, through welfare measures such as a reduction in stereotypic behaviour and faster recovery times, the ultimate goal being to improve the welfare of sea turtles held in confinement.

Declaration of interest

The authors wish to declare that they have no conflict of interest, or relationship, financial or otherwise, that might be perceived as influencing objectivity.

References

- Addison DS and Nelson KA** 2000 Recapture of a tagged, captive reared juvenile loggerhead turtle: an example of habituation? *Marine Turtle Newsletter* 89: 15-16
- Amorochio D and Reina R** 2008 Intake passage time, digesta composition and digestibility in East Pacific green turtles (*Chelonia mydas agassizii*) at Gorgona National Park, Colombian Pacific. *Journal of Experimental Marine Biology and Ecology* 360: 117-124. <https://doi.org/10.1016/j.jembe.2008.04.009>
- Arena P, Warwick C and Steedman C** 2014 Welfare and environmental implications of farmed sea turtles. *Journal of Agricultural and Environmental Ethics* 27: 309-330. <https://doi.org/10.1007/s10806-013-9465-8>
- Arthur K, McMahon K, Limpus C and Dennison W** 2009 Feeding ecology of green turtles (*Chelonia mydas*) from Shoalwater Bay, Australia. *Marine Turtle Newsletter* 123: 6-12
- Balazs G, Losey G and Privitera L** 1994 Cleaning symbiosis between the wrasse, *Thalassoma duperoy*, and the green turtle, *Chelonia mydas*. *Copeia* 3: 684-690. <https://doi.org/10.2307/1447184>
- Bannister CC, Thomson AJC and Cuculescu-Santana M** 2021 Can colored object enrichment reduce the escape behavior of captive freshwater turtles? *Zoo Biology* 40(2): 160-168. <https://doi.org/10.1002/zoo.21583>
- Barber JC and Mellen JD** 2013 Animal ethics and welfare. In: Irwin MD, Stoner JB and Cobaugh AM (eds) *Zookeeping: An Introduction to the Science and Technology* pp 53-61. University of Chicago Press: Chicago, USA
- Barnett J and Hemsworth P** 2009 Welfare monitoring schemes: using research to safeguard welfare of animals on the farm. *Journal of Applied Animal Welfare Science* 12: 114-131. <https://doi.org/10.1080/10888700902719856>
- Bartol S, Mellgren R and Musick J** 2003 Visual acuity of juvenile loggerhead sea turtles (*Caretta caretta*): a behavioral approach. *International Journal of Comparative Psychology* 16: 143-155
- Bennett A** 1982 The energetics of reptilian activity. *Biology Of The Reptilia* 13: 155-199
- Bjorndal KA** 1980 Nutrition and grazing behavior of the green turtle *Chelonia mydas*. *Marine Biology* 56(2): 147-154. <https://doi.org/10.1007/BF00397131>
- Bracke M and Hopster H** 2006 Assessing the importance of natural behavior for animal welfare. *Journal of Agricultural and Environmental Ethics* 19: 77-89. <https://doi.org/10.1007/s10806-005-4493-7>
- Brill R, Balazs G, Holland K, Chang R, Sullivan S and George J** 1995 Daily movements, habitat use, and submergence intervals of normal and tumor-bearing juvenile green turtles (*Chelonia mydas* L) within a foraging area in the Hawaiian islands. *Journal of Experimental Marine Biology and Ecology* 185: 203-218. [https://doi.org/10.1016/0022-0981\(94\)00146-5](https://doi.org/10.1016/0022-0981(94)00146-5)
- Broom D and Johnson KJ** 1993 Stress and strain, welfare and suffering. In: Broom D (ed) *Stress and Animal Welfare* pp 80-82. Kluwer Academic Publishers: The Netherlands. <https://doi.org/10.1007/978-94-024-0980-2>
- Bryant Z and Kother G** 2015 Environmental enrichment with simple puzzle feeders increases feeding time in fly river turtles (*Carettochelys insculpta*). *Herpetological Bulletin* 130: 3-5
- Burghardt G** 2005 *Genesis of Animal Play: Testing the Limits*. MT Press: Cambridge, UK. <https://doi.org/10.7551/mitpress/3229.001.0001>
- Burghardt G** 2013 Environmental enrichment and cognitive complexity in reptiles and amphibians: concepts, review, and implications for captive populations. *Applied Animal Behaviour Science* 147(3-4): 286-298. <https://doi.org/10.1016/j.applanim.2013.04.013>
- Burghardt G, Rosscoe R and Ward B** 1996 Problem of reptile play: environmental enrichment and play behavior in a captive Nile soft-shelled turtle, *Trionyx triunguis*. *Zoo Biology* 15: 223-238. [https://doi.org/10.1002/\(SICI\)1098-2361\(1996\)15:3<223::AID-ZOO3>3.0.CO;2-D](https://doi.org/10.1002/(SICI)1098-2361(1996)15:3<223::AID-ZOO3>3.0.CO;2-D)
- Cabanac M** 1999 Emotion and phylogeny. *Journal of Consciousness Studies* 6: 176-190
- Cabanac M and Bernieri C** 2000 Behavioral rise in body temperature and tachycardia by handling of a turtle (*Clemmys insculpta*). *Behavioral Processes* 49: 61-68. [https://doi.org/10.1016/S0376-6357\(00\)00067-X](https://doi.org/10.1016/S0376-6357(00)00067-X)

- Caldwell D** 1962 Growth measurements of young captive Atlantic sea turtles in temperate waters. *Contributions in Science* 50: 1-8. <https://doi.org/10.5962/p.241045>
- Case B, Lewbart G and Doerr P** 2005 The physiological and behavioural impacts of and preference for an enriched environment in the eastern box turtles (*Terrapene carolina carolina*). *Applied Animal Behaviour Science* 92: 353-365. <https://doi.org/10.1016/j.applanim.2004.11.011>
- Claxton AM** 2011 The potential of the human-animal relationship as an environmental enrichment for the welfare of zoo-housed animals. *Applied Animal Behaviour Science* 133(1-2): 1-10. <https://doi.org/10.1016/j.applanim.2011.03.002>
- Clegg IL, Borger-Turner JL and Eskelinen HC** 2015 C-Well: The development of a welfare assessment index for captive bottlenose dolphins (*Tursiops truncatus*). *Animal Welfare* 24(3): 267-282. <https://doi.org/10.7120/09627286.24.3.267>
- Davis K** 2009 *Sociality, cognition and social learning in turtles* (Emydidae). Dissertation, Tennessee Research and Creative Exchange, USA
- Davis K and Burghardt G** 2012 Long-term retention of visual tasks by two species of emydid turtles, *Pseudemys nelsoni* and *Trachemys scripta*. *Journal of Comparative Psychology* 126: 213-223. <https://doi.org/10.1037/a0027827>
- Davis A, Maney D and Maerz J** 2008 The use of leukocyte profiles to measure stress in vertebrates: A review for ecologists. *Functional Ecology* 22(5): 760-772. <https://doi.org/10.1111/j.1365-2435.2008.01467.x>
- Dawkins MS** 2008 The science of animal suffering. *Ethology* 114: 937-945. <https://doi.org/10.1111/j.1439-0310.2008.01557.x>
- deAzevedo CS, Cipreste CF and Young RJ** 2007 Environmental enrichment: a GAP analysis. *Applied Animal Behaviour Science* 102(3-4): 329-343. <https://doi.org/10.1016/j.applanim.2006.05.034>
- Deem SL and Harris HS** 2017 Nutrition. In: Manire CA, Norton TM and Stacy BA (eds) *Sea Turtle Health and Rehabilitation* pp 945-953. J Ross Publishing, Incorporated: FL, USA
- Eagan T** 2019 Evaluation of enrichment for reptiles in zoos. *Journal of Applied Animal Welfare Science* 22(1): 69-77. <https://doi.org/10.1080/10888705.2018.1490182>
- Englefield B, Blackman SA, Starling M and McGreevy PD** 2019 A review of Australian animal welfare legislation, regulation, codes of practice, and policy, and their influence on stakeholders caring for wildlife and the animals for whom they care. *Animals* 9(6): 335. <https://doi.org/10.3390/ani9060335>
- Fagen R** 1981 *Animal Play Behavior*. Oxford University Press: New York, USA
- Farm Animal Welfare Council** 1993 *Second report on priorities for research and development in farm animal welfare*. Farm Animal Welfare Council, MAFF: Tolworth, UK
- Flint J, Flint M, Limpus CJ and Mills P** 2017 Status of marine turtle rehabilitation in Queensland. *PeerJ* 5: e3132. <https://doi.org/10.7717/peerj.3132>
- Garner J** 2005 Part II: stereotypic behaviours as pathologies. In: Mason G and Rushen J (eds) *Stereotypic Animal Behaviour: Fundamentals and Applications to Welfare, Second Edition*. CABI Publishing: Cambridge, UK
- Glazebrook JS, Campbell RSF and Blair D** 1989 Studies on cardiovascular fluke (*Digenea: Spirorchidae*) infections in sea turtles from the Great Barrier Reef, Queensland, Australia. *Journal of Comparative Pathology* 101(3): 231-250. [https://doi.org/10.1016/0021-9975\(89\)90033-9](https://doi.org/10.1016/0021-9975(89)90033-9)
- Gordon AN, Kelly WR and Lester RJG** 1993 Epizootic mortality of free-living green turtles, *Chelonia mydas*, due to coccidiosis. *Journal of Wildlife Diseases* 29(3): 490-494. <https://doi.org/10.7589/0090-3558-29.3.490>
- Gregory L, Gross T, Bolten A, Bjorndal K and Guillette L** 1996 Plasma corticosterone concentrations associated with acute captivity stress in the loggerhead sea turtle (*Caretta caretta*). *General and Comparative Endocrinology* 104(3): 312-320. <https://doi.org/10.1006/gcen.1996.0176>
- Guy AJ, Curnoe D and Banks PB** 2013 A survey of current mammal rehabilitation and release practices. *Biodiversity and Conservation* 22(4): 825-837. <https://doi.org/10.1007/s10531-013-0452-1>
- Hall RJ, Robson SK and Ariel E** 2018 Colour vision of green turtle (*Chelonia mydas*) hatchlings: do they still prefer blue under water? *PeerJ* 6: e5572. <https://doi.org/10.7717/peerj.5572>
- Hart K and Fujisaki I** 2010 Satellite tracking reveals habitat use by juvenile green sea turtles, *Chelonia mydas*, in the Everglades, Florida, USA. *Endangered Species Research* 11: 221-232. <https://doi.org/10.3354/esr00284>
- Hawkings M and Willemsen M** 2004 Environmental enrichment for amphibians and reptiles. *ASZK Reptile Enrichment Workshop. Zoological Parks Board of NSW. Thylacinus vol 28, no 3*. <http://tester.enrichment.org/MiniWebs/Australasia/workshop01.pdf>
- Heithaus M, McLash J, Frid A, Dill L and Marshall G** 2002 Novel insights into green sea turtle behaviour using animal-borne video cameras. *Journal of the Marine Biological Association of the United Kingdom* 82: 1049-1050. <https://doi.org/10.1017/S0025315402006689>
- Hoopes LA, Koutsos EA and Norton TM** 2017 Nutrition. In: Manire CA, Norton TM and Stacy BA (eds) *Sea Turtle Health & Rehabilitation* pp 63-90. J Ross Publishing, Incorporated: FL, USA
- Hoy J, Murray P and Tribe A** 2010 Thirty years later: enrichment practices for captive mammals. *Zoo Biology* 29: 303-316. <https://doi.org/10.1002/zoo.20254>
- Hunt KE, Innis CJ, Kennedy AE, McNally KL, Davis DG, Burgess EA and Merigo C** 2016 Assessment of ground transportation stress in juvenile Kemp's Ridley sea turtles (*Lepidochelys kempii*). *Conservation Physiology* 4(1). <https://doi.org/10.1093/conphys/cov071>
- Hutchins M** 2006 Death at the zoo: the media, science, and reality. *Zoo Biology* 25: 101-115. <https://doi.org/10.1002/zoo.20085>
- Jessop T** 2001 Modulation of the adrenocortical stress response in marine turtles (*Cheloniidae*): evidence for a hormonal tactic maximizing maternal reproductive investment. *Journal of Zoology* 254: 57-65. <https://doi.org/10.1017/S0952836901000553>
- Jessop T, Knapp R, Whittier J and Limpus C** 2002a Dynamic endocrine responses to stress: evidence for energetic constraints and status dependence in breeding male green turtles. *General and Comparative Endocrinology* 129: 59-67. <https://doi.org/10.1006/gcen.2001.7769>

- Jessop T, Limpus C and Whittier J** 2002b Nocturnal activity in the green sea turtle alters daily profiles of melatonin and corticosterone. *Hormones and Behaviour* 41: 357-365. <https://doi.org/10.1006/hbeh.2002.1775>
- Jessop TS, Sumner JM, Limpus CJ and Whittier JM** 2004 Interplay between plasma hormone profiles, sex and body condition in immature hawksbill turtles (*Eretmochelys imbricata*) subjected to a capture stress protocol. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 137(1): 197-204. <https://doi.org/10.1016/j.cbpa.2003.09.029>
- Jones TT, Salmon M, Wyneken J and Johnson C** 2000 Rearing leatherback hatchlings: protocols, growth and survival. *Marine Turtle Newsletter* 90: 3-6
- Kanghae H, Thongprajukaew K, Inphrom S, Malawa S, Sandos P, Sotong P and Boonsuk K** 2021 Enrichment devices for green turtles (*Chelonia mydas*) reared in captivity programs. *Zoo Biology* 40(5): 407-416. <https://doi.org/10.1002/zoo.21613>
- Krams I, Vrublevska J, Cirule D, Kivleniece I, Krama T, Rantala MJ, Sild E and Hõrak P** 2012 Heterophil/lymphocyte ratios predict the magnitude of humoral immune response to a novel antigen in great tits (*Parus major*). *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 161(4): 422-428. <https://doi.org/10.1016/j.cbpa.2011.12.018>
- Lambert H, Carder G and D'Cruze N** 2019 Given the cold shoulder: A review of the scientific literature for evidence of reptile sentience. *Animals* 9(10): 821. <https://doi.org/10.3390/ani9100821>
- Latham N** 2010 Chapter 6. Brief introduction to welfare assessment: A 'Toolbox' of techniques. In: Hubrecht R and Kirkwood JK (eds) *The UFAW Handbook on the Care and Management of Laboratory and Other Research Animals, Eighth Edition*. Wiley: West Sussex, UK
- Levy Y, King R and Aizenberg I** 2005 Holding a live leatherback turtle in Israel: lessons learned. *Marine Turtle Newsletter* 107: 7-8
- Limpus C and Limpus D** 2000 Mangroves in the diet of *Chelonia mydas* in Queensland, Australia. *Marine Turtle Newsletter* 89: 13-15
- Limpus CJ, Limpus DJ, Savige M and Shearer D** 2012 *Health assessment of green turtles in south and central Queensland following extreme weather impacts on coastal habitat during 2011*. Department of Environment and Heritage Protection: QLD, Australia
- Lloyd J, Ariel E, Adams D and Owens L** 2012 Environmental enrichment for sea turtles in rehabilitation: preliminary study. *Proceedings of the 8th National Australian Wildlife Rehabilitation Conference*. 16-20 July 2012, Townsville, QLD, Australia
- Lopez J, Gomez Y, Rodriguez F, Broglio C, Vargas J and Salas C** 2001 Spatial learning in turtles. *Animal Cognition* 4: 49-59. <https://doi.org/10.1007/s100710100091>
- Manire CA, Norton TM, Walsh MT and Campbell LA** 2017 Buoyancy disorders. In: Manire CA, Norton TM and Stacy BA (eds) *Sea Turtle Health & Rehabilitation* pp 45-60. J Ross Publishing, Incorporated: FL, USA
- Maple T and Perdue B** 2013 *Zoo Animal Welfare*. Springer: Heidelberg, New York, Dordrecht, London. <https://doi.org/10.1007/978-3-642-35955-2>
- March DT, Marshall K, Swan G, Gerlach T, Smith H, Blyde D, Ariel E, Christidis L and Kelaher BP** 2021 The use of echocardiography as a health assessment tool in green sea turtles (*Chelonia mydas*). *Australian Veterinary Journal* 99(1-2): 46-54. <https://doi.org/10.1111/avj.13039>
- March DT, Vinette-Herrin K, Peters A, Ariel E, Blyde D, Hayward D, Christidis L and Kelaher BP** 2018 Hematologic and biochemical characteristics of stranded green sea turtles. *Journal of Veterinary Diagnostic Investigation* 30(3): 423-429. <https://doi.org/10.1177/1040638718757819>
- Mason G** 1991 Stereotypies: a critical review. *Animal Behaviour* 41: 1015-1037. [https://doi.org/10.1016/S0003-3472\(05\)80640-2](https://doi.org/10.1016/S0003-3472(05)80640-2)
- Mason G and Latham N** 2004 Can't stop, won't stop: is stereotypy a reliable animal welfare indicator? *Animal Welfare* 13: 57-69
- Mason G, Clubb R, Latham N and Vickery S** 2007 Why and how should we use environmental enrichment to tackle stereotypic behaviour? *Applied Animal Behaviour Science* 102(3-4): 163-188. <https://doi.org/10.1016/j.applanim.2006.05.041>
- Mason G and Mendl M** 1993 Why is there no simple way of measuring animal welfare? *Animal Welfare* 2: 301-319
- Mason G and Veasey JS** 2010 How should the psychological well-being of zoo elephants be objectively investigated? *Zoo Biology* 29(2): 237-255. <https://doi.org/10.1002/zoo.20256>
- Meager JJ and Limpus C** 2012 Marine wildlife stranding and mortality database annual report 2011 III. *Marine Turtle Conservation Technical and Data Report* 3: 1-46
- Mellen J and MacPhee M** 2001 Philosophy of environmental enrichment: past, present and future. *Zoo Biology* 20: 211-226. <https://doi.org/10.1002/zoo.1021>
- Mellgren R and Mann M** 1998 What can a green sea turtle learn? In: Abreu-Grobois F, Briseño-Dueñas R, Márquez R and Sarti L (eds) *Proceedings of the Eighteenth International Sea Turtle Symposium*. US Department of Commerce, NOAA Tech Memo. NMFS-SEFSC-436 pp 79-80. 3-7 March 1998, Mazatlán, Sinaloa, México. http://aquaticcommons.org/21311/1/Fish_TM_436.pdf
- Mellor DJ** 2017 Operational details of the Five Domains model and its key applications to the assessment and management of animal welfare. *Animals* 7: 60. <https://doi.org/10.3390/ani7080060>
- Mellor DJ and Beausoleil NJ** 2015 Extending the 'Five Domains' model for animal welfare assessment to incorporate positive welfare states. *Animal Welfare* 24(3): 241. <https://doi.org/10.7120/09627286.24.3.241>
- Mellor DJ and Reid CSW** 1994 Concepts of animal well-being and predicting the impact of procedures on experimental animals. *Improving the Well-being of Animals in the Research Environment* 3-18. <https://www.wellbeingintlstudiesrepository.org/cgi/viewcontent.cgi?article=1006&context=expawel>
- Melvin SD, March DT, Marshall K, Carroll AR and van de Merwe JP** 2021 Improving rehabilitation outcomes using metabolomics: Health, recovery and biomarkers of mortality in sick and injured green turtles (*Chelonia mydas*). *Biological Conservation* 254: 108943. <https://doi.org/10.1016/j.biocon.2020.108943>
- Monreal-Pawlowsky T, Marco-Cabedo V, Membrive GP, Sanjose J, Fuentes O, Jimenez E and Manteca X** 2017 Environmental enrichment facilitates release and survival of an injured loggerhead sea turtle (*Caretta caretta*) after ten years in captivity. *Journal of Zoo and Aquarium Research* 5: 182-186
- Newberry R** 1995 Environmental enrichment: increasing the biological relevance of captive environments. *Applied Animal Behaviour Science* 44: 229-243. [https://doi.org/10.1016/0168-1591\(95\)00616-Z](https://doi.org/10.1016/0168-1591(95)00616-Z)

- New South Wales Government** 2020 *Code of Practice for Injured and Sick Sea Turtles and Sea Snakes*. <https://www.environment.nsw.gov.au/research-andpublications/publications-search/code-of-practice-forinjured-and-sick-sea-turtles-and-sea-snakes>
- Norton T** 2005 Sea turtle conservation in Georgia and an overview of the Georgia Sea Turtle Center on Jekyll Island, Georgia. *Georgia Journal of Science* 63: 208-231
- Ogden J, Robinson L, Witlock K, Daganhardt H and Cebula R** 1983 Diel foraging patterns in juvenile green turtles (*Chelonia mydas*) in St Croix United States Virgin Islands. *Journal of Experimental Marine Biology and Ecology* 66: 199-205. [https://doi.org/10.1016/0022-0981\(83\)90160-0](https://doi.org/10.1016/0022-0981(83)90160-0)
- Phillips C, Jiang Z, Hatton A, Tribe A, Bouar M, Guerlin M and Murray P** 2011 Environmental enrichment for captive eastern blue-tongue lizards, *Tiliqua scincoides*. *Animal Welfare* 20: 377-384
- Schofield G, Katselidis K, Pantis J, Dimopoulos P and Hays G** 2007 Female-female aggression: structure of interaction and outcome in loggerhead sea turtles. *Marine Ecology Progress Series* 336: 267-274. <https://doi.org/10.3354/meps336267>
- Silvestre A** 2014 How to assess stress in reptiles. *Journal of Exotic Pet Medicine* 23: 240-243. <https://doi.org/10.1053/j.jepm.2014.06.004>
- Smith B and Litchfield C** 2010 An empirical case study examining effectiveness of environmental enrichment in two captive Australian sea lions (*Neophoca cinera*). *Journal of Applied Animal Welfare Science* 13:407-416. <https://doi.org/10.1080/10888700903371863>
- Stabenau EK and Vietti K** 2003 The physiological effects of multiple forced submergences in loggerhead sea turtles (*Caretta caretta*). *Fishery Bulletin* 101(4): 889-899
- Stamper AM, Harms CA and Lewbart GA** 2017 Environment/water quality/biosecurity. In: Manire CA, Norton TM and Stacy BA (eds) *Sea Turtle Health & Rehabilitation* pp 45-60. J Ross Publishing, Incorporated: FL, USA
- Tetzlaff SJ, Sperry JH and DeGregorio BA** 2018 Captive-reared juvenile box turtles innately prefer naturalistic habitat: Implications for translocation. *Applied Animal Behaviour Science* 204: 128-133. <https://doi.org/10.1016/j.applanim.2018.03.007>
- Tetzlaff SJ, Sperry JH and DeGregorio BA** 2019a Trade-offs with growth and behavior for captive box turtles head-started with environmental enrichment. *Diversity* 11: 40. <https://doi.org/10.3390/d11030040>
- Tetzlaff SJ, Sperry JH, Kingsbury BA and DeGregorio BA** 2019b Captive-rearing duration may be more important than environmental enrichment for enhancing turtle head-starting success. *Global Ecology and Conservation* 20: e00797. <https://doi.org/10.1016/j.gecco.2019.e00797>
- Therrien C, Gaster L, Cunningham-Smith P and Manire A** 2007 Experimental evaluation of environmental enrichment of sea turtles. *Zoo Biology* 26: 407-416. <https://doi.org/10.1002/zoo.20145>
- Tynes VV** 2010 *Behavior of Exotic Pets*. John Wiley & Sons: USA
- Van de Merve J, Ibrahim K, Lee S and Whittier J** 2009 Habitat use by green turtles (*Chelonia mydas*) nesting in Peninsular Malaysia: local & regional conservation implications. *Wildlife Research* 36: 367-645. <https://doi.org/10.1071/WR09099>
- Warwick C, Arena P, Lindley S, Jessop M and Steedman C** 2013 Assessing reptile welfare using behavioral criteria. *In Practice* 35: 123-131. <https://doi.org/10.1136/inp.fl197>
- Whitham J and Wielebnowski N** 2009 Animal-based welfare monitoring: using keeper ratings as an assessment tool. *Zoo Biology* 28: 545-560. <https://doi.org/10.1002/zoo.20281>
- Wickins-Drazilova D** 2006 Zoo animal welfare. *Journal of Agricultural and Environmental Ethics* 19: 27-36. <https://doi.org/10.1007/s10806-005-4380-2>
- Wilkinson A, Chan H and Hall G** 2007 Spatial learning and memory in the tortoise (*Geochelone carbonaria*). *Journal of Comparative Psychology* 121(4): 412-418. <https://doi.org/10.1037/0735-7036.121.4.412>
- Wilkinson A, Coward S and Hall G** 2009 Visual and response-based navigation in the tortoise (*Geochelone carbonaria*). *Animal Cognition* 12: 779-787. <https://doi.org/10.1007/s10071-009-0237-9>
- Wood J and Wood F** 1981 Growth and digestibility for the green turtle (*Chelonia mydas*) fed diets containing varying protein levels. *Aquaculture* 25: 269-274. [https://doi.org/10.1016/0044-8486\(81\)90188-5](https://doi.org/10.1016/0044-8486(81)90188-5)
- Young RJ** 2013 *Environmental Enrichment for Captive Animals*. John Wiley & Sons: USA
- Zamzow J** 1998 Cleaning symbioses between Hawaiian reef fishes and green sea turtles, *Chelonia mydas*. In: Abreu-Grobois FA, Briseño-Dueñas R, Márquez R and Sarti L (eds) *Proceedings of the Eighteenth International Sea Turtle Symposium*. US Dep Commerce. NOAA Tech Memo NMFS-SEFSC-436 pp 235-237. http://aquaticcommons.org/2131/1/Fish_TM_436.pdf