

Wind-borne dispersion of *Echinococcus multilocularis* eggs – a flight model

S. Siegert  and S. Neumann 

Institute of Veterinary Medicine, University of Göttingen, Göttingen D-37077, Germany

Research Paper

Cite this article: Siegert S, Neumann S (2022). Wind-borne dispersion of *Echinococcus multilocularis* eggs – a flight model. *Journal of Helminthology* **96**, e45, 1–7. <https://doi.org/10.1017/S0022149X22000360>

Received: 21 March 2022

Revised: 9 June 2022

Accepted: 10 June 2022

Key Words:

Echinococcus multilocularis; *E. multilocularis* eggs; zoonotic disease; Alveolar echinococcosis; Alveolar hydatid disease; dispersion; wind-borne dispersion; mathematical model; risk management; flight model; parameters of *E. multilocularis* eggs; rate of descent

Author for correspondence:

S. Siegert,

E-mail: stephan.siegert@uni-goettingen.de

Abstract

The alveolar hydatid disease, also known as alveolar echinococcosis, of humans is certainly one of the most dangerous zoonoses worldwide. The disease is caused by *Echinococcus multilocularis* – the fox tapeworm. Red foxes (*Vulpes vulpes*) are currently counted as the most important carriers (reservoirs) of *E. multilocularis* in the Northern Hemisphere. The possible routes of infection of *E. multilocularis* to humans are complex and still require research. Until now, it has been unknown whether *E. multilocularis* eggs can be moved by wind at all. This analysis shows, based on calculations, that *E. multilocularis* eggs can be transported by wind. Using a mathematical model, flight distances depending on wind speed and take-off heights are calculated for dense and less dense (coniferous) forest areas. The results – differentiated for seasons and as overall average – are based on mean values of wind speeds which were measured over a ten-year period in an experimental forest stand in the Solling (Germany). Due to their rate of descent, wind-related spreading of *E. multilocularis* eggs is possible. The average flight distance covered by *E. multilocularis* eggs in forest areas, depending on their starting altitude and wind speed, is between approximately 1.3 m and approximately 17 m. From the mathematical point of view, the wind factor can definitely be seen as one of the multiple vectors associated with environmental contamination by *E. multilocularis* eggs. Consequently, the possible wind-borne spread of *E. multilocularis* eggs poses an infection risk to humans that should be considered and requires further research.

Introduction

Alveolar hydatid disease, also known as alveolar echinococcosis (AE), ranks as one of the most dangerous helminthic zoonoses in the world (Ito *et al.*, 2007). Without appropriate medical measures and chemotherapeutic treatment strategies, this cancer-like disease has a death rate of more than 90% within ten years after diagnosis (Torgerson *et al.*, 2008). Also, the veterinary aspect is of great concern. Animals that are not part of the (sylvatic) transmission cycle, such as pigs, apes or domestic animals such as dogs and cats can not only be (seriously) infected with the disease, but also increase the risk of infection for humans (Deplazes & Eckert, 2001). The causative agent of AE is the metacestode of *Echinococcus multilocularis* (Deplazes & Eckert, 2001). The distribution of *E. multilocularis* covers almost the entire Northern Hemisphere. Its endemic areas reach from western Europe to Russia, parts of Asia (including Hokkaido, Japan), Alaska (including St. Lawrence), Canada and the continental United States (Torgerson *et al.*, 2010; Deplazes *et al.*, 2017).

Echinococcus multilocularis measures only 1.2 mm to 4.5 mm and contains in its proglottis from 200 up to 300 eggs (Thompson, 1986; Eckert *et al.*, 2011). Despite its small body size, *E. multilocularis* compensates with an enormous mass infestation. In individual cases, more than 100,000 worms can colonize a single host (Fischer *et al.*, 2005). This allows a release of an enormous number of eggs per host and defecation into the environment. For foxes, the definitive hosts, the maximum number of eggs excreted daily was found to be 98,638 (Kapel *et al.*, 2006). The consequential contamination of the environment can become a dangerous infection source.

Echinococcus multilocularis eggs belong to the Taeniidae. All of this family's eggs are similar in terms of their morphology (Lawson & Gemmell, 1983). They are round-elliptical in shape (Thompson, 1986). The size of *E. multilocularis* eggs varies between 28 × 39 µm and 30 × 40 µm (Veit *et al.*, 1995).

They are 'extremely resistant, enabling them to withstand a wide range of environmental temperatures' (Thompson, 1986) while sensitive to aridity and high temperatures (Lawson & Gemmell, 1983).

During an experiment in Germany exposing *E. multilocularis* eggs to natural climatic conditions, the eggs survived a maximum of 240 days during autumn and winter and 78 days in summer. Generally, it can be assumed that the eggs usually survive well over 100 days in the environment (Veit *et al.*, 1995).

© The Author(s), 2022. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.

The main infection path is hand–mouth contact after direct contiguity with affected primary hosts, namely foxes, but in some cases also dogs and other animals (see above), because the eggs get stuck in their fur (Conraths & Deplazes, 2015; Merck, 2018). Also, the handling of contaminated soil is mentioned (Romig *et al.*, 2017). Furthermore, *E. multilocularis* is third in the global ranking of food-borne parasites. Therefore, a transmission and infection by contaminated food, such as fruits, vegetables or even water should not be dismissed (Food and Agriculture Organization of the United Nations & World Health Organization, 2014). The routes of transmission and dispersion of *E. multilocularis*, respectively its eggs, are of highly complex pattern, yet, still insufficiently researched (Lawson & Gemmell, 1983; Ishikawa *et al.*, 2003; Tolnai *et al.*, 2013; Hegglin *et al.*, 2015).

There have been prior studies using mathematical models to describe transmission dynamics of *E. multilocularis* for estimating the risk of infection. These models considered influences of important mechanisms and variables regarding the transmission of the parasite (e.g. treatment of foxes, age structures, seasonality, habitat types, deworming of dogs, environmental disinfection, etc.) (Atkinson *et al.*, 2013; Lewis *et al.*, 2014; Hassan & Munganga, 2019; Khan *et al.*, 2020; Woolsey & Miller, 2021). While these models provide valuable insights into modulating infection risks, they ignore that there is a second category of transmission – the abiotic distribution. Meanwhile a well-known fact that dispersion of *E. multilocularis* eggs can be affected by various biotic factors such as herbivores, birds, numerous insects, especially flies, different beetles, butterflies, snails, etc. the abiotic distributors of eggs, rain and wind (Guo *et al.*, 2017) are longstanding objects of controversial discussion (Lawson & Gemmell, 1983). To this day, the evidence that wind could contribute to the dispersal of *E. multilocularis* eggs has been absent. A study detecting a uniform radial dispersion of *Taenia hydatigena* eggs may argue against a dispersion by wind (Gemmell

et al., 1978). In contrast, indications for the wind-borne dispersion of *Echinococcus granulosus* eggs have been shown by several studies (Bourée, 2001; Sánchez Thevenet *et al.*, 2019). This supports the hypothesis of the involvement of wind also for *E. multilocularis*. Therefore, it is of huge importance to investigate the role of wind. If wind has an influence on transmission dynamics of *E. multilocularis* eggs, this could increase the risk of infection for AE.

Material and methods

Flight model

To analyse the possible epidemiological impact of wind, we developed a mathematical flight model.

In order to develop such a model, it is necessary to understand the relationships between the physical parameters that enable the flight of an *E. multilocularis* egg. Those dependencies are shown in fig. 1. The graphic (fig. 1a) explains the relationships. The starting point (S) (shown here as a stylized tree stump) determines the take-off height (h). The wind acts on the egg with the wind speed (u) and carries it on. As soon as the egg starts its flight, gravity forces it back to earth at its specific rate of descent (v) – proportional to the wind speed (u). The distance (e), that the egg covers during this process to the landing point (Z), gives the flight distance of the egg. The flight range is described by the same angle (α) as the rate of descent in relation to the wind speed and the take-off height in relation to the distance. This flight path angle is determined as the equation in fig. 1b. Rearranging this equation to e results in the final formula for the calculation of the flight length (fig. 1c).

This simple model calculates the minimal flight distance of an *E. multilocularis* egg depending on the constant ‘rate of descent’ (v) and the variable parameters ‘wind speed’ (u) and ‘starting altitude’ (h), provided that the flight is not disturbed.

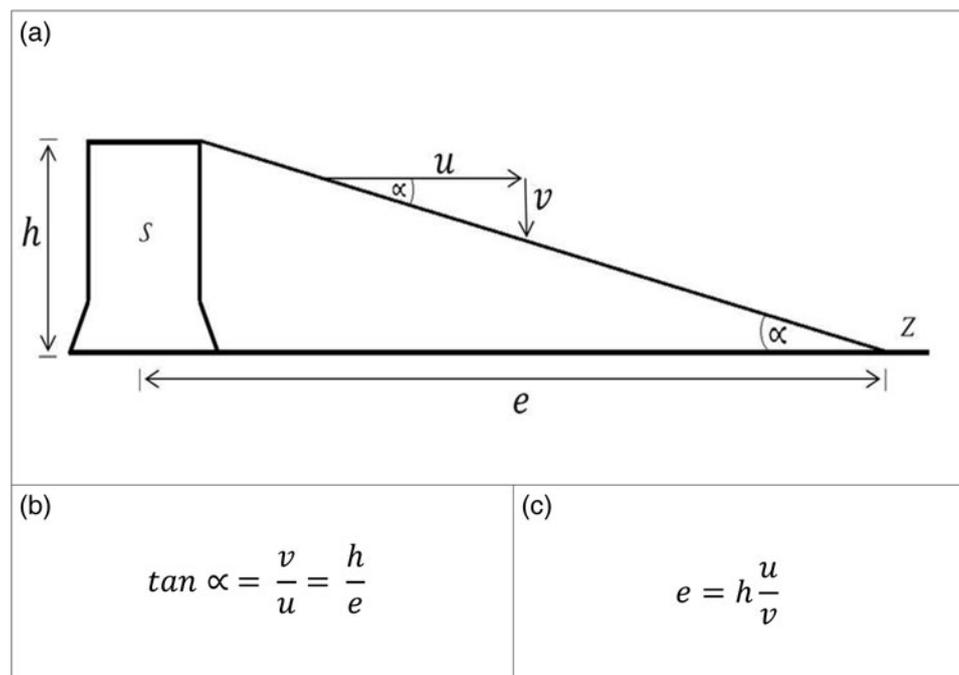


Fig. 1. Flight model describing the physical parameters and dependencies influencing the flight of an *Echinococcus multilocularis* egg (a) with mathematical description of the flight path angle (b) and the final formula for flight path calculation (c). h = take-off height, u = wind speed, v = rate of descent, e = distance, S = starting point (tree stump), Z = zero (finish).

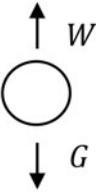
(a) 	(b) $W = 6 \cdot \pi \eta r v$	(c) $G = m \cdot g$
(d) $W = G$	(e) $6 \cdot \pi \mu v r = m \cdot g$	(f) $v = \frac{m \cdot g}{6 \cdot \pi \cdot \mu \cdot r}$

Fig. 2. Graphical illustration describing the two important forces affecting the egg (a). Formula of frictional resistance (b) and gravitation (c) equated (d and e) and rearranged for calculation of the rate of descent (v) for an *Echinococcus multilocularis* egg. (Avison, 1989; Bird *et al.*, 2007) W = frictional resistance (according to Stokes), η = dynamic viscosity of the fluid (liquid or gas) in which the body moves (here: $18.2 \cdot 10^{-6}$ Nsec./mm²), r = radius (here: $0.017125 \cdot 10^{-3}$ m), v = rate of descent, G = gravitation, m = mass (here: $0.024502 \cdot 10^{-9}$ kg), g = gravitational acceleration (here: 9.81 m/sec²).

Determination of the 'rate of descent' (v)

Two important forces are affecting the egg (fig. 2a): the frictional resistance (W) (fig. 2b) (Bird *et al.*, 2007) which has to be overcome in order to enable transport by air; and the gravitation (G) (fig. 2c) (Avison, 1989) which forces the flying object back to the ground. The flight ability of an ovum is therefore largely dependent on its rate of descent.

To determine the rate of descent, the size and the mass of an *E. multilocularis* egg are required. While the average size of these eggs is known (Veit *et al.*, 1995), we needed to determine the weight of an *E. multilocularis* egg. Figure 3 shows the mathematical methods used to approximate the volume and weight. The diameter measures between $28 \times 39 \mu\text{m}$ and $30 \times 40 \mu\text{m}$ (Veit *et al.*, 1995). In our calculations (fig. 3) we applied the mean value 0.029×0.0395 mm. According to the round but slightly elliptical shape of the egg (Thompson, 1986; Veit *et al.*, 1995), we decided to calculate three different volumes and intersect them, to incorporate the special shape of the *E. multilocularis* egg as closely as possible in our approximation. Therefore, volume values were determined for both an ellipsoid (fig. 3a) (Weisstein, 2002) and a sphere (fig. 3b) (Weisstein, 2002). The spherical volume was calculated for a 'small' sphere (related to 0.029 mm), and for a 'large' sphere (related to 0.0395 mm). Both volumes were averaged. This 'mean sphere' was intersected with the volume of the ellipsoid, resulting in an *E. multilocularis* volume of $2 \cdot 10^{-5}$ mm³.

Since the specific weight of eggs of the Taeniidae family is given as 1.2251 g/ml (David & Lindquist, 1982), the weight of a single *E. multilocularis* egg equals 0.024502 μg .

In order to determine the rate of descent of the egg, W and G are equated (figs. 2d and e) and rearranged to v (fig. 2f). Consequently, the rate of descent is $v = 4.09 \cdot 10^{-2}$ m/s, respectively, $v = 40.9$ mm/s.

Determination of wind speeds (u)

To obtain reliable wind data for applying into the model, data sets originating from a dense stocked Norway spruce (*Picea abies*) stand (test area F1) in the Solling (Germany) were accessed (Meesenburg *et al.*, 1999; Paar *et al.*, 2013).

We averaged the hourly measured wind speeds of a ten-year period (1990 to 1999) as follows: maximum and minimum values of each month of each year were read out of the data pool, while error-values were ignored. In step two these minima and maxima were allocated to the four seasons of the year. In a third step, the determination of the means per season per year followed, as well as the mean over the entire year. The fourth step was the compilation of all mean values of the individual years and their seasons in a table in order to determine the mean values over all ten years. By this step, the ten-year mean values at a height of 1 m for spring, summer, autumn, winter and as an annual total could be generated.

Determination of starting heights (h)

Based on the fox's behaviour about the placement of faeces in forest areas, there are many assumable heights for potential starting points (altitudes) of *E. multilocularis* eggs (e.g. litter, tree stumps, stones, trunks, etc.) (Goszczyński, 1990). Consequently, to the behaviour (Goszczyński, 1990) and body size (Cavallini, 1995) of foxes, we took the heights 0.3, 0.5, 0.7, 0.8, 0.9 and 1.0 (in m) into consideration. Since wind speed data of only a height of 1 m were available, we needed to extrapolate the wind speeds for these heights using the formula in fig. 4 (Gualtieri & Secci, 2011). Here, the roughness length is required. The roughness length is a fictitious height above the ground at which the average wind speed approaches zero. This (vertical) length, usually named z_0 , is used in the logarithmic law of wind and describes the

(a) $V = \frac{4}{3} \cdot \pi \cdot a \cdot b \cdot c$	(b) $V = \frac{4}{3} \cdot \pi \cdot r^3$
--	--

Fig. 3. Formula for calculating the volume of an ellipsoid (a) and a sphere (b). (Weisstein, 2002) V = volume, r = radius, a , b , c = half-axes.

$$v_2 = v_1 \frac{\ln\left(\frac{h_2}{z_0}\right)}{\ln\left(\frac{h_1}{z_0}\right)}$$

Fig. 4. Formula for the extrapolation of wind speeds (Gualtieri & Secci, 2011). v_2 = wind speed in m/s in the height h_2 , v_1 = reference wind speed in m/s measured on the reference height h_1 , z_0 = roughness length, h_2 = height above the ground, \ln = natural logarithm.

$$e = h \frac{u}{40.9 \frac{\text{mm}}{\text{s}}}$$

Fig. 5. Formula to estimate flight lengths of *Echinococcus multilocularis* eggs. h = take-off height, u = wind speed, e = distance.

roughness of the ground surface, for example, the presence of grass or bushes (American Meteorological Society, 2019). The roughness length for dense areas of the forest (e.g. stocked stands) is given 0.8 m and 0.2 m for less dense areas (e.g. windthrow areas) (Pertmann, 2011). By applying these values, we were able to extrapolate the wind speeds downwards using the formula in fig. 4 (Gualtieri & Secci, 2011) and thereby to simulate the flight of an *E. multilocularis* egg in two different environments: dense forest

areas (with z_0 : 0.8 m, starting altitudes (in m): 1.0, 0.9, 0.8); and less dense forest areas (with z_0 : 0.2 m, starting altitudes (in m): 0.7, 0.5, 0.3). A further extrapolation downwards would be superfluous, since at heights below the roughness length calm prevails.

Results

As described in the material and methods section, we developed the following mathematical model as a practical approach to estimate flight lengths of *E. multilocularis* eggs in dependence of wind speed and starting altitude summarized by the definitive formula (fig. 5).

Table 1 presents the calculated flight distances of an *E. multilocularis* egg in a (coniferous) forest stand depending on the wind speed (Wind), the take-off height (Height) above the ground and – with extrapolated values – the roughness length for the two scenarios ‘Forest area (dense)’ and ‘Forest area (less dense)’. The model analysis has revealed, that in forest areas, an egg can be transported several metres by wind and that its flight distances are influenced by season. Further it showed the more the roughness length is lowered, the further the transport length rises.

Flight distances – overall

As can be seen in table 1, the results of this analysis reveal in the overall average of the analysed ten-year period the following: at a starting altitude (height) of 1 m above the ground, an *E. multilocularis* egg is carried approximately 17 m by the wind. From a starting point with a height of 90 cm, this distance is roughly halved. On less dense forest areas (with a lower roughness length),

Table 1. Flight length of an *Echinococcus multilocularis* egg in relation to the mean wind speed of a whole year according to the ten-year average and in relation to the mean wind speeds in seasons according to the ten-year average.

	Forest area (dense):			Forest area (less dense):		
	reference values	extrapolated values		extrapolated values		
roughness length (m):	n/a	0.8		0.2		
height (m):	1.0	0.9	0.8	0.7	0.5	0.3
<i>overall:</i>						
wind (m/s):	0.704	0.372	0	0.548	0.401	0.177
flight length (m):	17.22	8.18	0	9.38	4.90	1.30
<i>spring:</i>						
wind (m/s):	0.674	0.355	0	0.524	0.383	0.170
flight length (m):	16.47	7.82	0	8.97	4.69	1.25
<i>summer:</i>						
wind (m/s):	0.644	0.340	0	0.501	0.367	0.162
flight length (m):	15.74	7.48	0	8.58	4.48	1.19
<i>autumn:</i>						
wind (m/s):	0.731	0.386	0	0.569	0.416	0.184
flight length (m):	17.86	8.49	0	9.73	5.08	1.35
<i>winter:</i>						
wind (m/s):	0.769	0.406	0	0.598	0.438	0.194
flight length (m):	18.80	8.93	0	10.24	5.35	1.42

The values given in boldface represent the final calculated flight distances (output), while values in regular type represent parameters implemented in the flight model/equations (input).

the egg can still fly more than 9 m from a starting point with a height of 70 cm. As soon as the start takes place only 50 cm above the ground, the *E. multilocularis* egg manages to cover a distance of almost 5 m. At a starting altitude of 30 cm, the wind still launches the egg, but just a little more than 1 m from its original starting point it will reach the ground.

Flight distances – seasons (spring, summer, autumn and winter)

It is noticeable when comparing the four seasons (table 1), in both cases, concerning the dense and the less dense forest areas, deviations regarding the flight distances in the overall annual mean can be observed. The flight range of the egg tends to decrease in spring. This declining tendency is found reinforced in summer, where also the lowest values occurred. In autumn, the flight length is slightly higher than the annual average. Here, the egg is transported about 60 cm further at a starting height of 1 m, compared to the annual average. The longest flight distances emerged in winter. At a height of 1 m, the trajectory was approximately 1.5 m longer than the annual standard value. Between summer and winter seasons, the achieved flight length differs by approximately 3 m.

Discussion

As outlined in the introduction, the possible distribution paths of the *E. multilocularis* eggs are generally uncertain, of highly complex pattern and showing knowledge gaps (Lawson & Gemmill, 1983; Eckert & Thompson, 2017). At least since 1981 the impact of wind as one possible vector in the infection process of *E. multilocularis* is discussed (World Health Organization, 1982). Over time, this aspect and the research on it faded into the background.

In this study, the rate of descent of *E. multilocularis* eggs was calculated to verify whether an egg is physically able to be transported by wind. We found the eggs rate of descent is similar to hornbeam pollen (*Carpinus betulus*) (Rempe, 1937). In the context of this work, the theoretical flight ability of *E. multilocularis* eggs can be regarded as confirmed. In addition, a mathematical model was developed to calculate possible flight distances of the egg in forest areas depending on starting altitude and wind speed. In dependence of its (examined) starting altitudes, the flight lengths varied between approximately 8 m (height: 0.9 m) and approximately 17 m (height: 1 m) in dense forest areas and between 1.3 m (height: 0.3 m) and 9.38 m (height: 0.7 m) in less dense forest areas. While in the spring and summer seasons, the reached distances slightly declined, in winter and autumn they exceeded the overall average. Therewith, wind-related spreading of eggs is possible.

To the knowledge of the authors, these results are the first of their kind. These findings, being in line with those of uttering of Guo *et al.* (2017), confirm the assumptions of the World Health Organization from 1981 that wind is a possible vector of *E. multilocularis* eggs (World Health Organization, 1982). They differ from the voices of those who argued against the involvement of wind as a possible distributor of *E. multilocularis* eggs (Lawson & Gemmill, 1983). Furthermore, this work not only confirms the transport by wind, but also, by introducing a simple mathematical model with high practical applicability on a wide range of landscapes, helps to estimate flight lengths and thereby the impact of wind as an epidemiological vector (in this study done for forest landscapes). The impact of different starting

altitudes, landscape situations and climatic situations (seasons) could be shown. Compared to previously observed dispersals of *Echinococcus* and *Taenia* eggs, in which wind may have been involved, our calculated flight distances are settled below the distances from the deposition site found in real endemic scenarios (e.g. 115 m (Sánchez Thevenet *et al.*, 2019) and 180 m (Bourée, 2001)). This could be explained by the different landscape types and the associated higher wind speeds, as well as the simultaneous influence of biotic vectors (e.g. beetles, flies and birds).

The significance of potential flight ranges, of course, are difficult to assess, regarding the risk to humans without determining the location of the starting point (e.g. distances to hiking trails, tourist attractions, etc.). Nevertheless, airborne spreading of *E. multilocularis* eggs, especially in fox-inhabited areas, can therefore be assumed as a considerable risk to hikers and berry pickers. The shorter flight distances, which were determined for spring and summer, can be seen as a positive factor reducing the potential risk of infection due to higher temperatures and aridity, which substantially reduce the viability of the eggs. Anyhow, the plentiful presence of fruits and berries, especially since these vector plants are often located on shady and cooler areas which could increase the survival time, can buffer the shorter flight lengths by serving as potential infection sources. While in the autumn and winter seasons flight distances raise due to higher wind-speeds. Increased humidity and lower temperatures enhance the durability of the eggs. The resulting risk of infection appears to be notably increased. A mitigating factor in relation to the hand–mouth infection during these months is the extensive lack of fruits (berries) – except for mushrooms in autumn.

Anyhow, some limitations of this model are worth noting. Although the flight lengths are an exciting first glimpse of the impact of wind, a more complex model should be developed in which non-linear flight paths, multiple restarts, vertical and lateral wind movements, respectively, and updraughts are considered. The model presumes that the flight is not disturbed. Under real conditions, this might not always be the case. Obstacles, such as bushes, shrubs, ground vegetation or collisions with trees or animals may interrupt the flight. Nevertheless, eggs that got stuck in this way can be available for further (airborne) transport. Furthermore, the flight model neglects mechanical and thermal turbulence which could abbreviate the flight or catapult the eggs to higher altitudes with higher wind speeds and hence extend the flight of the *E. multilocularis* egg – similarly to observation with pollen (Miki *et al.*, 2022). In the model the flight of a single egg is simulated. Under real conditions, however, it can be assumed that a group of eggs will start in the majority of flight events. Just as pollen generally do not fly solitary, but rather move in cloud structures (Hattemer *et al.*, 2001), this could also be assumed for *E. multilocularis* eggs, concomitant with a raised potential of contamination and infectivity.

The demonstrated model can be applied to a wide range of simulations. Consequently, there calculations should be conducted for other forest stands such as managed needle stands, deciduous stands, mixed stands and further terrains such as fields, meadows and urban areas, where much higher wind speeds are expected.

For further verification it would be of utmost interest to put such calculations in context to other real data such as the prevalence of infected foxes, rodents or humans in a chosen region under continuous investigation. Additionally, the question whether *E. multilocularis* eggs lose or retain their contagiousness after transport by wind remains open and requires further research.

Despite the limitations shown, the model provides clear information. The resulted flight lengths, which vary greatly depending on the population structure and starting altitude, underline the potential epidemiological importance of wind with regard to the spread of *E. multilocularis* eggs. It is hoped that these findings will bring ‘new life’ in the research about wind as an impact factor on the infection paths of *E. multilocularis*. Future work should consolidate the initial gained knowledge by conducting a field experiment with accurate surrogate eggs.

Acknowledgements. We thank Andreas Dillmann from the German Aerospace Institute in Göttingen for his support in developing a flight length model and Heinrich Kreilein from the Bioclimatology Department of the Faculty of Forest Sciences and Forest Ecology in Göttingen, who made the wind data accessible. Furthermore, we thank the anonymous reviewers for their careful reading of our manuscript and their insightful comments and suggestions.

Author contributions. All of the authors made significant contributions to the concept, design, execution and interpretation of the research and the development of the outline. S.N. proposed and conducted the research and supervised the first draft. S.S. discussed and shaped the research and improved upon the manuscript.

Financial support. The authors declare that there was no financial support for this paper.

Conflict of interest. None.

Ethical approval. None.

References

- American Meteorological Society (2019) *Aerodynamic roughness length secondary aerodynamic roughness length*. Available at https://glossary.amet-soc.org/w/index.php?title=Aerodynamic_roughness_length&oldid=1090 (accessed 17 February 2022).
- Atkinson J-AM, Williams GM, Yakob L, Clements ACA, Barnes TS, McManus DP, Yang YR and Gray DJ (2013) Synthesising 30 years of mathematical modelling of echinococcus transmission. *PLoS Neglected Tropical Diseases* 7(8), e2386.
- Avison J (1989) *The world of physics*. 2nd edn. Cheltenham, Nelson.
- Bird RB, Stewart WE and Lightfoot EN (2007) *Transport phenomena*. 2nd edn. New York/Chichester/Weinheim/Brisbane/Singapore/Toronto, John Wiley & Sons, Inc.
- Bourée P (2001) Hydatidosis: dynamics of transmission. *World Journal of Surgery* 25(1), 4–9.
- Cavallini P (1995) Variation in the body size of the red fox. *Annales Zoologici Fennici* 32(4), 421–427.
- Conraths FJ and Deplazes P (2015) *Echinococcus multilocularis*: epidemiology, surveillance and state-of-the-art diagnostics from a veterinary public health perspective. *Veterinary Parasitology* 213(3–4), 149–161.
- David ED and Lindquist WD (1982) Determination of the specific gravity of certain helminth eggs using sucrose density gradient centrifugation. *The Journal of Parasitology* 68(5), 916–919.
- Deplazes P and Eckert J (2001) Veterinary aspects of alveolar echinococcosis – a zoonosis of public health significance. *Veterinary Parasitology* 98(1–3), 65–87.
- Deplazes P, Rinaldi L, Alvarez Rojas CA, et al. (2017) Chapter Six – global distribution of alveolar and cystic echinococcosis. *Advances in Parasitology* 95, 315–493.
- Eckert J and Thompson RCA (2017) Chapter One - historical aspects of echinococcosis. *Advances in Parasitology* 95, 1–64.
- Eckert J, Deplazes P and Kern P (2011) Alveolar echinococcosis (*Echinococcus multilocularis*): and neotropical forms of echinococcosis (*Echinococcus vogeli* and *Echinococcus oligarthrus*). pp. 669–699 in Palmer SR, Soulsby L, Torgerson P and Brown DWG (Eds) *Oxford textbook of zoonoses: Biology, clinical practice, and public health control*. Oxford, Oxford University Press.
- Fischer C, Reperant LA, Weber JM, Hegglin D and Deplazes P (2005) *Echinococcus multilocularis* infections of rural, residential and urban foxes (*Vulpes vulpes*) in the canton of Geneva, Switzerland. *Parasite* 12(4), 339–346.
- Food and Agriculture Organization of the United Nations & World Health Organization (2014) Multicriteria-based ranking for risk management of food-borne parasites. pp. 1–302. *Microbiological risk assessment series*. Rome, Food Safety and Codex Unit, FAO of United Nations.
- Gemmell MA, Johnstone PD and Boswell CC (1978) Factors regulating tapeworm populations: dispersion patterns of *Taenia hydatigena* eggs on pasture. *Research in Veterinary Science* 24(3), 334–338.
- Goszczyński J (1990) Scent marking by red foxes in central Poland during the winter season. *Acta Theriologica* 35(1), 7–16.
- Gualtieri G and Secci S (2011) Comparing methods to calculate atmospheric stability-dependent wind speed profiles: a case study on coastal location. *Renewable Energy* 36(8), 2189–2204.
- Guo Y, Xiao Y, Wang H, Liu W and Tang Y (2017) Helminthiasis.. pp. 83–193 in Li H (Ed) *Radiology of parasitic diseases: A practical approach*. Dordrecht, Springer.
- Hassan AS and Munganga JMW (2019) Mathematical global dynamics and control strategies on *Echinococcus multilocularis* infection. *Computational and Mathematical Methods in Medicine* 2019, 3569528.
- Hattemer HH, Ziehe M, Finkeldey R and Fromm M (2001) Genetic diversity and differentiation of individual effective pollen clouds in trees.. pp. 223–237 in Müller-Starck G and Schubert R (Eds) *Genetic response of forest systems to changing environmental conditions*. Dordrecht, Springer.
- Hegglin D, Bontadina F and Deplazes P (2015) Human–wildlife interactions and zoonotic transmission of *Echinococcus multilocularis*. *Trends in Parasitology* 31(5), 167–173.
- Ishikawa H, Ohga Y and Doi R (2003) A model for the transmission of *Echinococcus multilocularis* in Hokkaido, Japan. *Parasitology Research* 91(6), 444–451.
- Ito A, Nakao M and Sako Y (2007) Echinococcosis: serological detection of patients and molecular identification of parasites. *Future Microbiology* 2(4), 439–449.
- Kapel CMO, Torgerson PR, Thompson RCA and Deplazes P (2006) Reproductive potential of *Echinococcus multilocularis* in experimentally infected foxes, dogs, raccoon dogs and cats. *International Journal for Parasitology* 36(1), 79–86.
- Khan A, Ahmed H, Sohail A, Alam F and Simsek S (2020) A mathematical modelling approach for treatment and control of *Echinococcus multilocularis*. *Parasitology* 147(3), 376–381.
- Lawson JR and Gemmell MA (1983) Hydatidosis and cysticercosis: the dynamics of transmission. *Advances in Parasitology* 22, 261–308.
- Lewis FI, Otero-Abad B, Hegglin D, Deplazes P and Torgerson PR (2014) Dynamics of the force of infection: insights from *Echinococcus multilocularis* infection in foxes. *PLoS Neglected Tropical Diseases* 8(3), e2731.
- Meesenburg H, Meiwes K-J and Bartens H (1999) Veränderung der Elementvorräte im Boden von Buchen- und Fichtenökosystemen im Solling [Changes in element stocks in the soil of beech and spruce ecosystems in Solling]. *Berichte Freiburger Forstliche Forschung* 7, 109–114. [In German.]
- Merck (2018) *The merck manual of diagnosis and therapy*. 20th edn. Kenilworth, NJ, Merck Sharp & Dohme Corp., A Subsidiary of Merck & Co., Inc.
- Miki K, Kawashima S, Kobayashi S, Takeuchi S, Tseng Y-T and Nakamura K (2022) Evaluating inaccurate pollen concentrations caused by turbulence using passive sampler. *Aerobiologia* 38, 1–12.
- Paar U, Evers J and Eichhorn J (2013) Ergebnisse der Waldökosystemforschung im Solling [Results of forest ecosystem research in Solling]. pp. 1–138 in Paar U, Evers J and Eichhorn J (Eds) *Jahrestagung der AFSV vom 11. – 14. September 2013 im Solling (NW-FVA)* [Annual meeting of the AFSV from September 11th to 14th, 2013 in Solling (NW-FVA)]. Solling, Nordwestdeutsche Forstliche Versuchsanstalt (NW-FVA).
- Pertmann I (2011) Estonian wind farms’ need for full balance power. *Oil Shale* 28, 193–202.
- Rempe H (1937) Untersuchungen über die Verbreitung des Blütenstaubes durch die Luftströmungen [Studies on the distribution of pollen by air currents]. *Planta* 27(1), 93–147. [In German.]
- Romig T, Deplazes P, Jenkins D, Giraudoux P, Massolo A, Craig PS, Wassermann M, Takahashi K and De La Rue M (2017) Chapter five -

- ecology and life cycle patterns of *Echinococcus* species. *Advances in Parasitology* **95**, 213–314.
- Sánchez Thevenet P, Alvarez HM, Torrecillas C, Jensen O and Basualdo JA** (2019) Dispersion of *Echinococcus granulosus* eggs from infected dogs under natural conditions in Patagonia, Argentina. *Journal of Helminthology* **94**(1), 1–7.
- Thompson RCA** (1986) Biology and systematics of *Echinococcus*. pp. 5–43 in Thompson RCA (Ed) *The biology of echinococcus and hydatid disease*. London, George Allen & Unwin.
- Tolnai Z, Széll Z and Sréter T** (2013) Environmental determinants of the spatial distribution of *Echinococcus multilocularis* in Hungary. *Veterinary Parasitology* **198**(3–4), 292–297.
- Torgerson PR, Schweiger A, Deplazes P, Pohar M, Reichen J, Ammann RW, Tarr PE, Halkik N and Müllhaupt B** (2008) Alveolar echinococcosis: from a deadly disease to a well-controlled infection. Relative survival and economic analysis in Switzerland over the last 35 years. *Journal of Hepatology* **49**(1), 72–77.
- Torgerson PR, Keller K, Magnotta M and Ragland N** (2010) The global burden of alveolar echinococcosis. *PLoS Neglected Tropical Diseases* **4**(6), e722.
- Veit P, Bilger B, Schad V, Schäfer J, Frank W and Lucius R** (1995) Influence of environmental factors on the infectivity of *Echinococcus multilocularis* eggs. *Parasitology* **110**(1), 79–86.
- Weisstein EW** (2002) *CRC concise encyclopedia of mathematics*. 2nd edn. New York, Chapman and Hall/CRC.
- Woolsey ID and Miller AL** (2021) *Echinococcus granulosus sensu lato* and *Echinococcus multilocularis*: a review. *Research in Veterinary Science* **135**, 517–522.
- World Health Organization** (1982) *Report on FAO/UNEP/WHO meeting on research requirements in echinococcosis/hydatidosis and taeniasis/cysticercosis*. Geneva: World Health Organization. 29 April–1 May 1981.